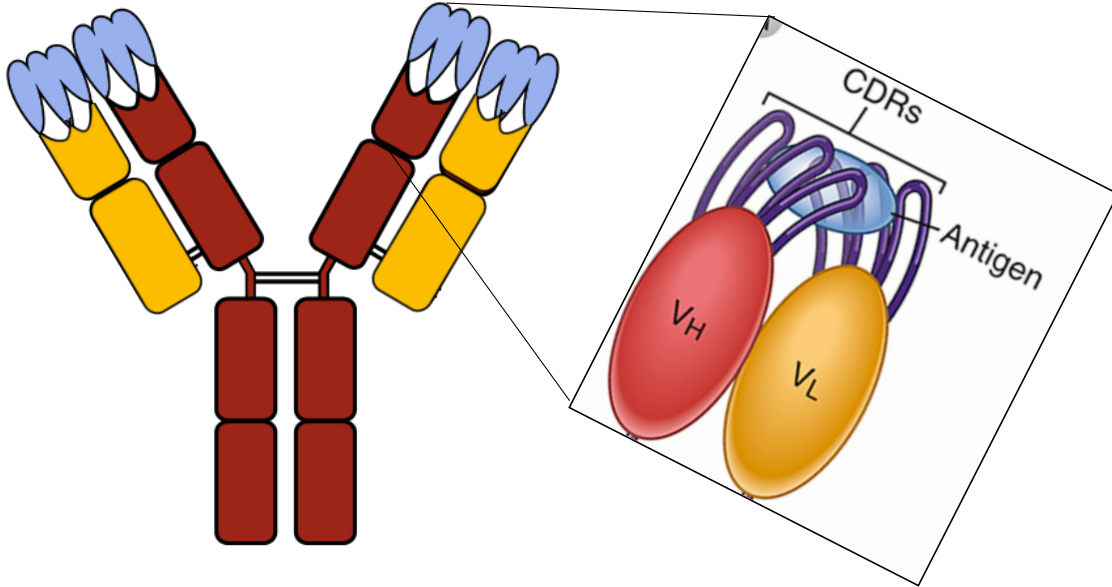


The antigen- antibody interaction

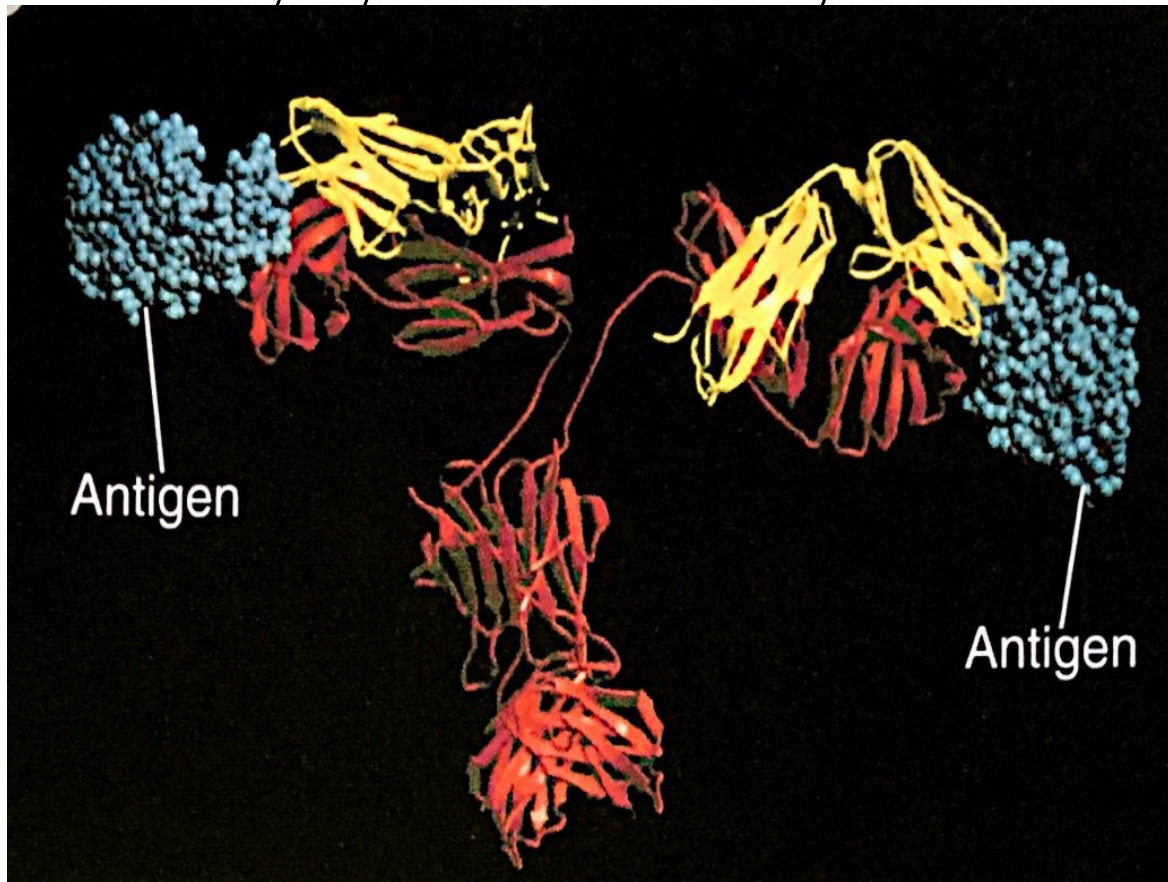
Characterization of scFvs that bind lysozyme



- The goal of this screen is to find a scFv clone with improved binding to lysozyme
- Antibody with a lower K_d for its antigen means a more stable interaction and a higher affinity
- We sorted a library of scFv yeast that bind to lysozyme
- Today will determine the dissociation constant of a single clone scFv with lysozyme

CDRs generate antigen binding site specificity

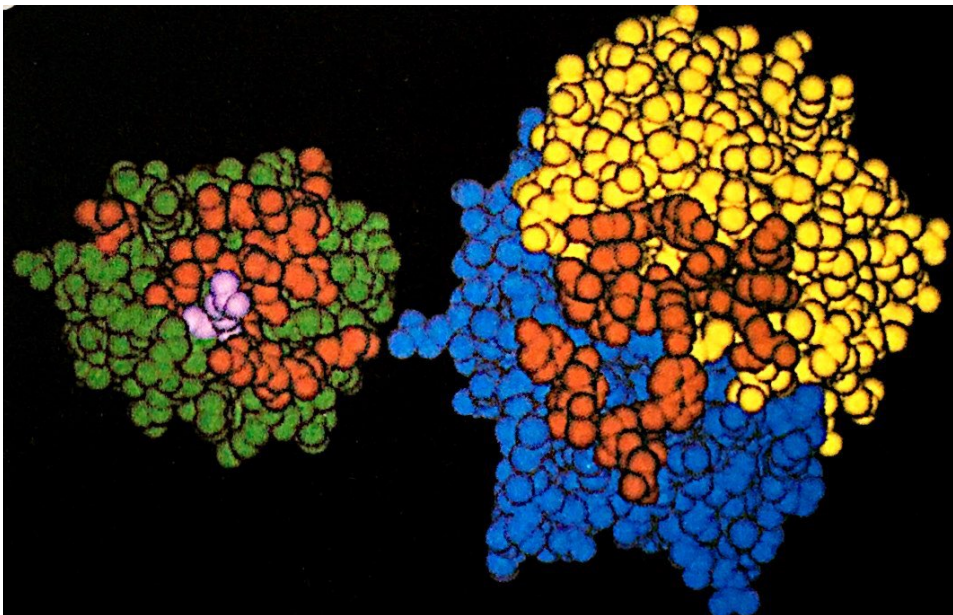
Lysozyme bound to antibody



- Specificity, degree to which an antibody differentiates between different antigens
- Finger-like CDRs usually recognize 15-22 amino acids
- Basic antibody structure maintained (β strands) when variability confined to CDR loops

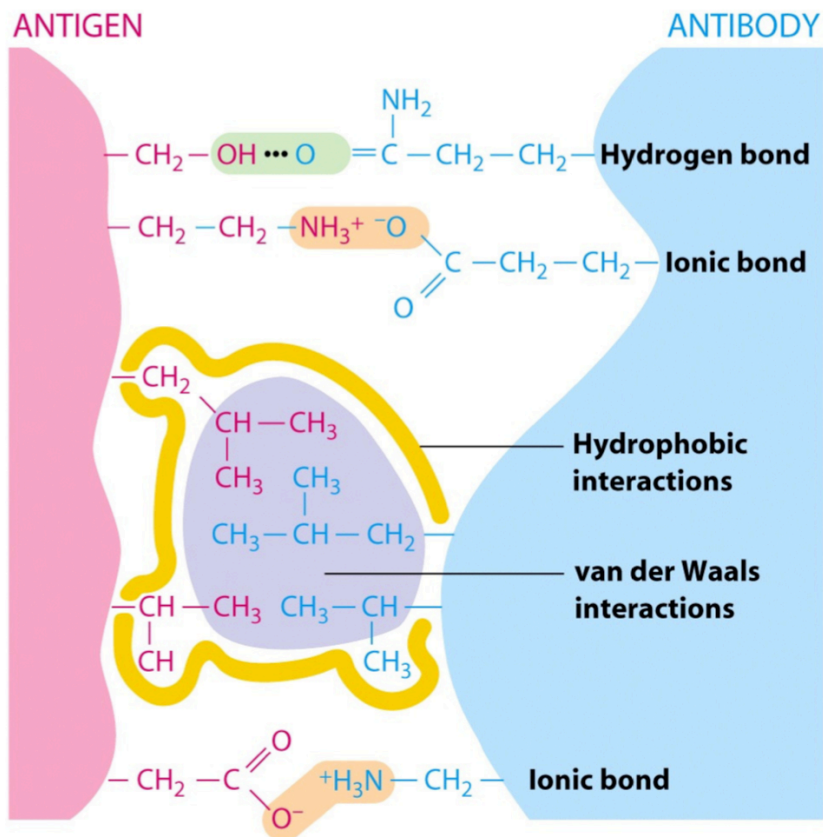
The Antigen - Antibody interaction forms multiple contacts

3D: Lysozyme bound to variable region



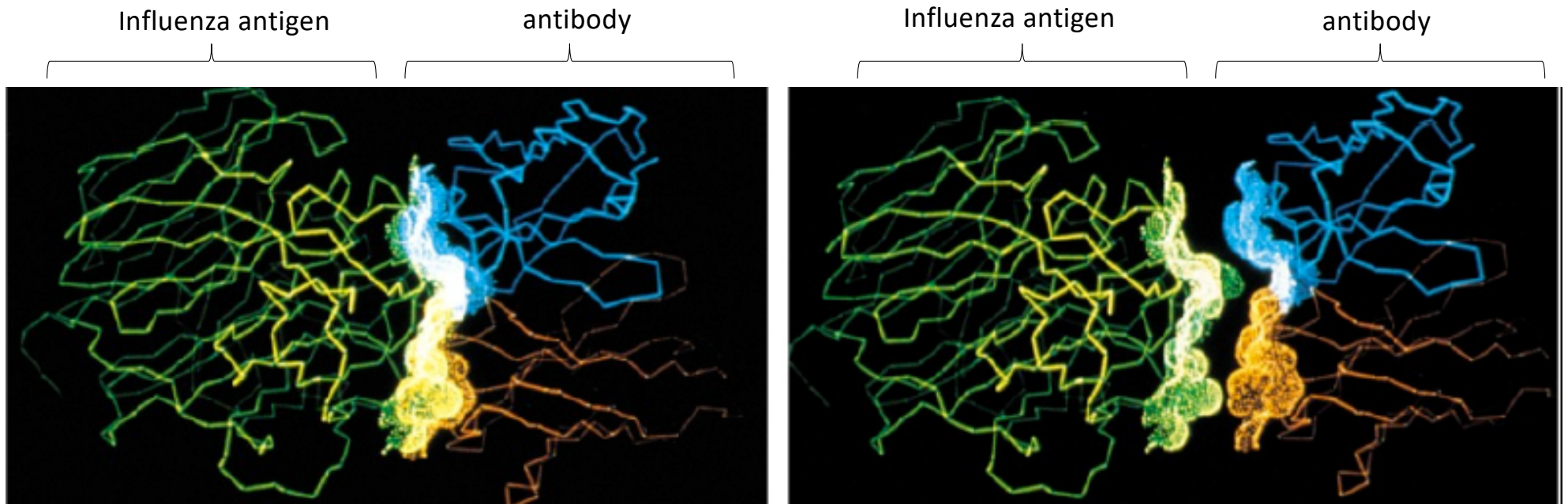
- Green: lysozyme
- Blue/Yellow: V_L and V_H
- Red amino acids that interact
- Pink critical glutamine residue fits into cleft of CDR

Noncovalent bonds form the basis of the antibody binding site

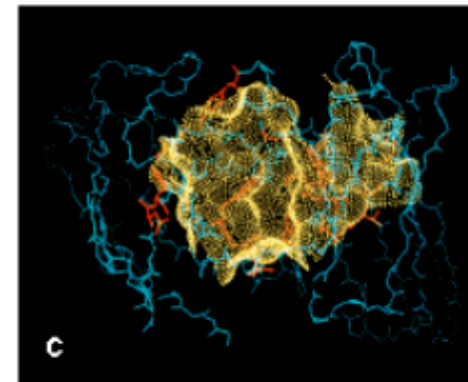
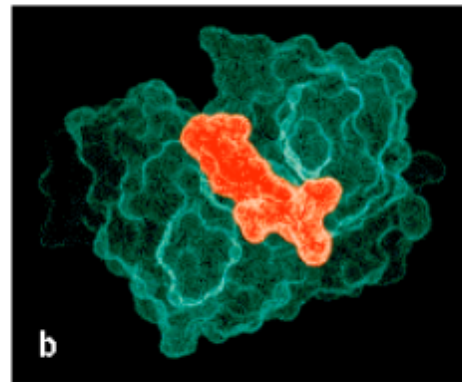
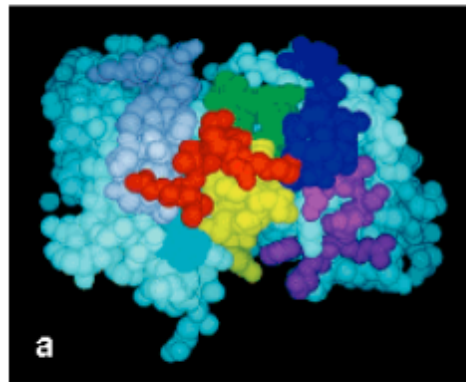
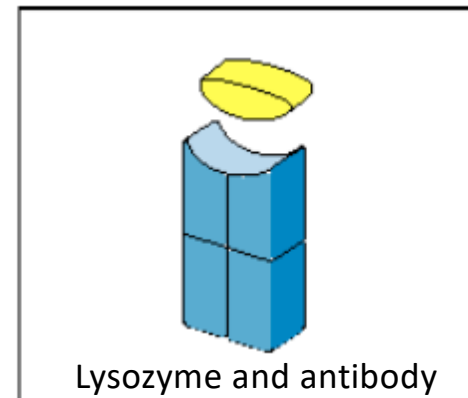
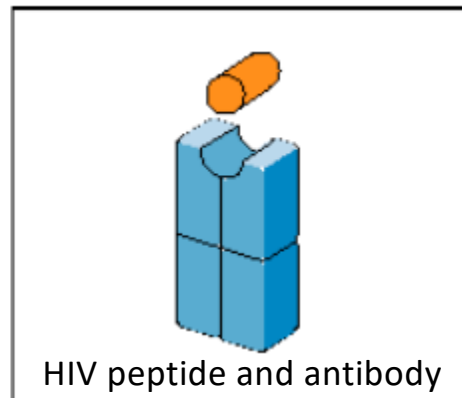
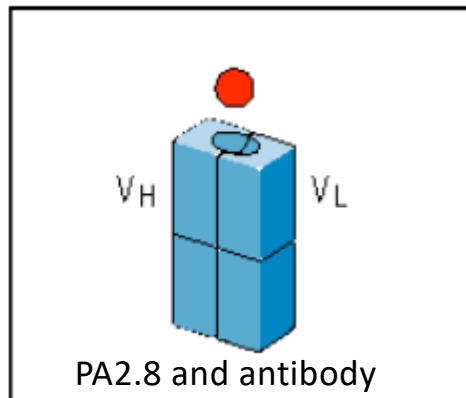


- Strength of each of these noncovalent interactions is weak
 - Many noncovalent bonds are required to form a strong interaction
- Each of these interactions operates over a very small distance ($\sim 1 \text{ \AA}$)
- This requires a high degree of complementarity between the CDR of the antibody and the antigen

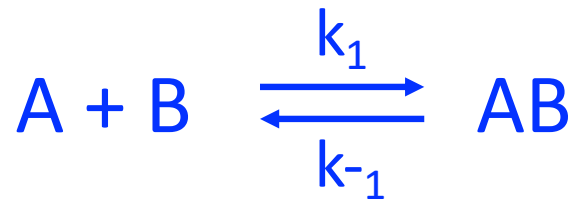
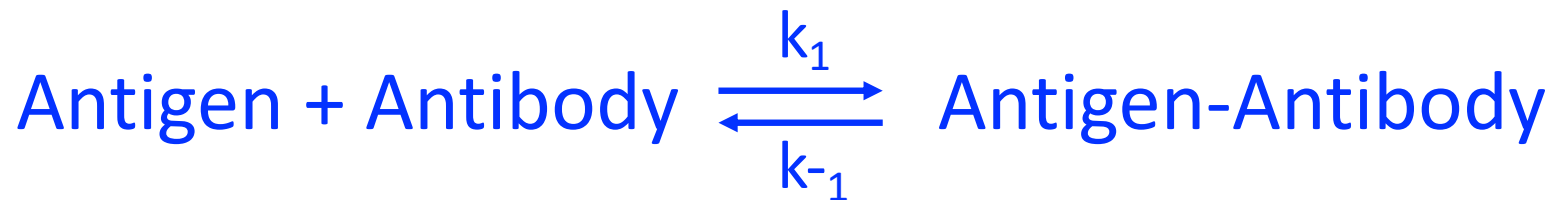
Influenza antigen and antibody binding illustrates complementary when separated by 8 Å



Large variation in antibody binding pockets



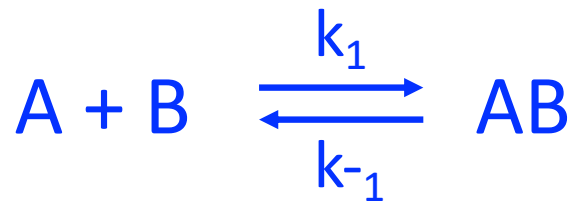
Binding a monovalent antigen by an antibody can be described by a bimolecular equation




k_1 =rate of association

k_{-1} =rate of disassociation

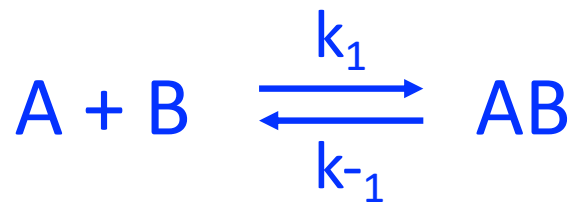
The equilibrium association constant (K_a) is a good indicator for antibody affinity




$$K_a = \frac{[AB]}{[A][B]}$$


- Ratio of products to reactants
- Affinity, the strength of the total noncovalent interactions between one antigen and antibody
- Units of K_a are concentration⁻¹
- Example: nM⁻¹

Equilibrium dissociation constant (K_d) is an indicator of the stability of a complex



$$K_d = \frac{[A][B]}{[AB]}$$


- Ratio of reactants to products
- Antibodies produced in a typical immune response usually varied from $K_d = 10^{-7}$ (~100nM) to 10^{-9} (~1nM)
- Units of K_d are concentration
- The smaller the K_d the more stable the interaction

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Range of biologically important interactions

Antibody-antigen
interactions

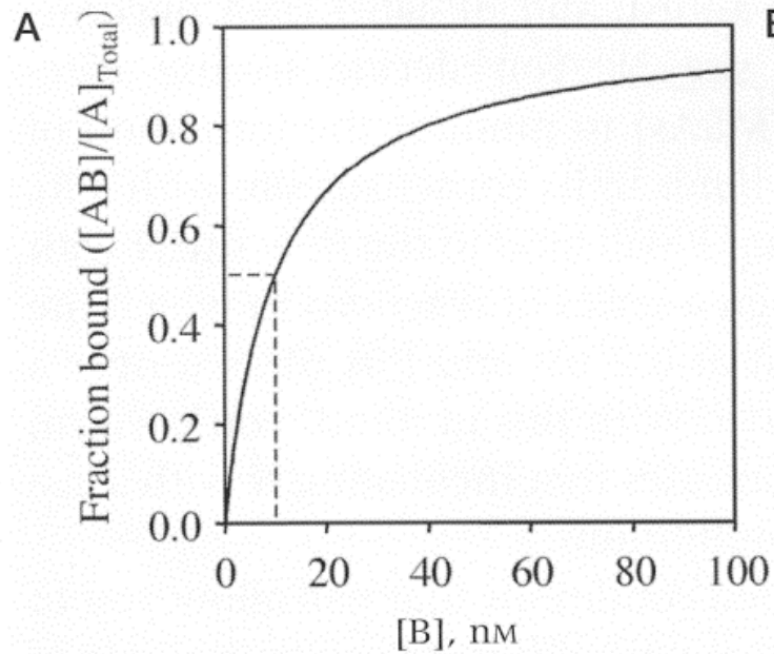
Type of Interaction	K_D (molar)	ΔG_{bind}^0 (at 300K) kcal/mol
Enzyme:ATP	$\sim 1 \times 10^{-3}$ to $\sim 1 \times 10^{-6}$ (millimolar to micromolar)	-4 to -8 kcal/mol
signaling protein binding to a target	$\sim 1 \times 10^{-6}$ (micromolar)	-8 kcal/mol
Sequence-specific recognition of DNA by a transcription factor	$\sim 1 \times 10^{-9}$ (nanomolar)	-12 kcal/mol
small molecule inhibitors of proteins (drugs)	$\sim 1 \times 10^{-9}$ to $\sim 1 \times 10^{-12}$ (nanomolar to picomolar)	-12 to -17 kcal/mol
biotin binding to avidin protein (strongest known non-covalent interaction)	$\sim 1 \times 10^{-15}$ (femtomolar)	-21 kcal/mol

higher K_D value
weaker interaction

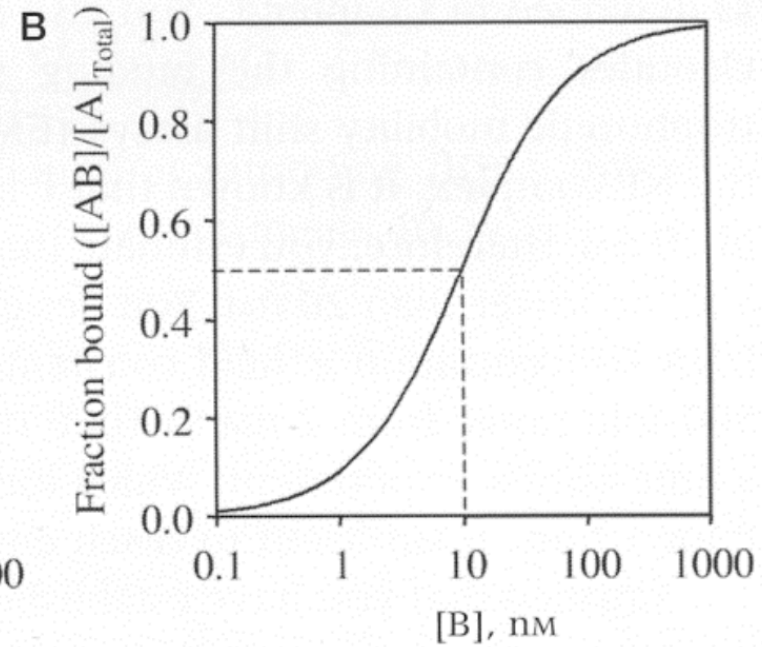
lower K_D value
stronger interaction

Logarithmic vs. Linear display of data

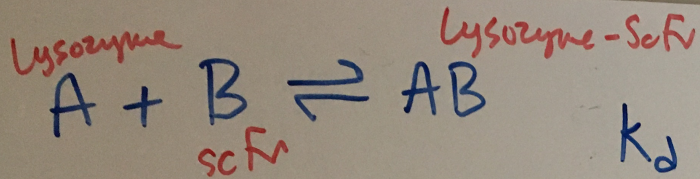
historic convention



current convention

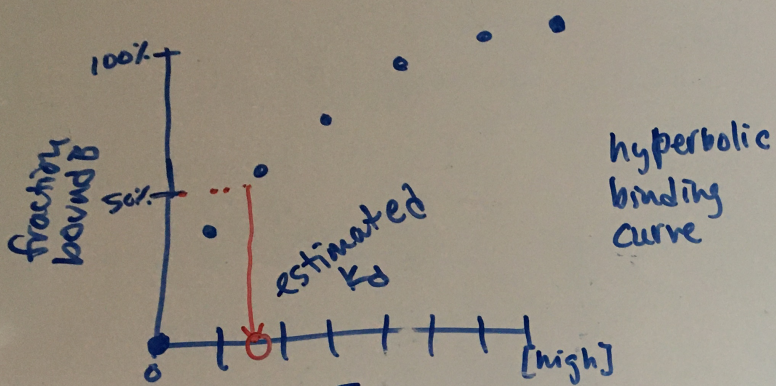


Biomolecular binding interaction at equilibrium:
Why is antibody dissociation constant (K_d) equal to the antigen concentration at which 50% antibody is bound to antigen?



Lysozyme-ScFv

$$K_d = \frac{[A][B]}{[AB]}$$



$$\text{fraction bound B} = \frac{[AB]}{[B] + [AB]} = \frac{\frac{[A][B]}{K_d}}{\frac{[B]}{K_d} + 1} = \frac{[A]}{K_d + [A]}$$

$$\frac{[B]}{[AB]}$$

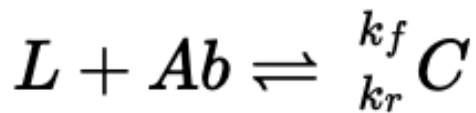
$$K_d = \frac{[A][B]}{[AB]}$$

$$\frac{1}{[A]} = \frac{[B]}{[AB]}$$

$$\text{fraction bound B} = \frac{[A]}{K_d + [A]} = \frac{1}{2} \text{ or } 50\%$$

$$K_d = [A]$$

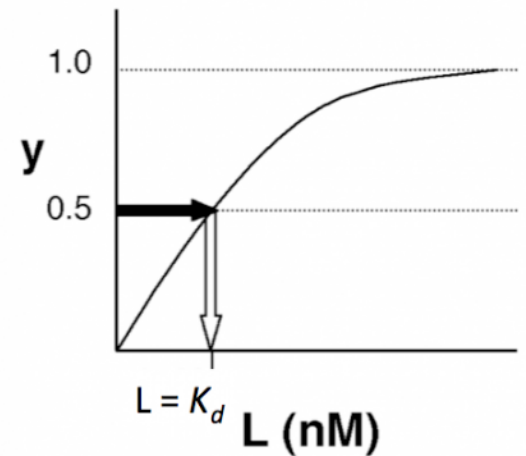
Mathematical relationship between fraction bound and free reactant makes estimations easy



$$y = \frac{[L]}{[L] + K_d}$$

If L in excess (in solution), and $[L] = L$ constant

- at $L = K_d$ $y = 0.5$
- if $L \ll K_d$ then $y \approx \frac{[L]}{K_d}$ (linear relationship)
- if $L \gg K_d$ then $y \approx 1$ (at saturation)



Alternative methods to measure binding dynamics without necessitating equilibrium

Binding of the antibody to the antigen alters the resonance readout and can translate to affinity

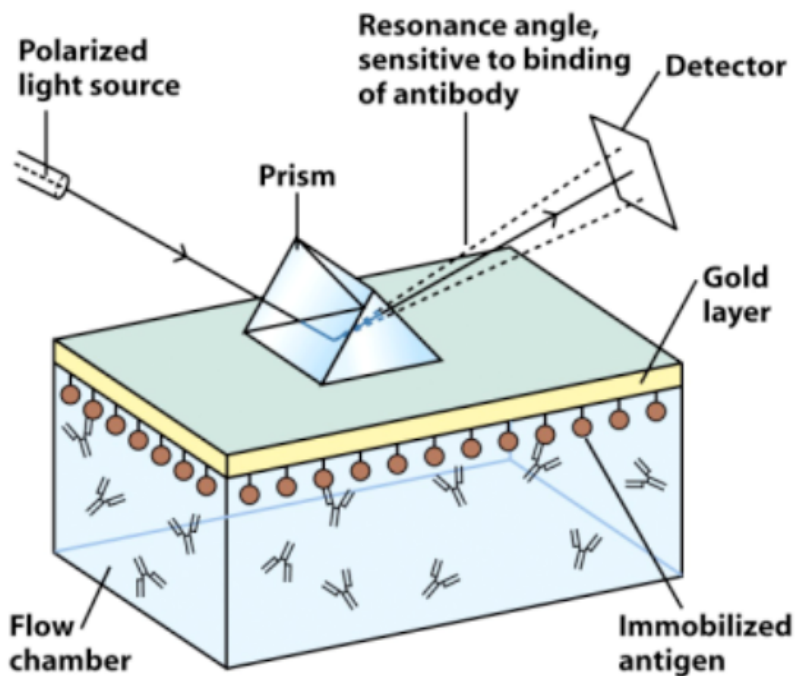
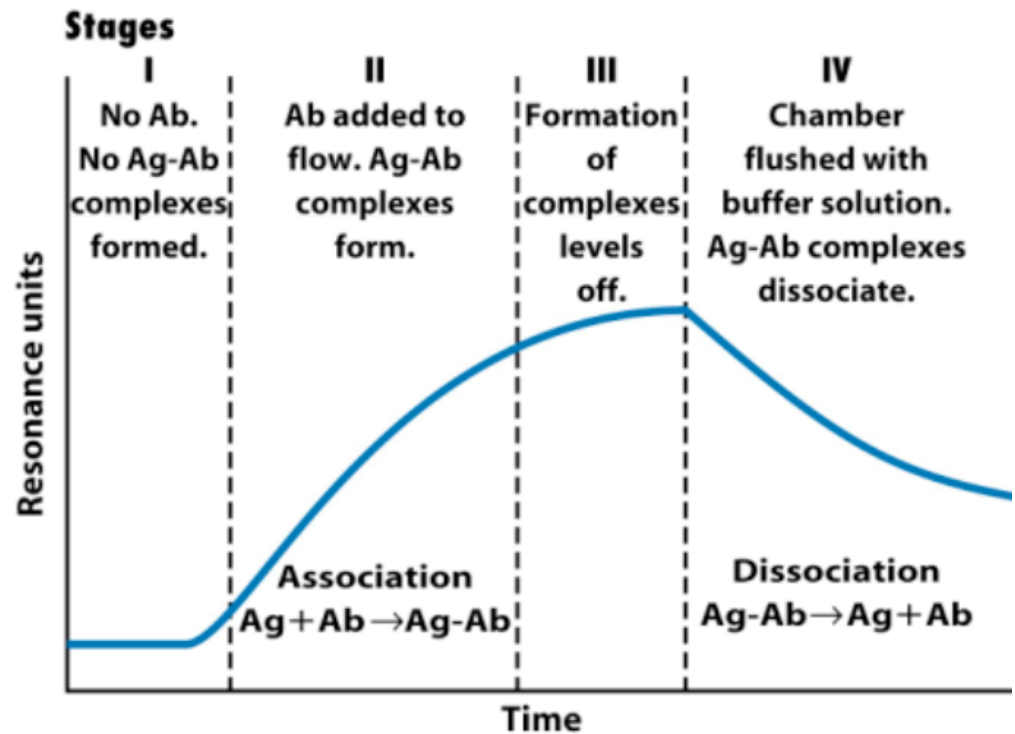
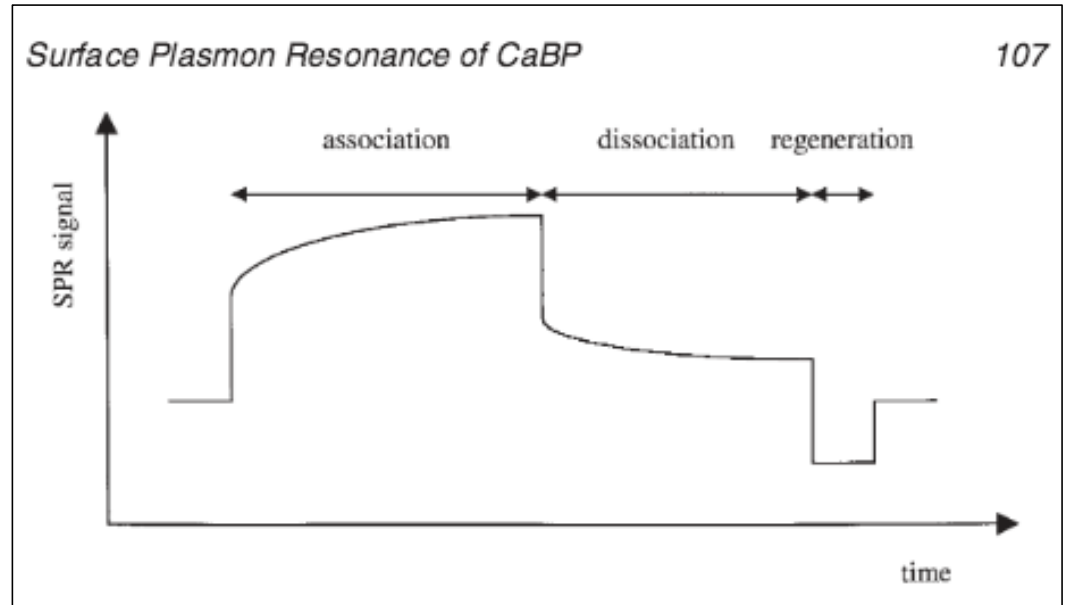
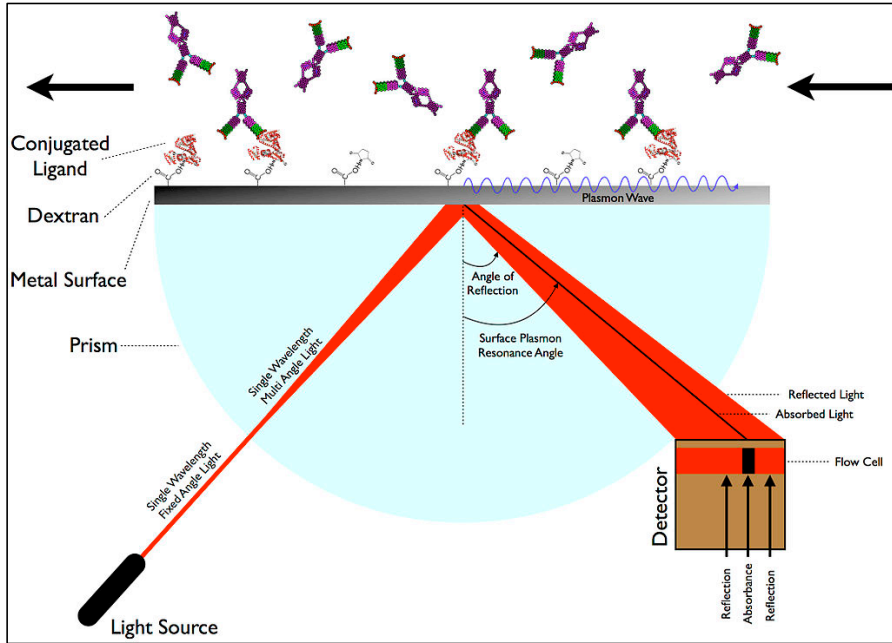


Figure 6-4a
Kuby IMMUNOLOGY, Sixth Edition
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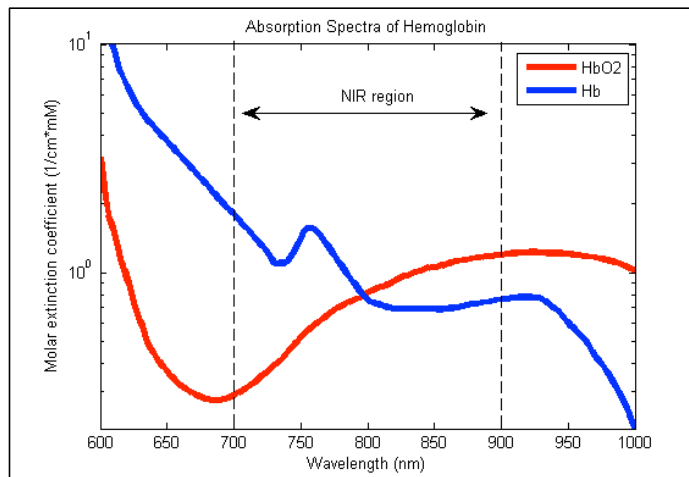


Surface plasmon resonance (Biacore)

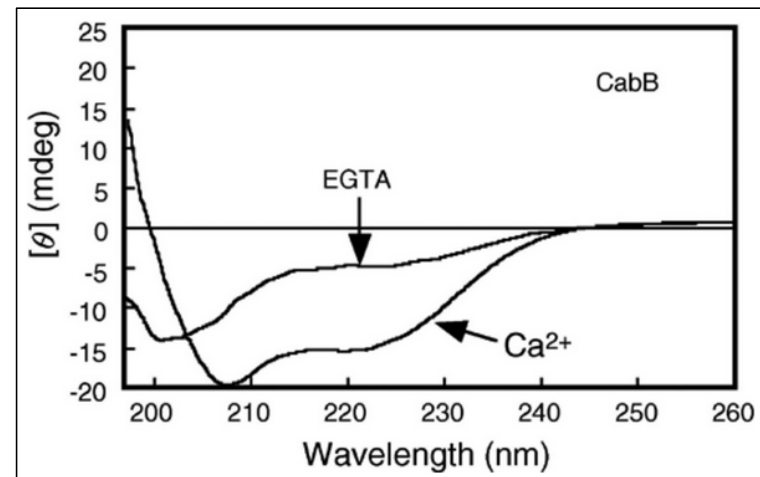


Binding may be quantified using methods other than fluorescence

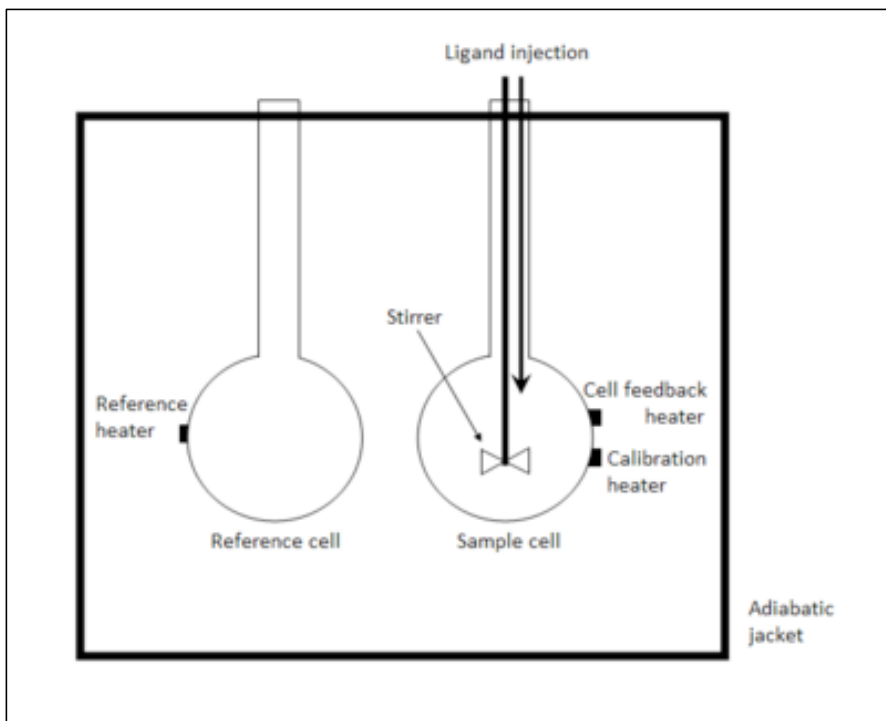
- absorbance spectroscopy
e.g. hemoglobin binding to O₂



- circular dichroism
e.g. Ca²⁺ binding to CabB

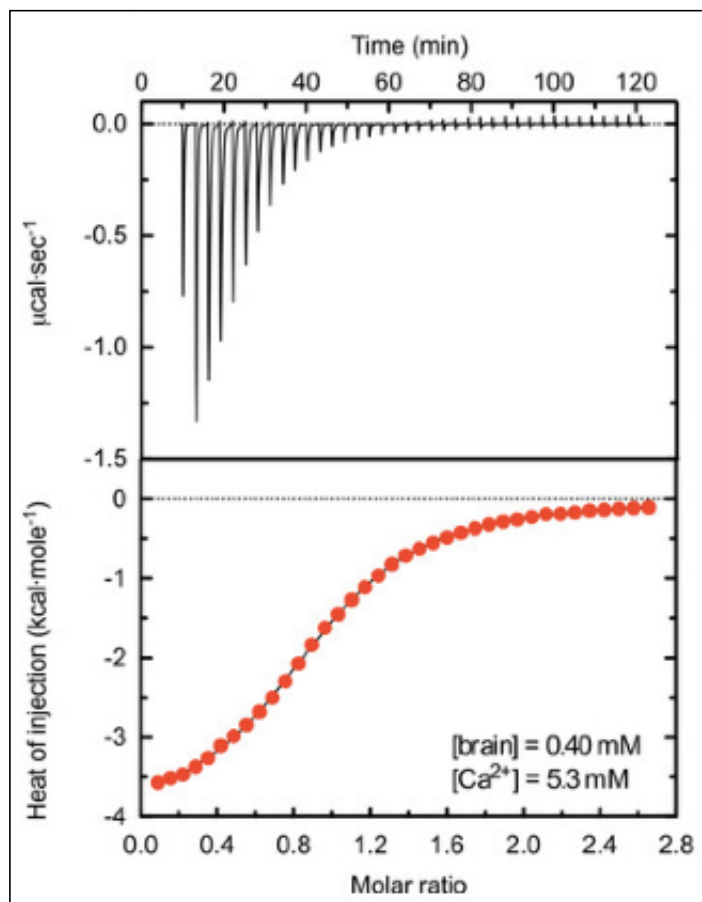


Isothermal titration calorimetry measures thermodynamic parameters of interactions



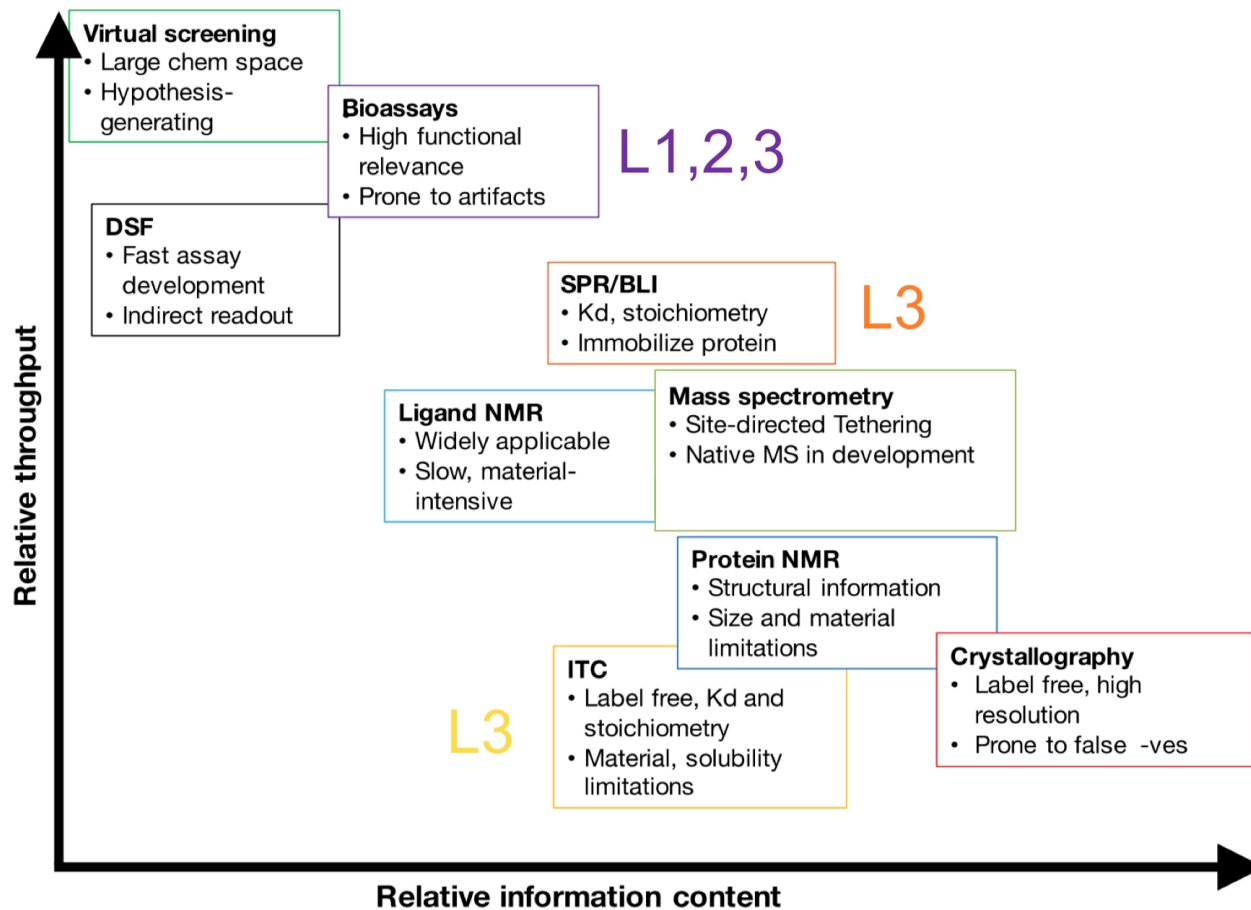
$$\Delta G = -RT \ln K_a = \Delta H - T\Delta S$$

Backman (2015) *PeerJ* 3: e944



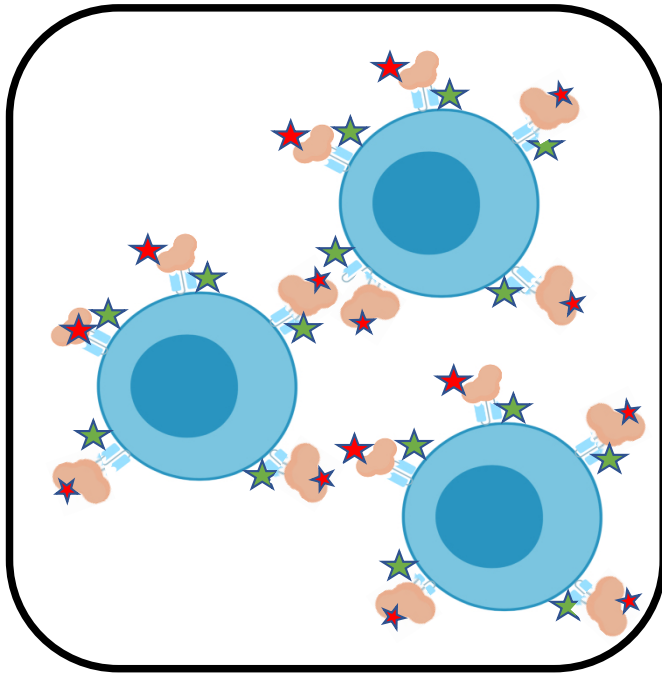
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Methods to evaluate binding interactions

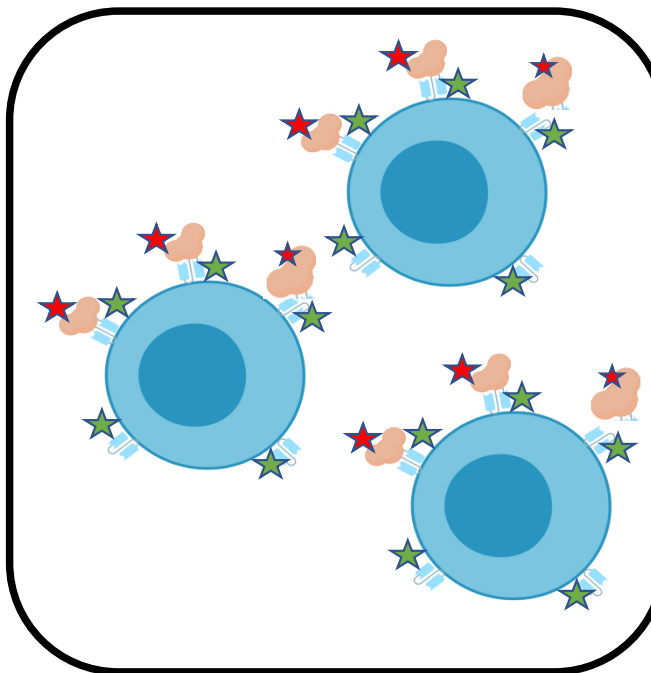


Practically: how will we measure equilibrium binding with different antigen concentrations?

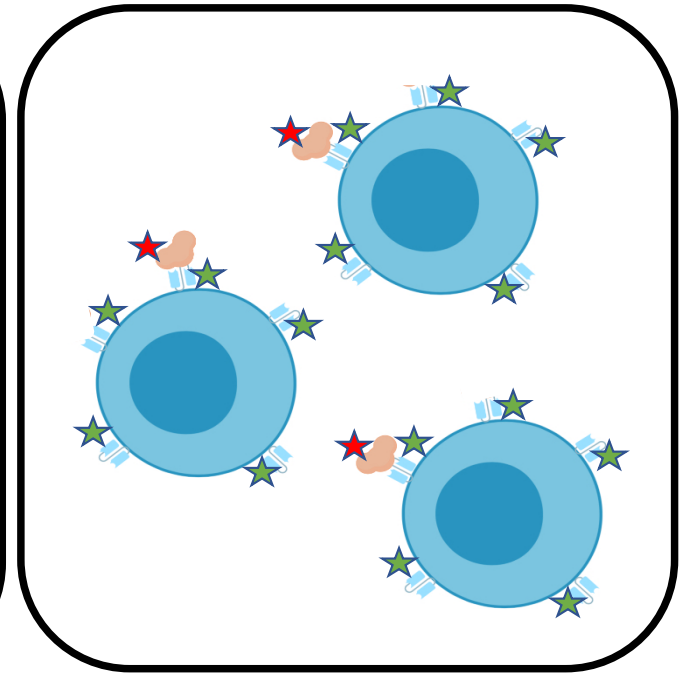
Tube #1



Tube #2

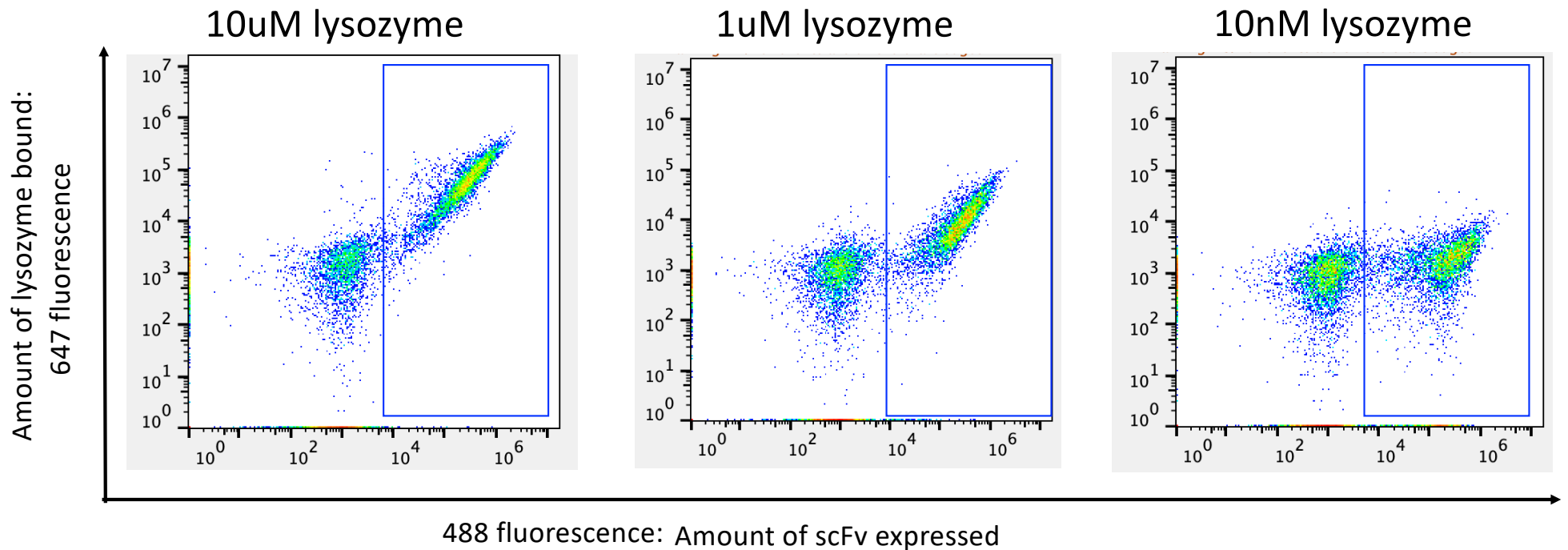


Tube #3

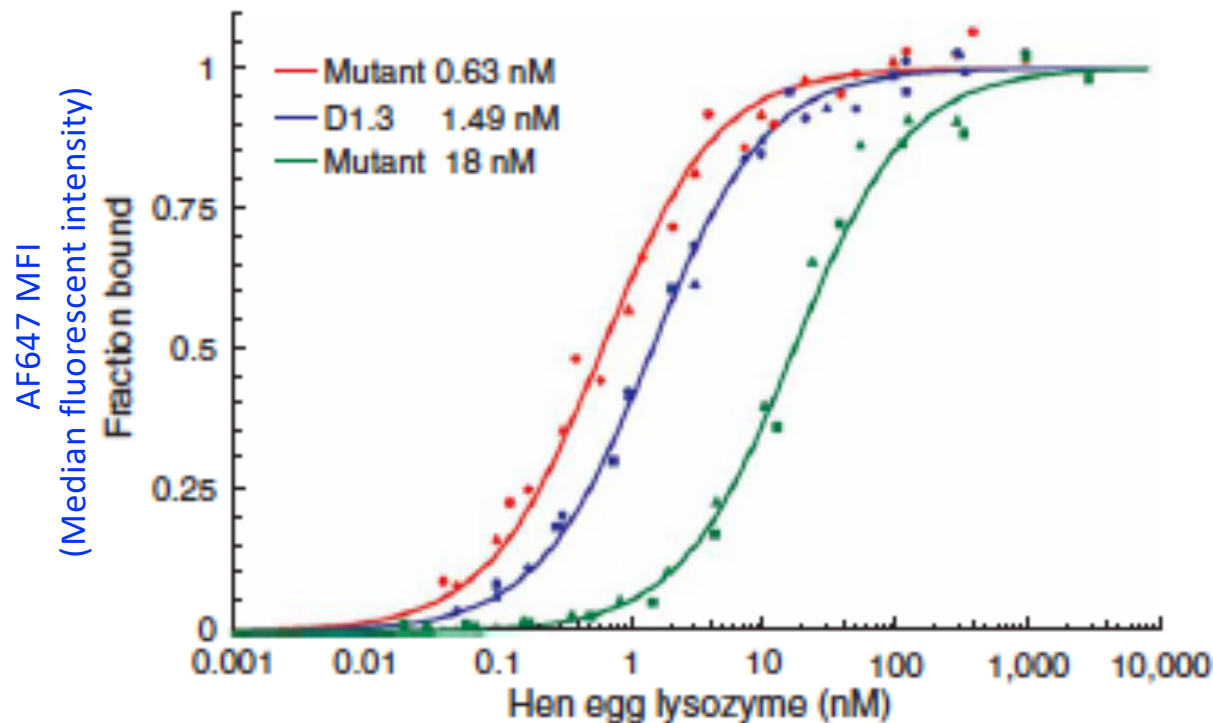


Median fluorescent intensity of gated scatterplot is to fraction bound

scFv Clone 14989 (650nM K_d) incubated with:



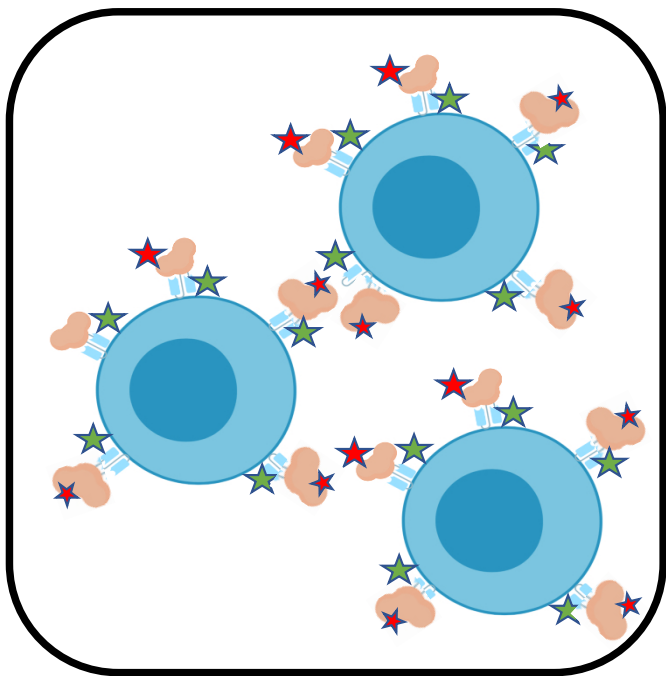
Plotted MFI illustrates fraction of antigen bound to antibody



Do you agree with the K_d ?

Today in “lab”

1) Set up titration of equilibrium binding reactions



2) Analyze flow cytometry data

