

- Announcements, Review HW
- Lab Quiz
- Pre-lab Lecture
 - ❖ Where we are/going
 - ❖ DNA Ligation, part 2
 - ❖ Bacterial Transformation
 - ❖ Controls, Expected Outcomes
 - ❖ Safety + Technical Tips

Old HW, problem 2

Enzyme activity measure in arbitrary units U

1 U = amount of enzyme to digest 1 μ g DNA
within 1 hr. at 37°C in total of 50 μ l

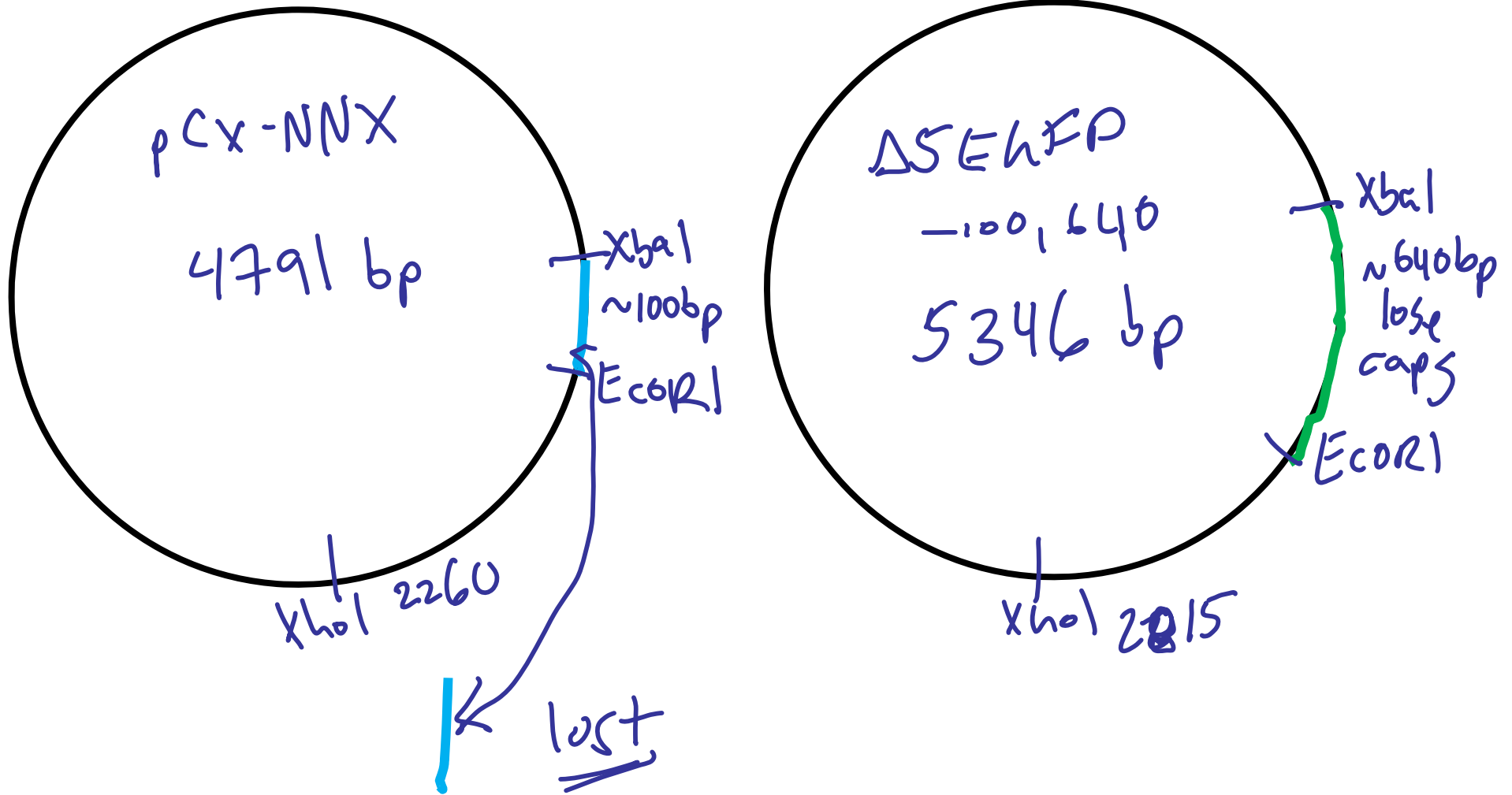
Will 75% XbaI and 100% EcoRI activity always give a
3:1 ratio of double-cut to single-cut product?

extreme cases

v. little DNA (<<1 μ g) ~ all double-cut

a lot DNA not even all single-cut

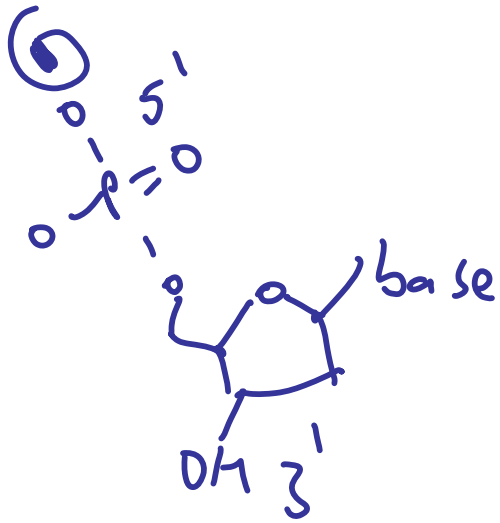
Old HW, problem 1



Where we are/going

- 🚩 make the desired clone (ligation)
- 🚩 amplify and select the clone in *E. coli*
- 🚩 next time: test candidate clones

DNA Ligation



Reaction creates new phosphodiester bond

Reaction requires ATP

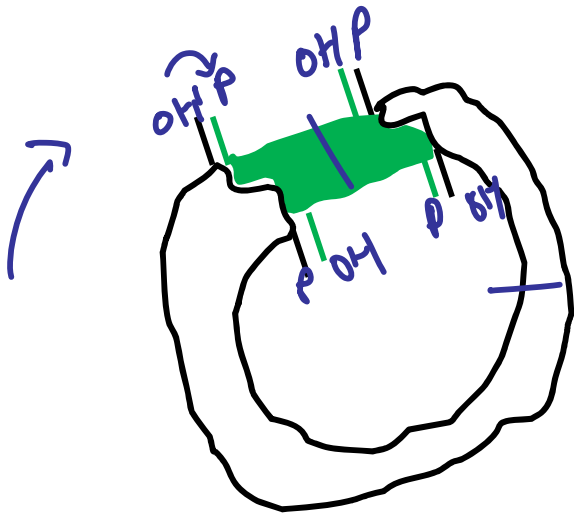
What factors affect yield?

- conc. of DNA
- ratio of α : β : γ
- enzyme (conc., T, age)
- time for rxn.

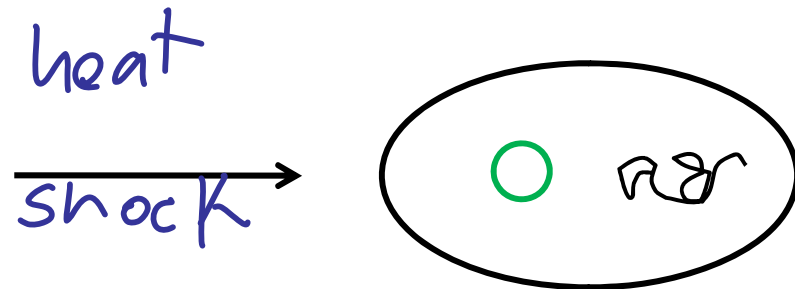
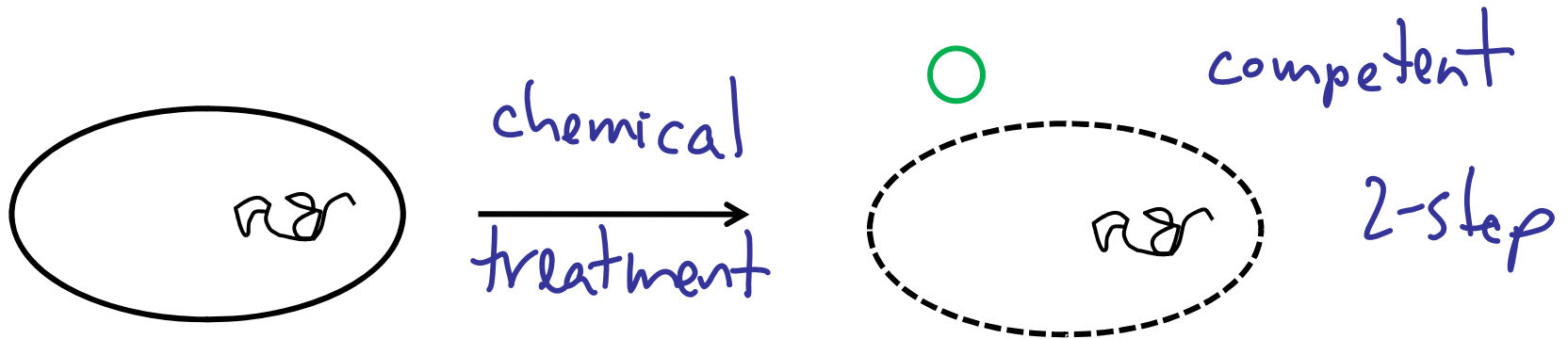
How do we assess if it worked?

* diagnostic digest

(sequencing)



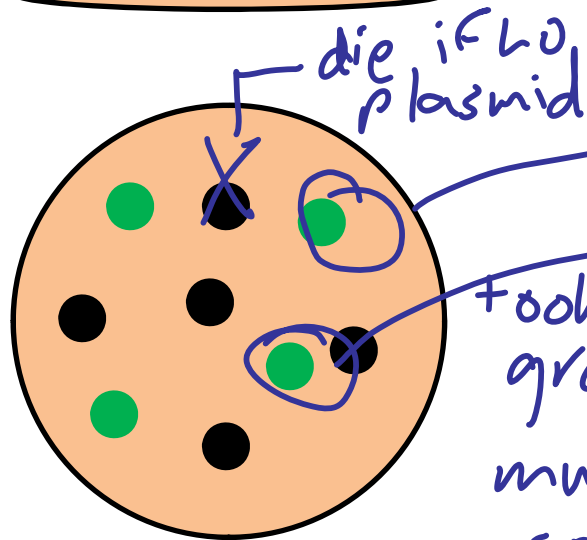
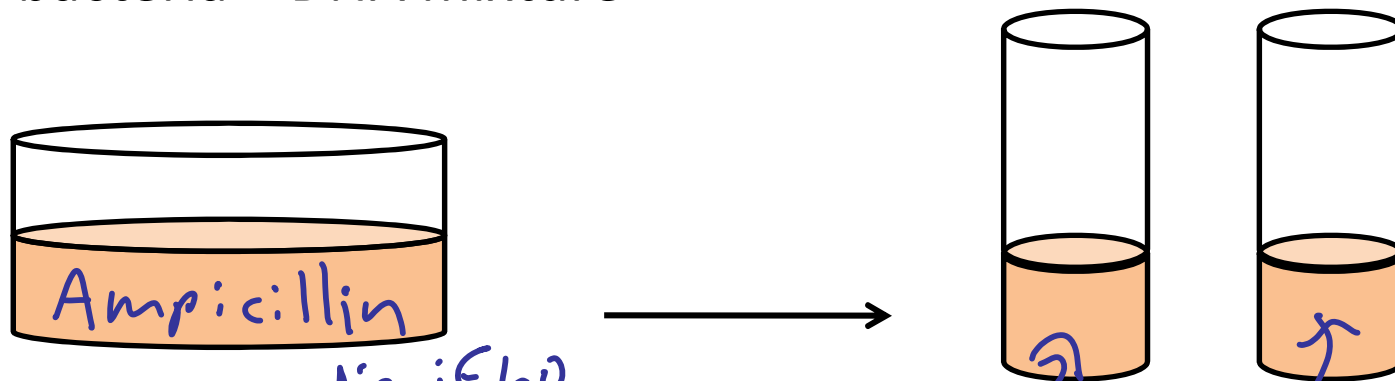
Bacterial transformation



other methods (1-step)
- electroporation
- ballistics

DNA Amplification in Bacteria

Plate bacteria + DNA mixture



die if no plasmid

Why grow/test multiple candidates?

took up plasmid,
grow into
multi-cell
colonies

not all
are correct

Ligation Controls + Outcomes

Sample	Expectation	Role
pCX-EGFP	lots (of colonies)	positive control
no DNA	none	neg. control
bkb + ins, no ligase	some	control for uncut plasmid
bkb + ligase	some (present)	control for single-cut plasmid
bkb + ins, + ligase	Some - many	experiment

} digestion efficiency

Ligation Controls + Outcomes

Sample	What if?
pCX-EGFP	none <i>wrong DNA bad cells bad plate</i>
no DNA	some <i>wrong cell line</i>
bkb + ins, no ligase	many <i>wrong DNA</i>
bkb + ligase	many <i>bad digestion (enzymes bad)</i>
bkb + ins, + ligase	none <i>exp. stuff or new plasmid kills bacteria</i>

In general, keep in mind:

* consider all outcomes together

* rxns. do not go to completion

(not in our case)

Today in Lab

- Keep ligase *and* ligase buffer (ATP) cold
- DNA precipitation after ligation reaction
 - Yeast tRNA → "carrier" to visualize
 - Ethanol → precipitates DNA
- Be gentle with competent cells *Keep cold, don't vortex*
- Sterile technique for transformations – demo