

- Announcements
- Lab Quiz
- Pre-lab Lecture
  - ❖ Library screen
  - ❖ Engineering tools/analogues
  - ❖ Today in Lab (Mod 2 Day 3)

# Announcements

- Brief discussion of previous quiz
- Notebooks due at end of lab today
- Next time
  - come to *lab* first to set up experiment
  - no quiz!
  - we'll go together to 16-336 for j. club talks
- J. club grading rubric available up front
- Error: Kan<sup>R</sup> is for the EnvZ deletion, not the integration (see strain description)

\* Slides  
due  
@ 12am

# Measuring LacZ: original system

Team Color	B-gal units (dark)	B-gal units (light)
Green	358	173
Blue	1644	531
Pink	616	1319
Purple	1334	475

$$1 \text{ Miller Unit} = 1000 * \frac{(Abs_{420} - (1.75 * Abs_{550}^{OD}))}{(t * v * Abs_{600}^{OD})}$$

Slide modified from N. Kuldell

# Genetic library screen: concepts

- Goal: improve contrast of bact. photographs
- Specifically, make plates darker in the dark
- Which mutations should have this effect?
- Why randomize at or near those sites?
- Control/reference mutation: *kinase dead, i-11*  
*red: k<sup>+</sup>p<sup>-</sup>*

F11:	EnvZ		<b>A239T</b>	<b>G240E</b>	<b>V241G</b>	<b>S242D</b>	<b>H243A</b>		<b>blue = K-P+</b>
F12:	<u>EnvZ</u>	<b>H243</b>	D244	L245	R246	<b>T247R (K+P-)</b>	P248		
	wt seq	CAC	GAC	TTG	CGC	<b>Thr = ACG</b>	CCG		
	<u>Cph8</u>	<b>H537</b>	D538	L539	R540	<b>T541</b>	P542		
	to test:	<b>Kinase Dead mutant</b>				<b>NNY mutagenesis</b>			

*H537A*

Images from F11 and F12 wikis

# Genetic library screen: methods

- Electroporation of library into cells
  - modified bacterial photography strain:  $\Delta$ Cph8
  - library of T247 randomized to NNY *no stop codons*  
*541*

- Selective media:

## MacConkey MUG agar

- contains lactose and red pH/metabolic indicator
- high [B-gal] leads to more red colonies (cleavage)

*as in redder, not #*



Image from wiki (M2D4 "Talk" page)

# Registry of Std Bio Parts

levels of abstraction

Goal: design protein generator that functions in *E. coli*

## Browse parts and devices by function

This section replaces the previous *Featured parts* pages.

### Browse parts by type

Catalog

List



**Promoters (?)**: A promoter is of the downstream DNA sequence.



**Ribosome Binding Sites (?)**: can bind and initiate translation.



**Protein domains (?)**: Protein up a protein coding sequence target the protein for cleavage.



**Protein coding sequences**: Note that some protein coding protein from start codon to stop also included here.



**Translational units (?)**: They begin at the site of translation.



**Biosynthesis**: Parts involved in the production of biological molecules.



**Cell-cell signaling and quorum sensing**: Parts involved in communication between cells.



**Cell death**: Parts involved in killing cells.



**Coliroid**: Parts involved in taking a bacterial cell.

### Browse parts and devices by standard

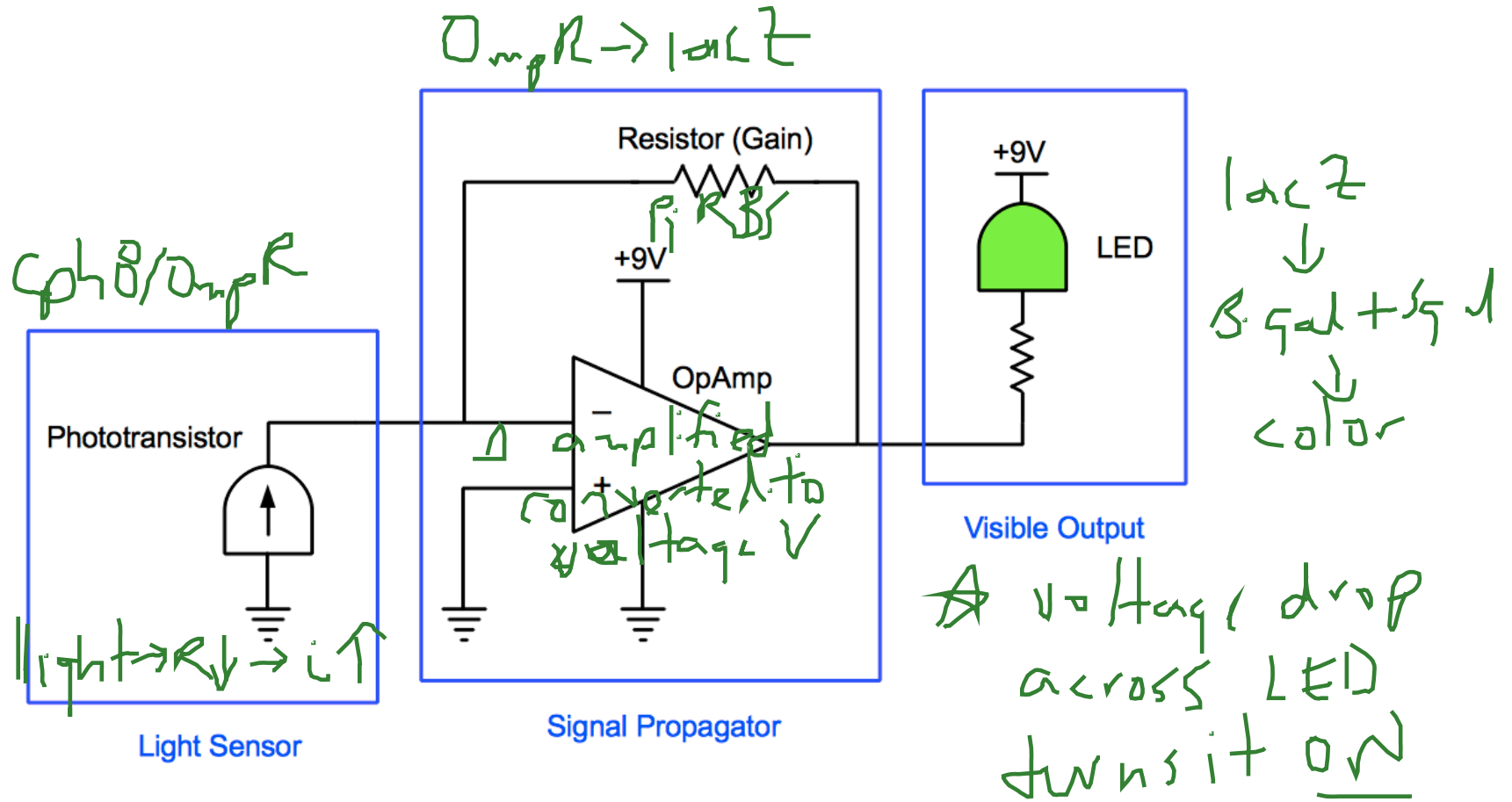
Unless otherwise specified, most parts in the Registry comply with this assembly standard.

**Assembly standard 10 (?)**: Assembly standard 10, parts that comply with this assembly standard.

**Assembly standard 23 (?)**: Assembly standard 23, parts that comply with this assembly standard.

Images from parts.mit.edu

# EE analogue to BP system

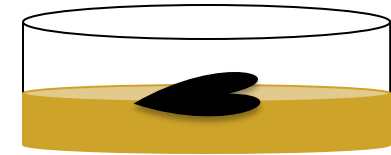


Goal: vary R and observe outcomes

limit cases:  $R=0, \infty$

# Today in Lab: M2D3

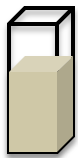
Observe/document *coli*roid from last time



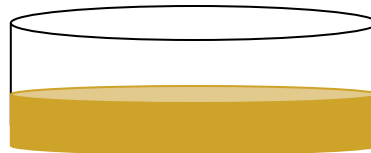
Set up library screen

1. electroporate

2. plate cells (BPΔCph8)



incubate 1 hr\*

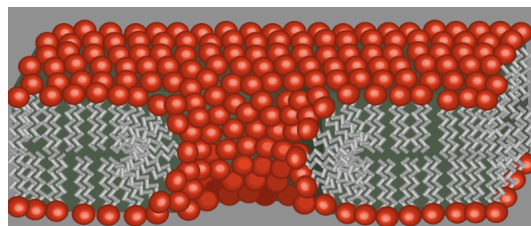
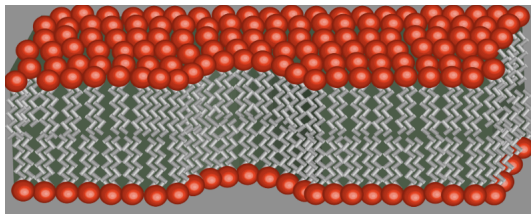


Explore EE analogue

Explore parts.mit.edu

Explore TinkerCell

\*transfer to SOC medium  
(has extra sugar, etc.)



Membrane model from Wikimedia Commons, public domain image