MEMS based sensors for cellular studies

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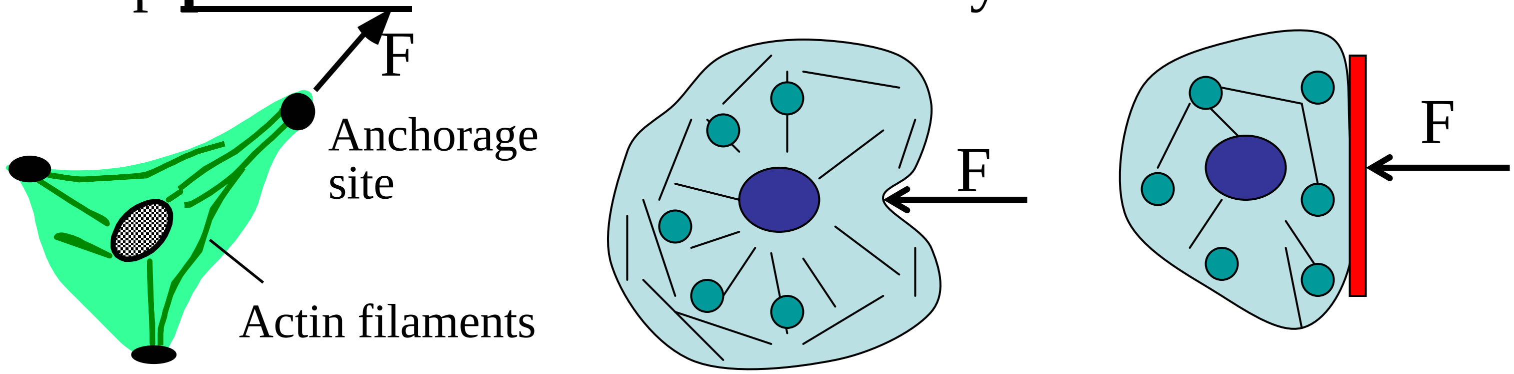
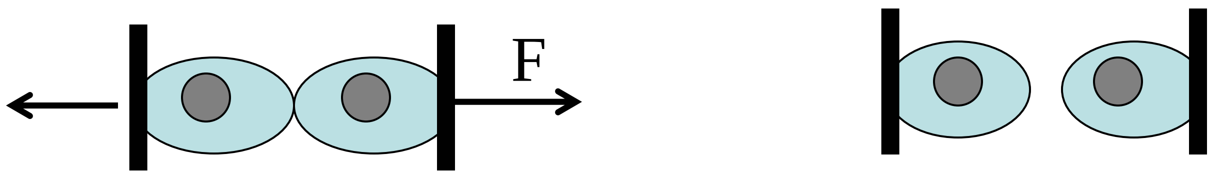
Part of GEM4 Summer School lectures on
instruments for cell mechanics studies (Aug 10, 2006, MIT)

Objective

Develop **portable** micro sensors to study:



• Cell mechanical response

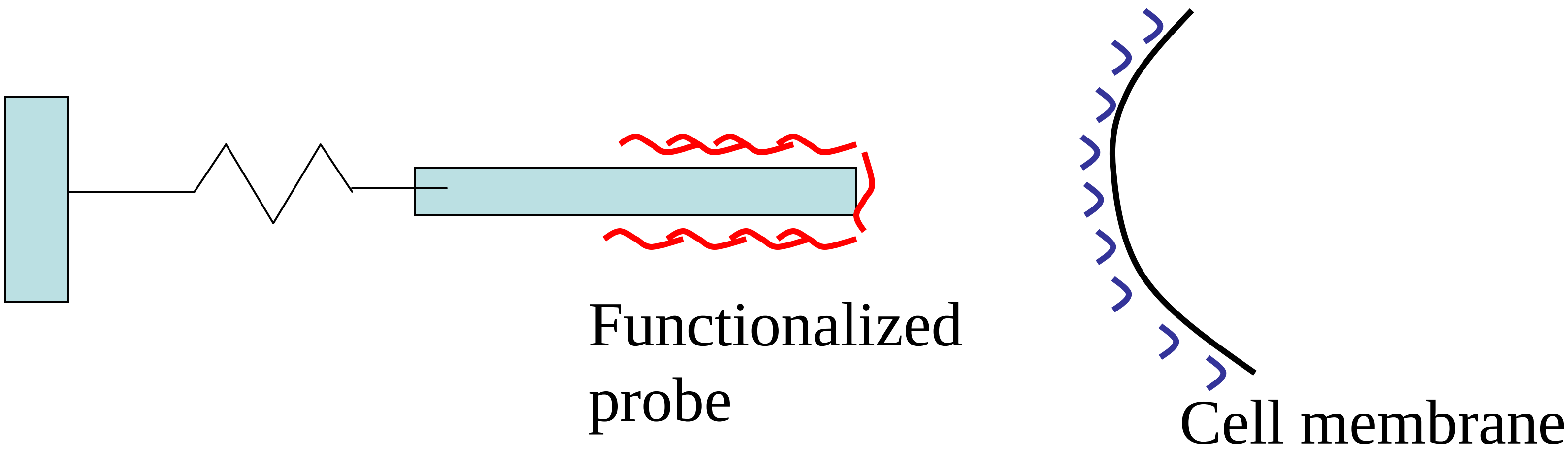


• Cell adhesion

in different biochemical environments
to explore mechanotransduction and disease detection

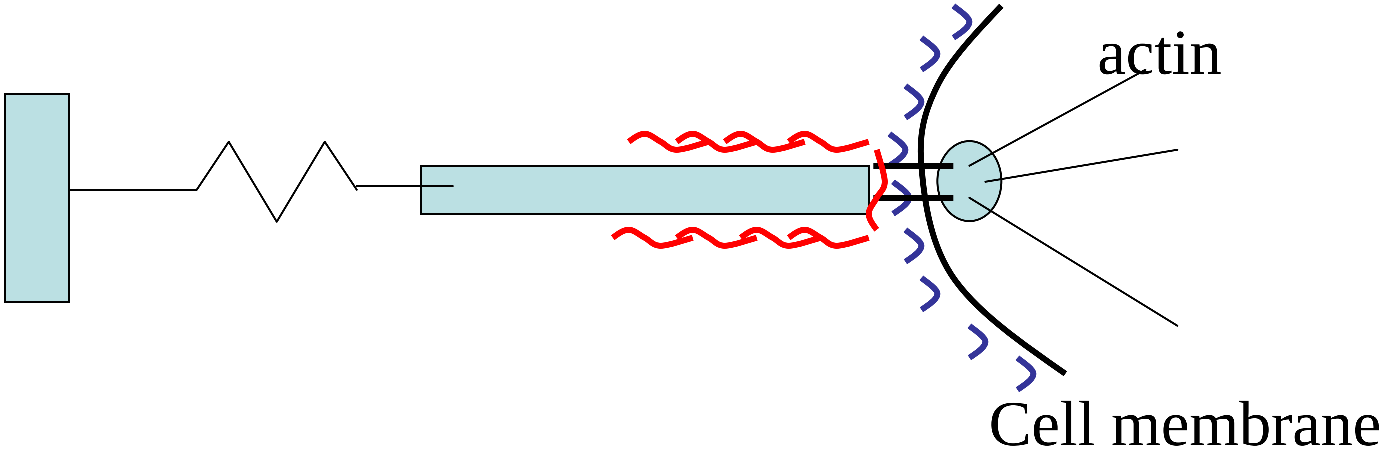
Basic idea

A micro spring is used to measure cell force



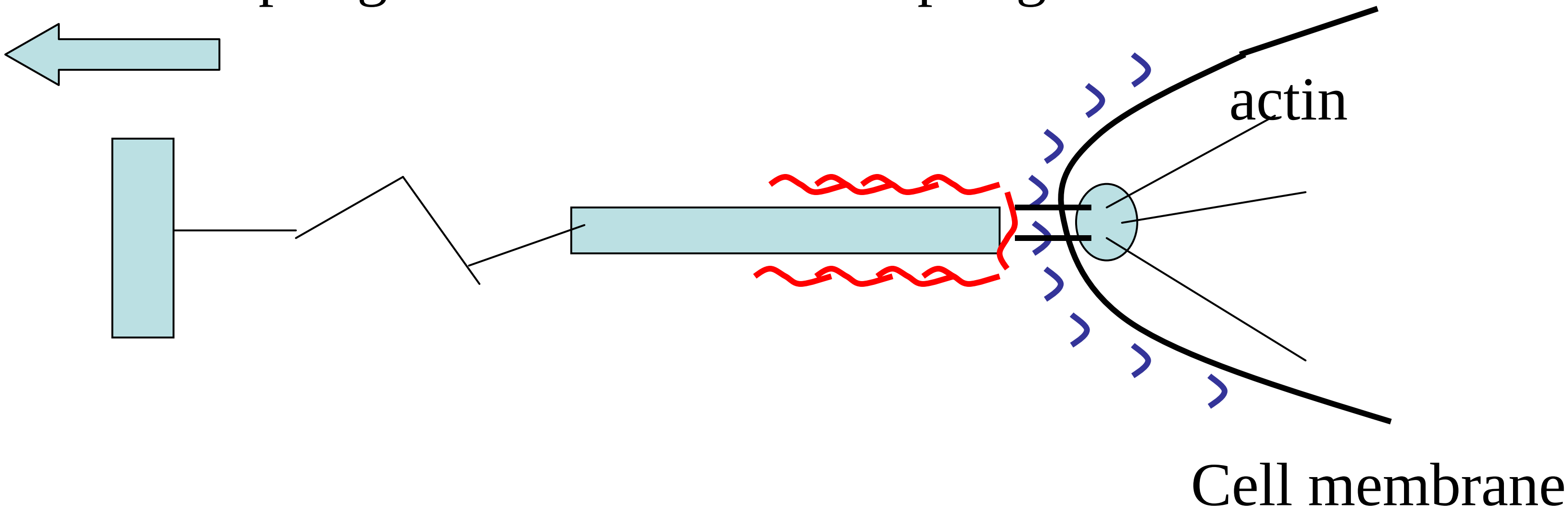
Basic idea

Functionalized probe contacts a cell and forms adhesion site



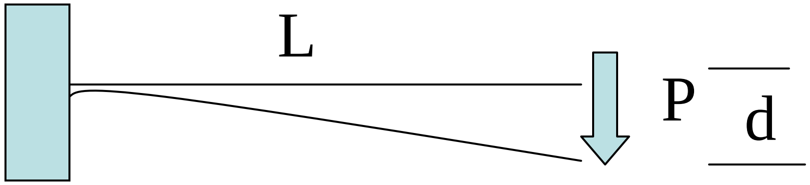
Basic idea

Probe is moved away from the cell. The cell applies a force on the spring. The force is measured from the spring deformation and its spring constant.



The cell may also be compressed or indented.

Cantilever as a mechanical spring



K (spring constant) = $3EI/L^3$

P = Kd

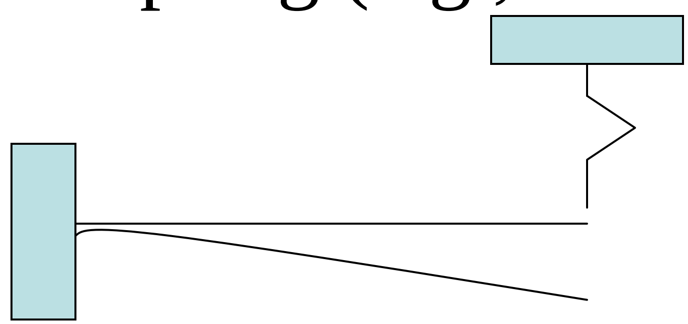
I=moment of inertia = width x depth^3/12

Typical K ~ 10 nN/µm

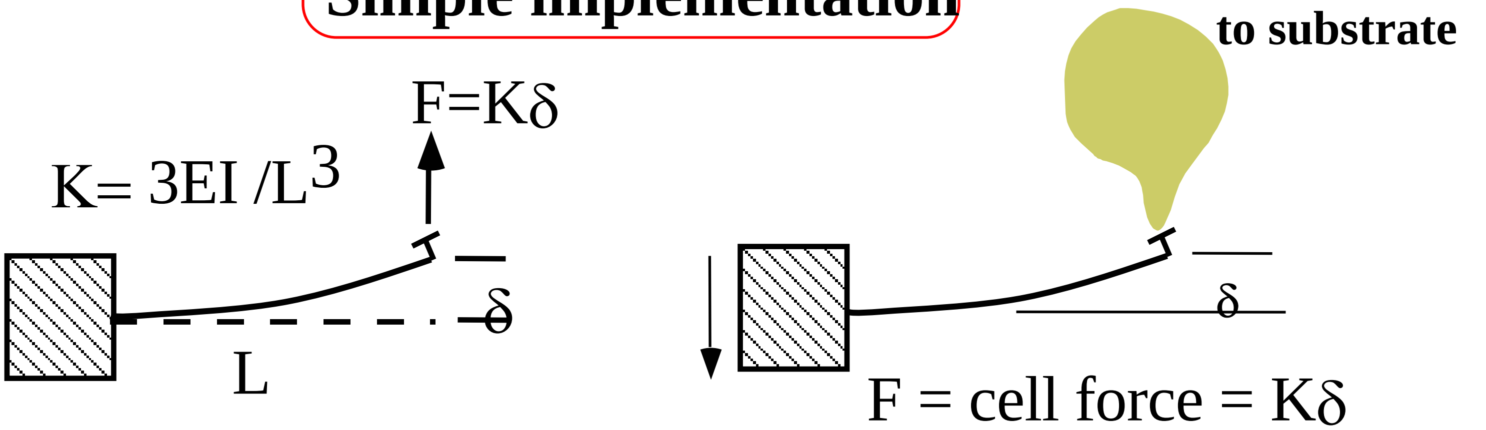
Calibration:

1) Resonant frequency, geometry, elastic property

2) Comparing with another spring (e.g., AFM)



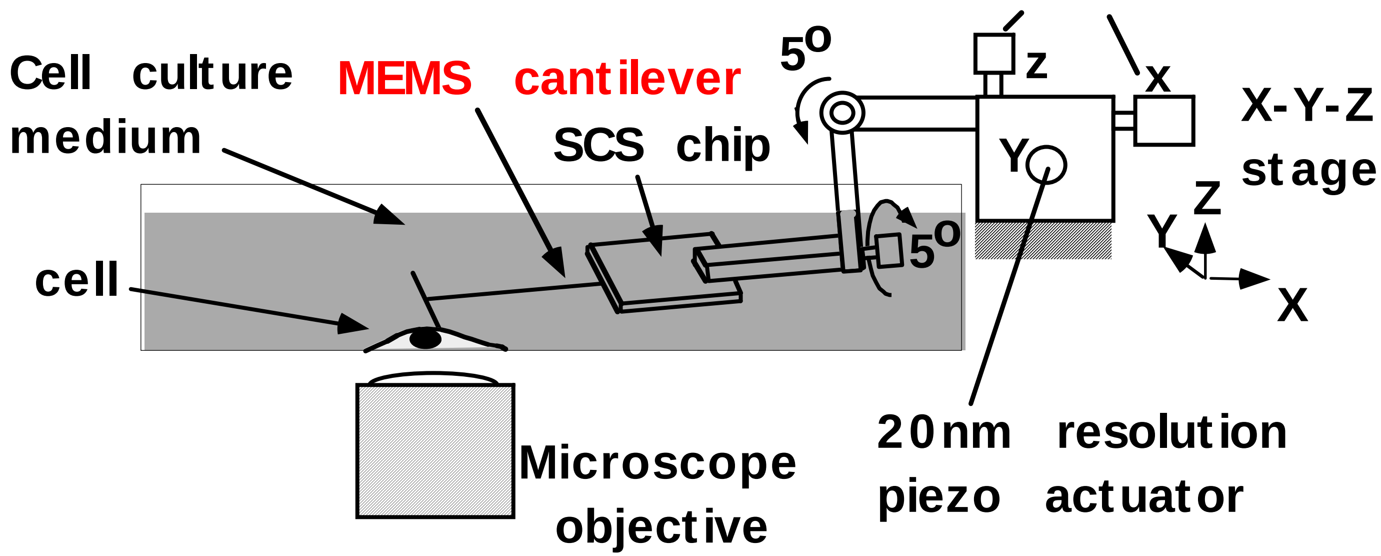
Simple implementation

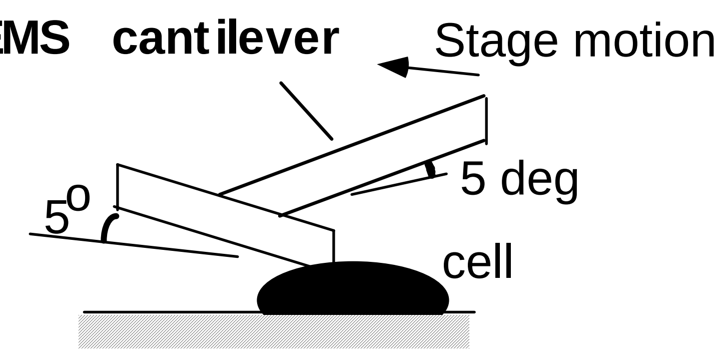


- Force sensor (such as a cantilever) is coated by fibronectin
- It is calibrated to determine spring constant, K
- The sensor tip is brought in contact with the cell - focal adhesion sites form
- It is moved away from the cell. The cell force is measured from the deformation δ

Advantages:
- Force sensor independently calibrated
- Force is applied at an anchorage site
- In-situ observation

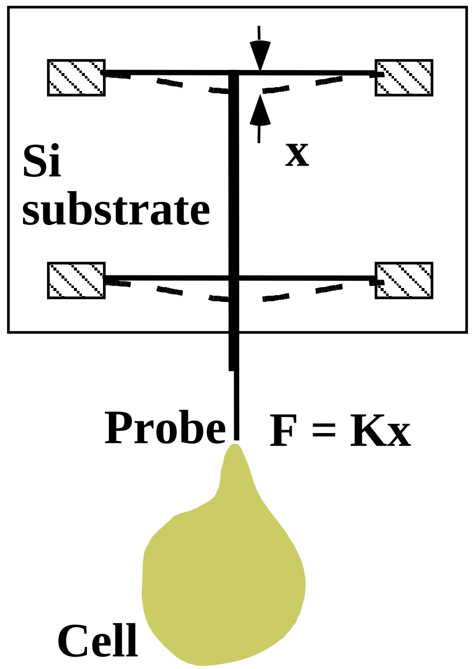
Experimental setup





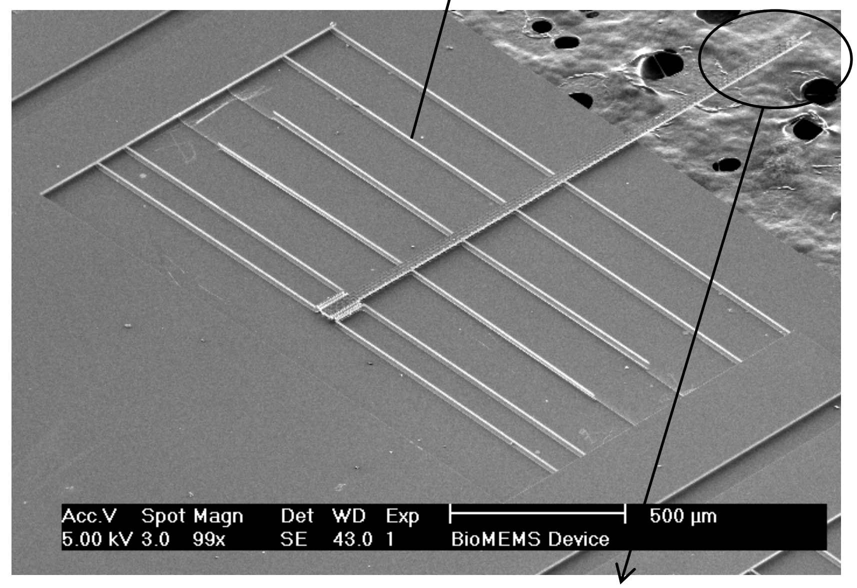
MEMS force sensor: beams anchored at both ends

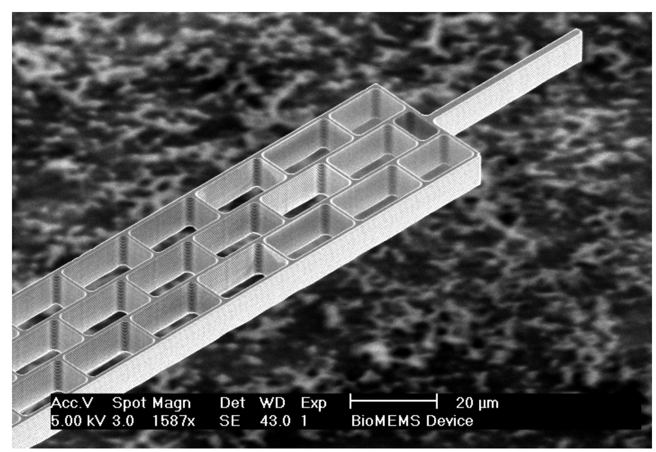
Example



K = 3.4 nN/µm

Flexural springs (1µm wide)



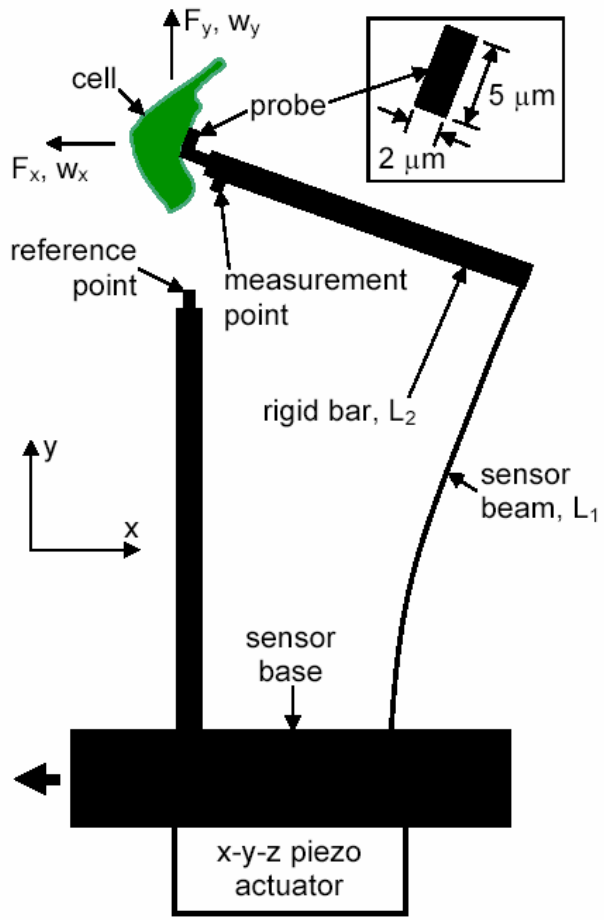


Probe

Yang and Saif.
Review of Scientific Instruments 76, 044301 (2005).

Example

2D force sensor

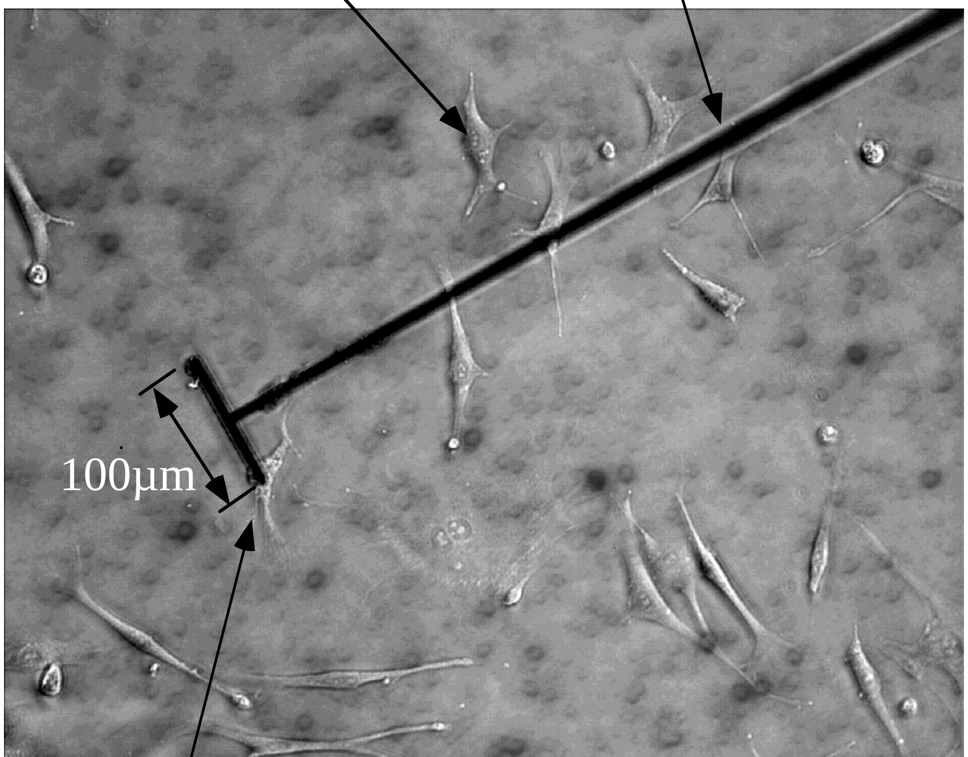


Endothelial cells in a culture dish with MEMS cantilever

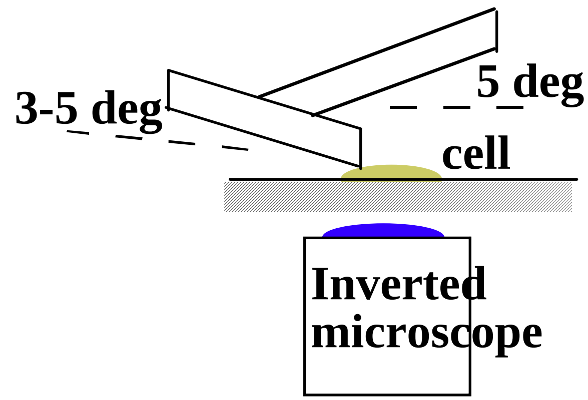
Example

Bovine endothelial cells on a substrate

MEMS cantilever coated with fibronectin (K=**18nN/μm**)

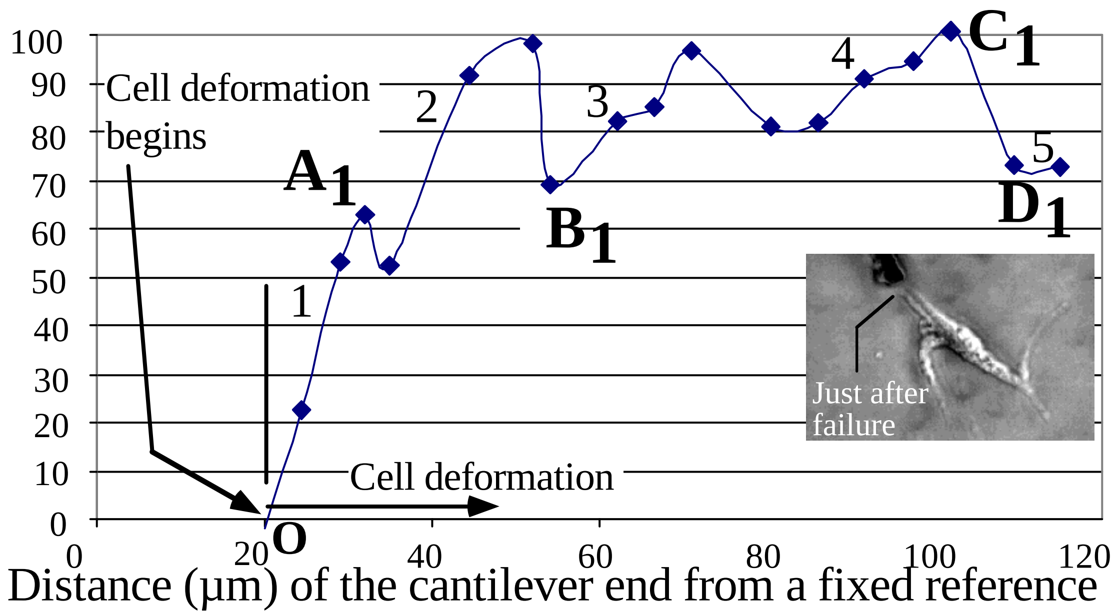


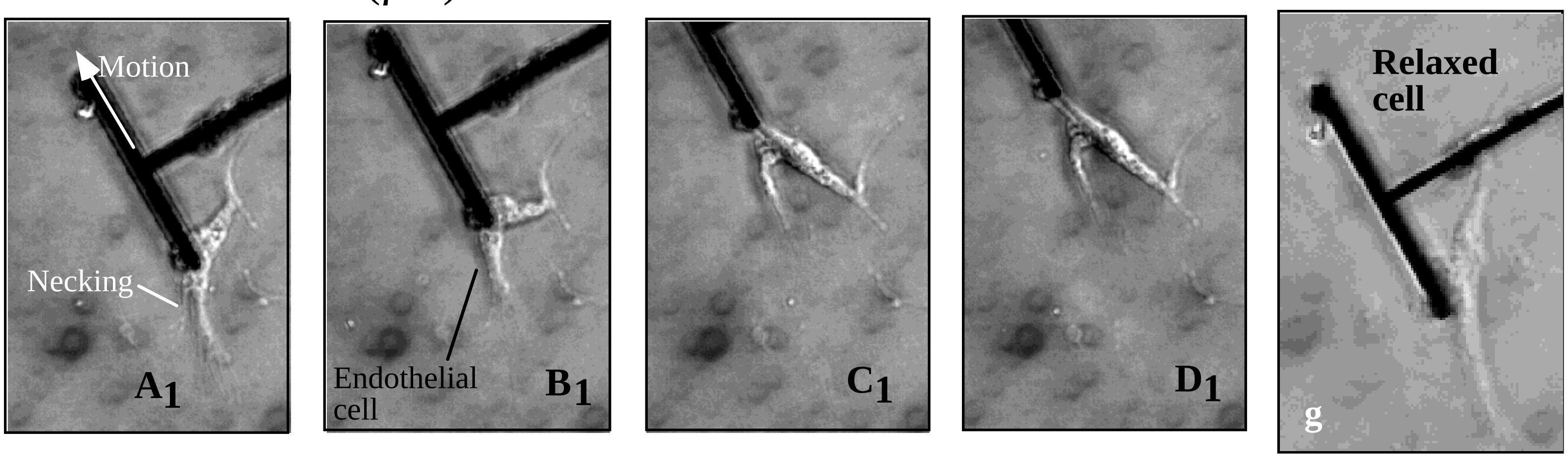
The MEMS cantilever has formed adhesive contact with the cell



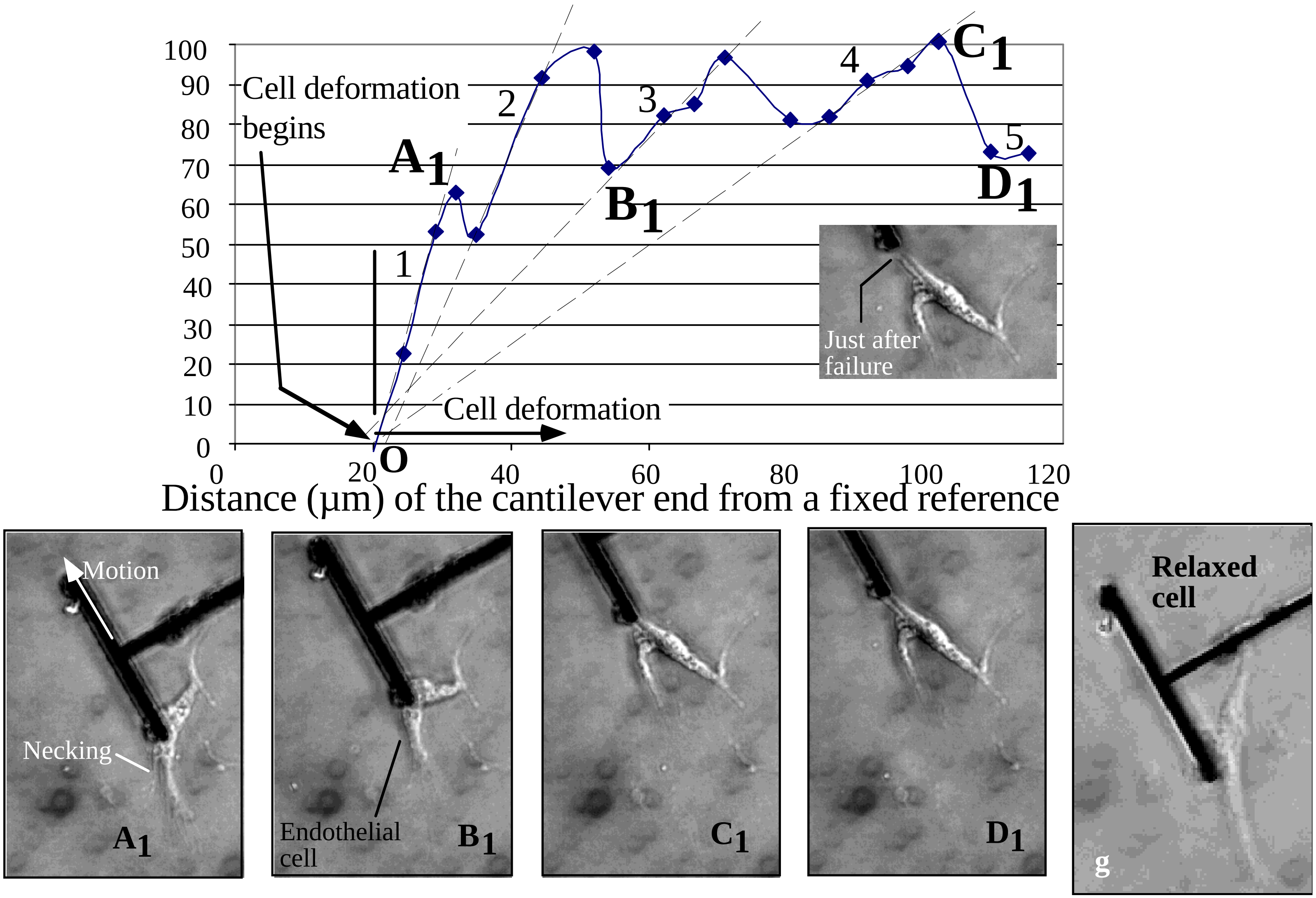
Schematic of the MEMS cantilever

Force response of an endothelial cell

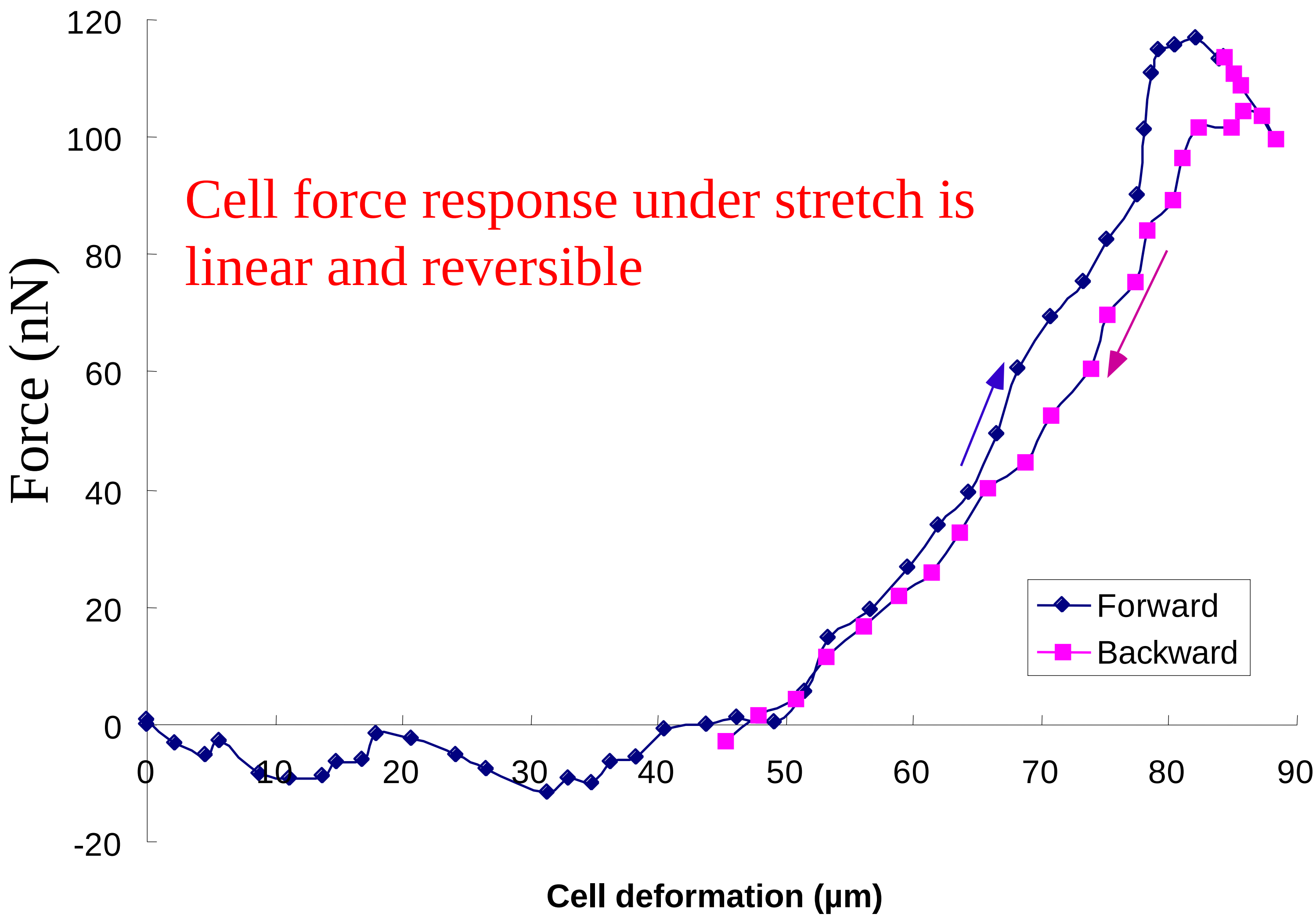




Force response of an endothelial cell

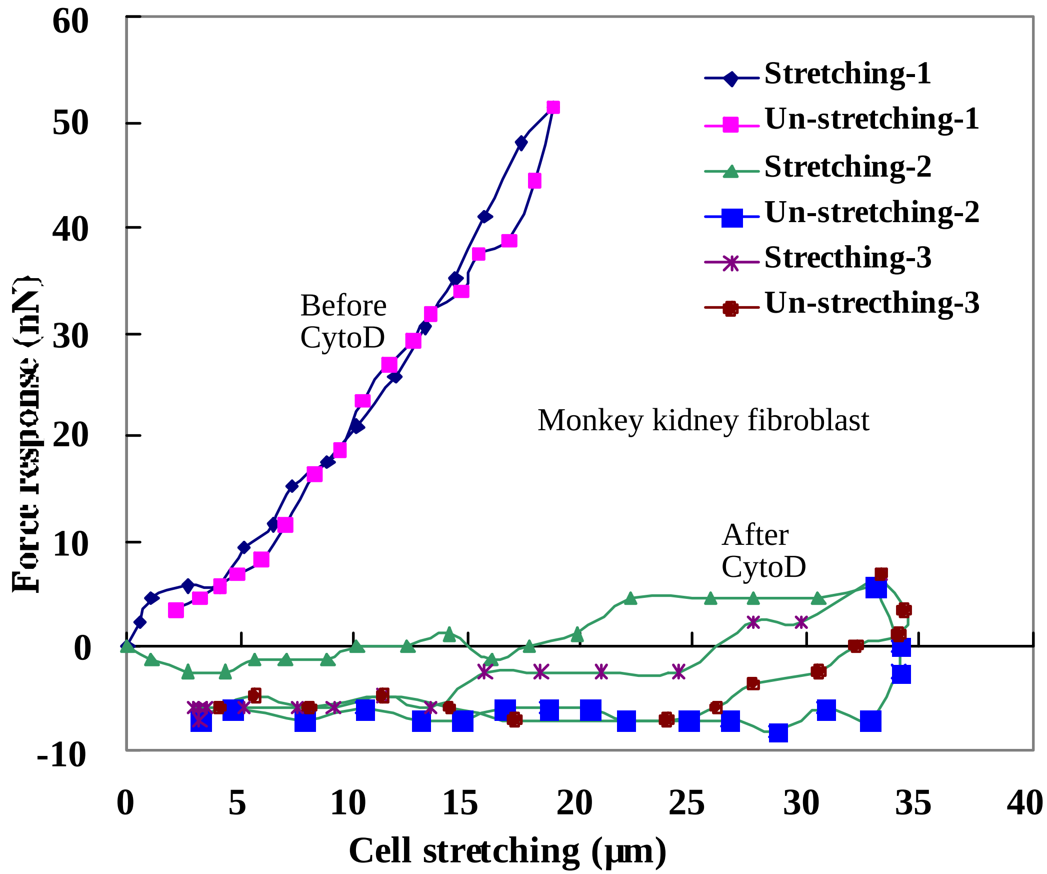


Force response of a monkey kidney fibroblast cell



Yang and Saif. Experimental Cell Research 305 (2005) 42– 50.

Cyto-D treatment disrupts force bearing capacity

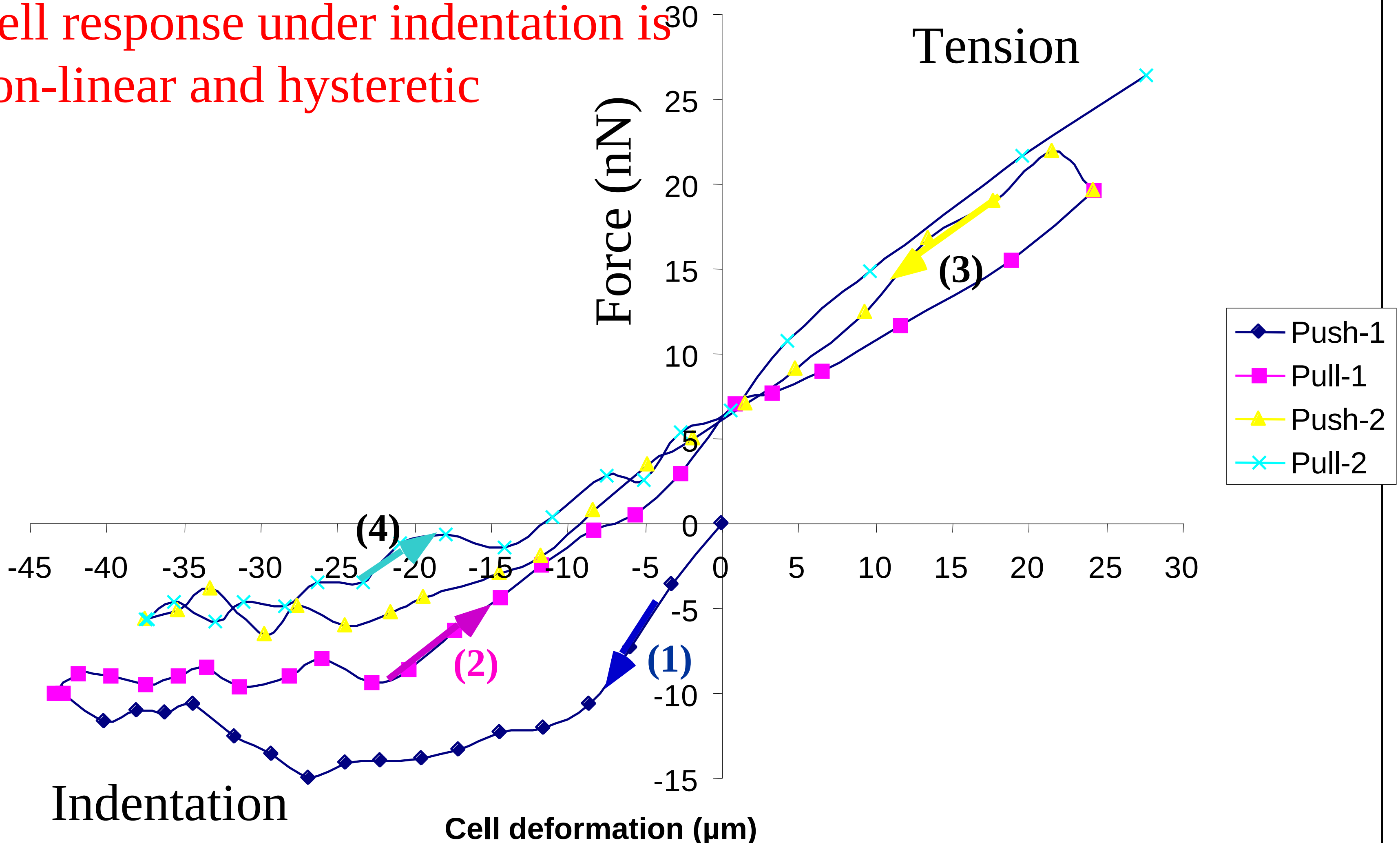


Cyto-D disrupts actin network

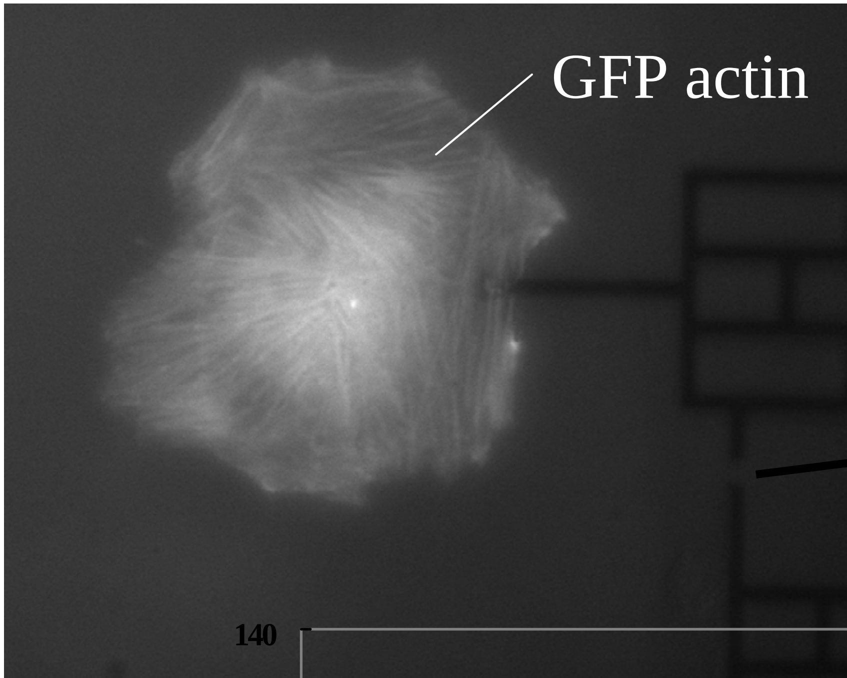
Yang and Saif. Experimental Cell Research 305 (2005) 42– 50.

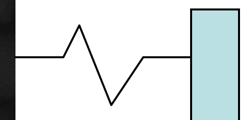
Force response of a monkey kidney fibroblast cell

Cell response under indentation is non-linear and hysteretic

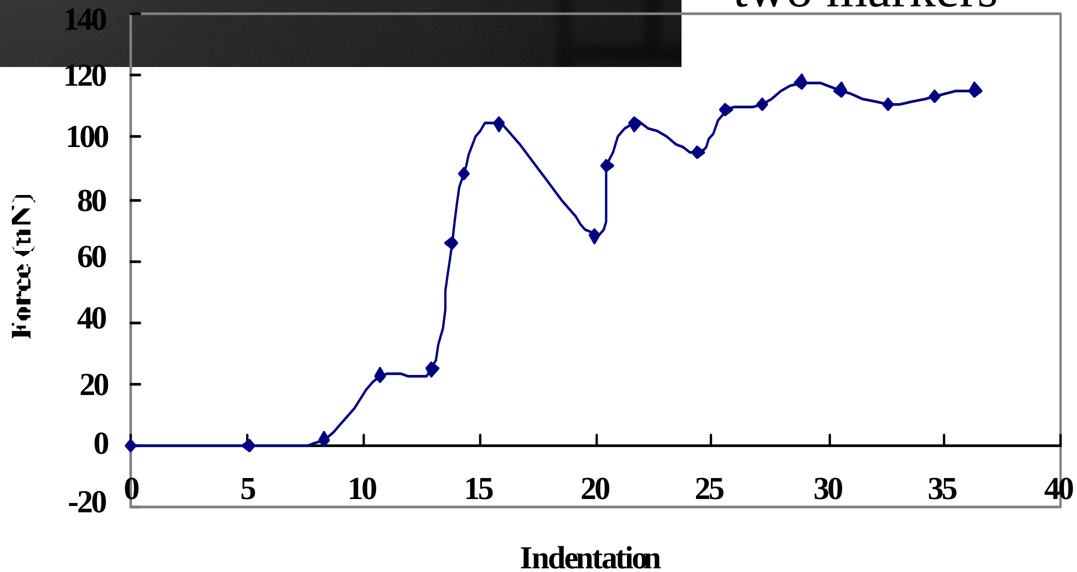


Mechanism of non-linearity and irreversibility under indentation

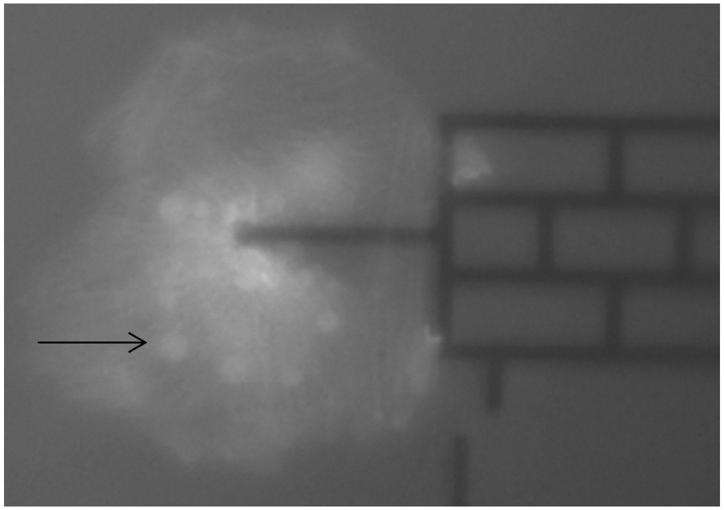


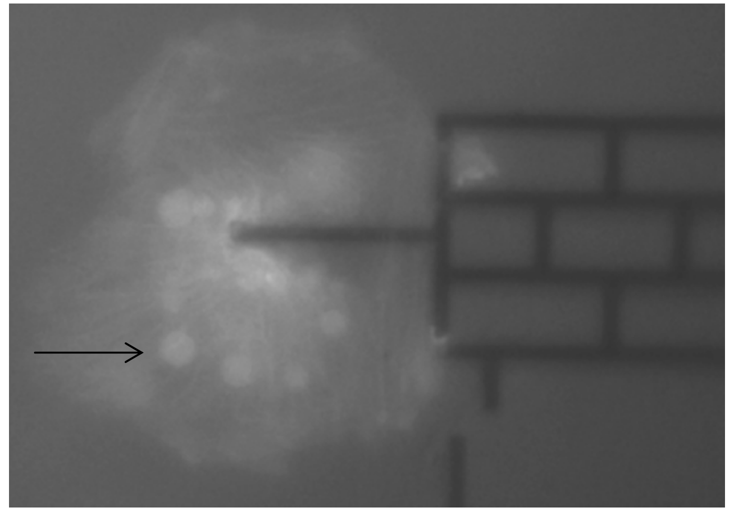


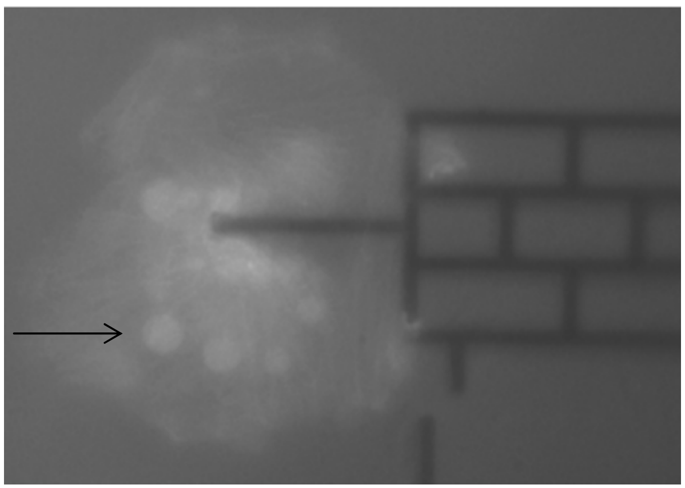
Force measured from change of gap between two markers

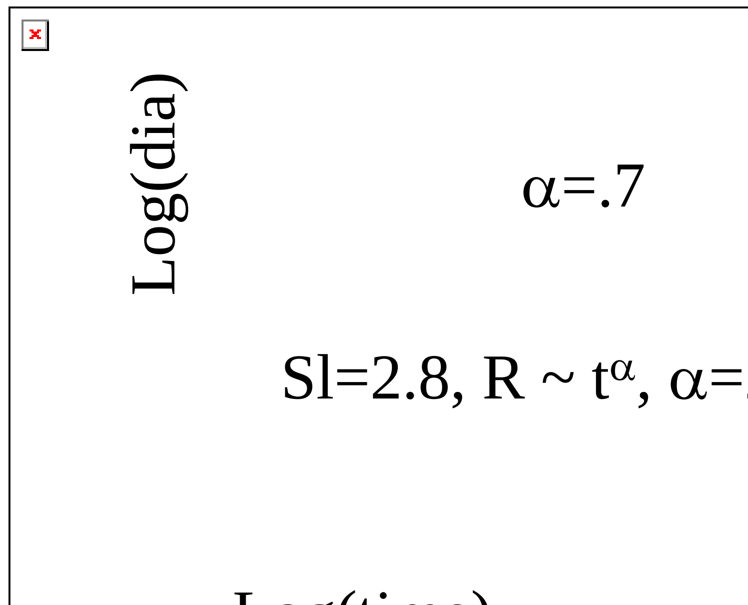


Actin agglomerates irreversibly under indentation



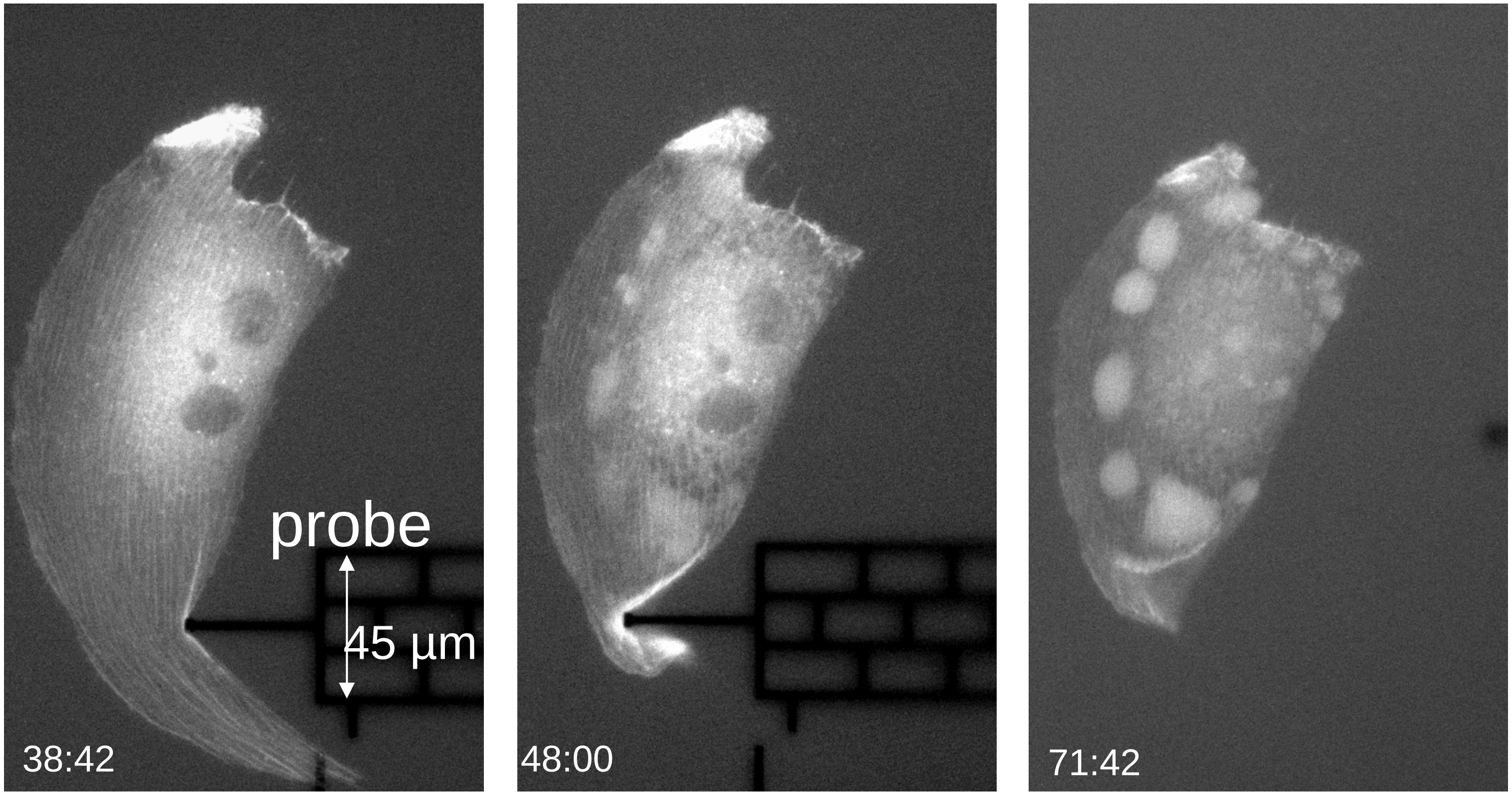






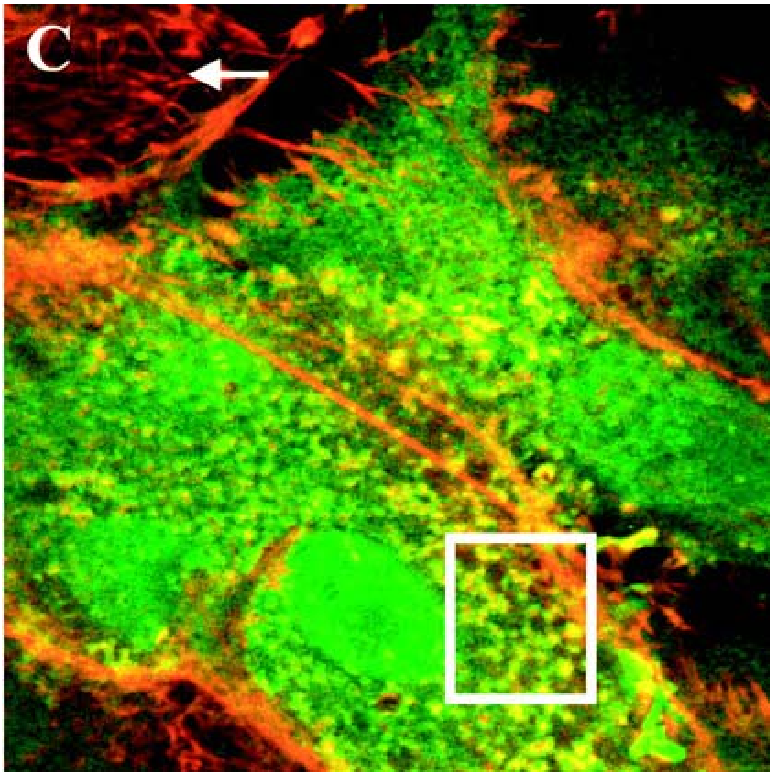
Yang and Saif
Actabiomaterialia 2006 (in press)

More evidence of actin agglomeration



Monkey kidney fibroblast subjected to mechanical indentation (injury simulation). Here actin stress fibers are highlighted by green florescent protein (GFP). In response to indentation, the cell signals local actin agglomeration at discrete locations. Such actin agglomeration is also observed in various physiological conditions such as during ischemic attack in kidney cells. This is the first evidence of actin agglomeration due to mechanical stimulus (Shengyuan and Saif, Actabiomaterialia 2006, in press).

Actin agglomeration in physiological condition: ischemic attack



Porcine kidney cells

Ashworth et al. Am J. Physiol Renal Physiol 284: F852, 2003.

Why MEMS bio sensors:

1. Force range: 1-100 nN (natural progression from optical tweezer, magnetic beads, AFM)

1. Flexibility of design (cell contact region may be designed in a variety of fashions)

3. Large cell deformation range (sub μm-10s of μm)