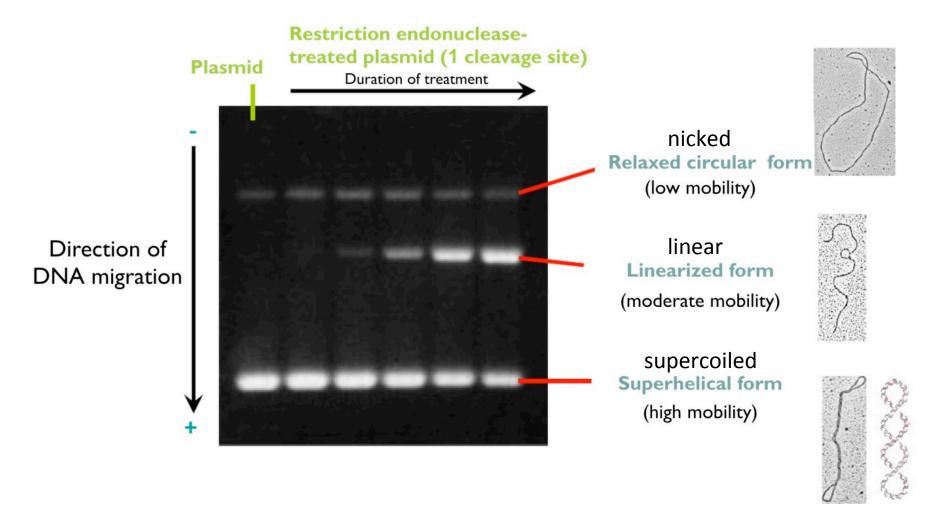
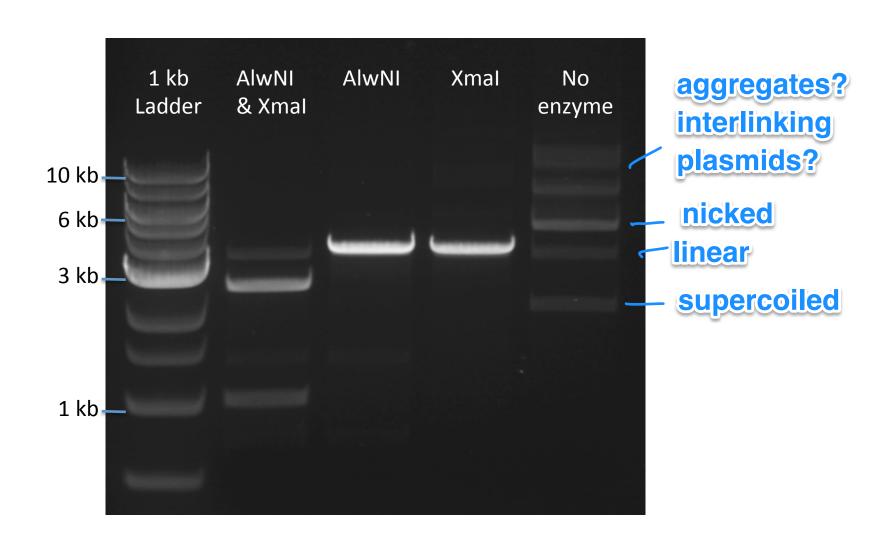
M1D3: Purify protein for secondary assays

- 1. Pre-lab discussion part 1
- 2. Purify FKBP12 protein
- 3. During 1 hour incubation (2pm):
 - Pre-lab discussion part 2
 - Select ligands (a.k.a. compounds) from identified SMM hits
- 4. Continue with FKBP12 purification & ligand selection

Deciphering confirmatoin digest results



Deciphering confirmatoin digest results



Other potential explanations

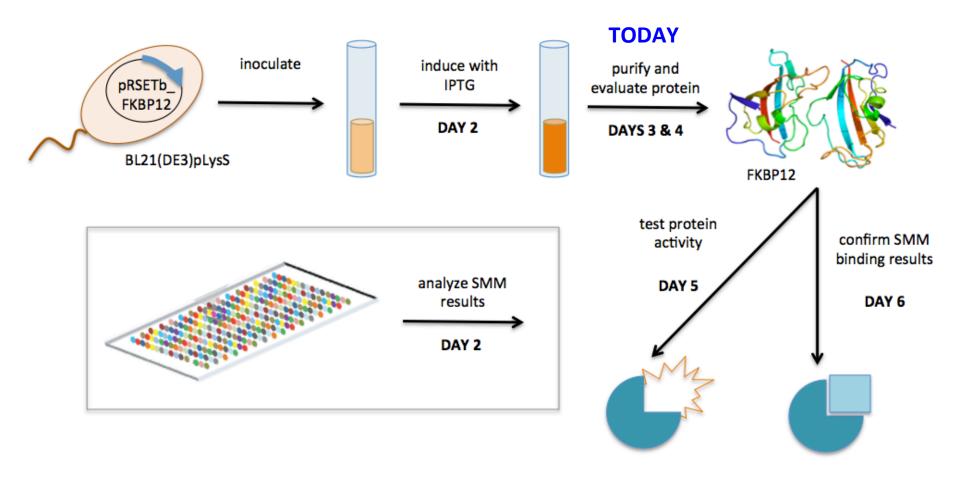


Home > Tools & Resources > Troubleshooting Guides > Restriction Enzyme Troubleshooting Guide

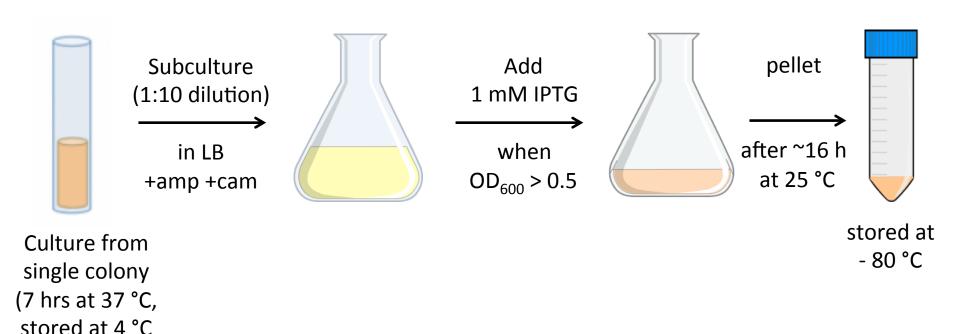
Restriction Enzyme Troubleshooting Guide

- Smear?
- Incomplete and / or no cutting?
- Extra bands?

Overview of Mod1 experiments



How did we induce protein expression?



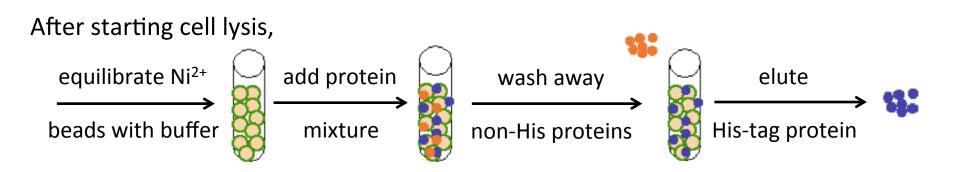
In addition to your induced sample, you will also examine and un-induced sample for FKBP12 expression

overnight)

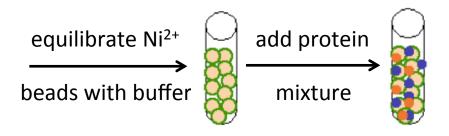
How will we retrieve our protein?

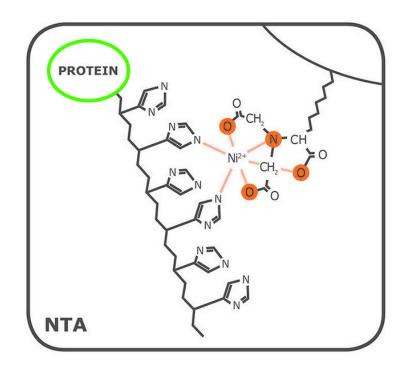
Cell lysis buffer components:

- protease inhibitor (AEBSF)
 eliminates proteases
- deoxyribonuclease (DNase)
 degrades DNA
- tris / salts buffer maintain pH & osmotic pressure
- dithiothreitol (DTT) reducing agent
- glycerol stabilizer
- lysozyme degrades bacterial cell wall



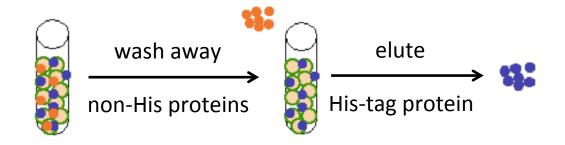
6x His residues enable binding to Ni²⁺





 Ni²⁺ chelated onto agarose resin via nitrilotriacetic acid (NTA) ligand

Wash and elute protein: Imidazole competes for binding to Ni²⁺



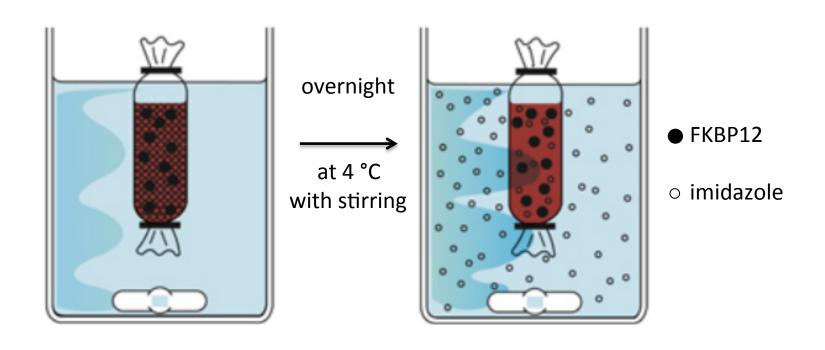
histidine

pKa ~7 — N N H

- Low concentration of imidazole included in wash buffer
- Increased 25-fold in elution buffer

Dialyze to remove imidazole

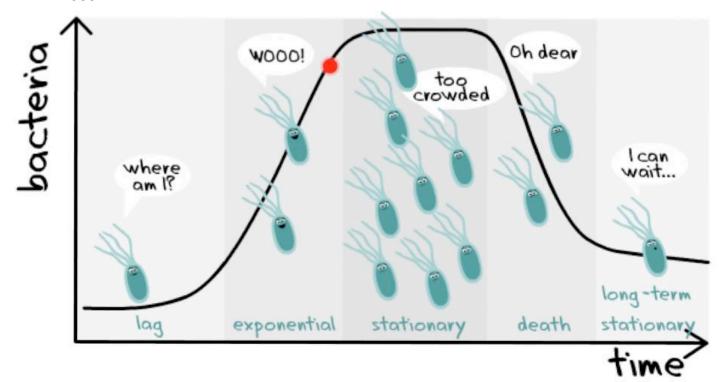
- Dialyze ('dilute out') imidazole with semi-permeable membrane of cross-linked polymers
- Molecular weight cutoff = 2000 Da
 - FKBP12-6His ~15 kDa, imidazole ~68 Da



Start lysis now

Why do we induce at $OD_{600} = 0.5-0.8$?

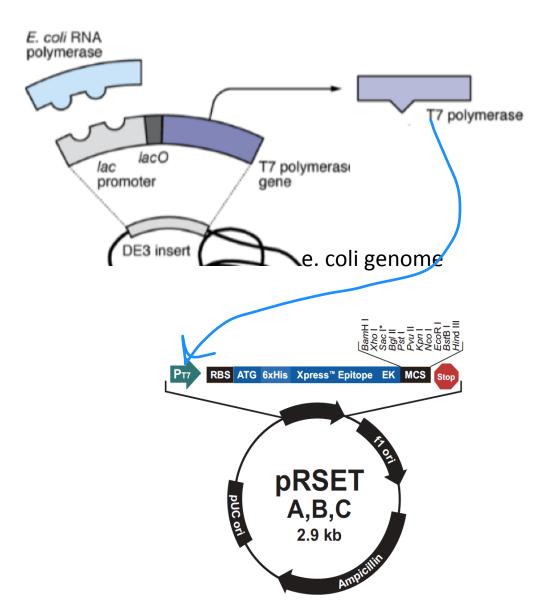
- Optical density (OD) is a measure of light scattering
 - E. coli scatter 600 nm light
- OD measures turbidity rather than absorption
 - Indicates # of cells present in culture
 - OD_{600} of 1 ≈ 8 x 10⁸ cells / mL



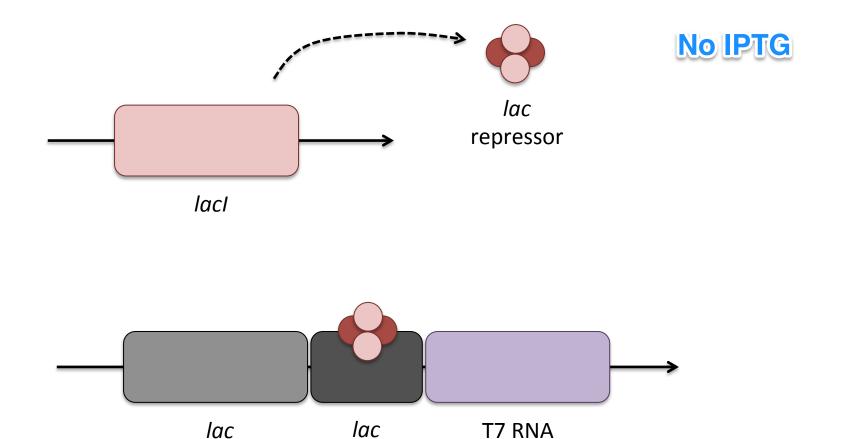
BL21(DE3)pLysS used for protein expression

e. coli

DE3



DE3: Lac promoter controls expression of T7 RNA polymerase

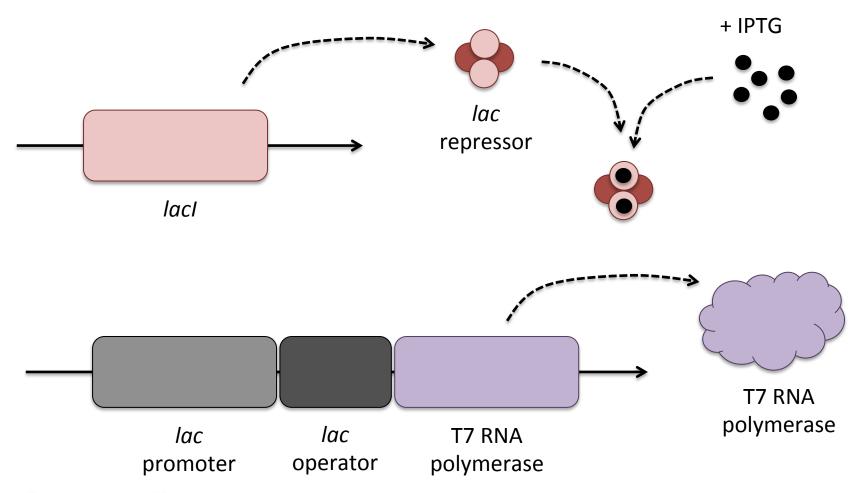


polymerase

operator

promoter

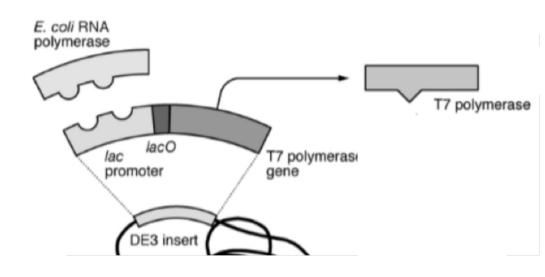
IPTG 'induces' protein expression



derepression

BL21(DE3)pLysS used for protein expression

• DE3

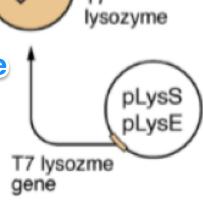


pLysS

T7 lysozyme binds T7 polymerase and degrades it

controls for leaky expression

maintained by chloramphenicol (cam)



IPTG induction of FKBP12 expression in words...

- In absence of IPTG:
 - LacI repressor binds *lac* operator;
 <u>represses/activates</u> transcription of T7 RNA polymerase
 - Leaky expression of T7 RNA polymerase corrected for by T7 lysozyme
- In presence of IPTG:
 - IPTG binds LacI; enables/prevents binding to lac operator
 - T7 RNA polymerase transcribed and binds at P_{T7}; initiates transcription of FKBP12

In lab today...

- Sign-up for compounds at the front laboratory bench
 - Each group will test two compounds

For next time...

- There will be a Comm Lab workshop in 56-614
- Draft schematic (image, title, caption)
- Outline introduction using topic sentences
 - Include reference information!!

Notes on topic sentences:

Used to introduce each paragraph

 Should 'funnel' from big picture topic to your specific research project

Specific background

Big picture motivation

 All claims should be supported by trusted sources

Your specific research question and hypothesis