M2D2: Perform protein purification protocol

1. Prelab discussion

2. Protein purification

3. Assess RE digest



"Don't pick it up," I say, and he says, "It's just a *plasmid*, what harm could it do?" Well just look at him now...who knows *what* protein he's expressing!

Homework

Crafting a slide for the Journal Article Presentation

Craft 1-2 slides using your journal article so you present key data from 1 figure

- Each slide should show a single message
- The title should state the take-home message of the data that are shown.
- Your slide(s) should show the data and highlight the key finding(s).
- The information should be clear and large enough to read.
- Keep text to a minimum. (NO figure captions on slide!)

EXAMPLE SLIDE: Football coaches are the highest paid \$1,057,305 academic employees at doctoral-granting universities

- Data represent expression of Y using method A
- Possibly something about the control(s), if applicable
- Important notes about the data (how were they generated?) and findings that are not already stated in the title
- Transition to next slide...
 (can also be done verbally)



Labwork

Purification of 6xHis-PfFKBP35

Overview of M2: drug discovery

Research goal: Test small molecules for binding to the *Plasmodium falciparum* FKBP35 protein using a functional assay.



PfFKBP35

- Pf = Plasmodium falciparum
- Essential to parasite survival
- Known drug target for:
 - Rapamycin
 - FK506
- Problem: Has a human ortholog in FKBP12
- How can we target the parasite protein and not the human?



Very Brief Outline of Induction

- 1) Addition of rhamnose causes Genomic Rhamnose Operon to make T7 RNA Polymerase
- 2) T7 RNA Polymerase binds T7 promoter and makes our protein



















Less Brief Outline of Induction

- 1) Rhamnose binds RharR protein that recruits RNA poly to the RharSR promoter **in the genome**
- 2) RharR & RharS get made
- 3) Rhamnose binds RharS that recruits RNA poly to the RhaBAD promoter
- 4) RhaBad promoter makes T7 RNA Poly
- 5) T7 RNA Polymerase binds T7 promoter **on our plasmid** and makes our protein

How do we induce protein expression?



Why do we add kanamycin to our culture? Why do we induce protein expression at $OD_{600} = 0.6$?

Addition of Rhamnose to induce protein expression occurs during the Exponential/log phase of growth



Component	Function	If it doesn't work?
BPER Bacterial Extraction Reagent		
Lysozyme		
DNAsel		
Proteinase Inhibitor Cocktail		



Component	Function	If it doesn't work?
BPER Bacterial Extraction Reagent	Detergent & Buffers break open the cell	
Lysozyme		
DNAsel		
Proteinase Inhibitor Cocktail		



Component	Function	If it doesn't work?
BPER Bacterial Extraction Reagent	Detergent & Buffers break open the cell	
Lysozyme	Chews through cell wall	
DNAsel		
Proteinase Inhibitor Cocktail		



Component	Function	If it doesn't work?
BPER Bacterial Extraction Reagent	Detergent & Buffers break open the cell	
Lysozyme	Chews through cell wall	
DNAsel	Chews up DNA	
Proteinase Inhibitor Cocktail		



Component	Function	If it doesn't work?
BPER Bacterial Extraction Reagent	Detergent & Buffers break open the cell	
Lysozyme	Chews through cell wall	
DNAsel	Chews up DNA	
Proteinase Inhibitor Cocktail	Inhibits Proteinases	



Component	Function	If it doesn't work?
BPER Bacterial Extraction Reagent	Detergent & Buffers break open the cell	No Lysis
Lysozyme	Chews through cell wall	No Lysis
DNAsel	Chews up DNA	Hard to purify (snotty)
Proteinase Inhibitor Cocktail	Inhibits Proteinases	No protein



6xHis tag binds to Ni²⁺ resin / column to allow purification of protein of interest via affinity purification





- Ni²⁺ chelated onto agarose resin via nitrilotriacetic acid (NTA) ligand
- His tag chelates to Ni²⁺ causing protein to 'stick' to resin / column

Non-specific binders washed from Ni²⁺ resin / column using a low concentration of imidazole



- Low concentration of imidazole included in wash buffer
- Imidazole competes for binding to Ni²⁺ resin
 - Low affinity binders / non-specific binders are outcompeted and released from the resin

High concentration of imidazole is used to elute the protein from the Ni²⁺ resin / column



- Similar concept to wash
 - Wash uses 50mM imidazole
 - Elution used 250mM imidazole
- Instead of competing away non-specific binding, we can now out-compete the His Tag

Purification process (and where you will save samples)



DNA electrophoresis review



How do you visualize the migration through the gel? Tracking dye / Dye front – Bromophenol blue How do you visualize DNA bands in the gel? SYBR Safe DNA Stain

For today...

- 1. Purify your protein for validation assay
- 2. During a centrifugation step, electrophorese your RE digest

For M2D3...

1. Answer question prompts on the wiki homework to think about how you will create a story from figures in the paper