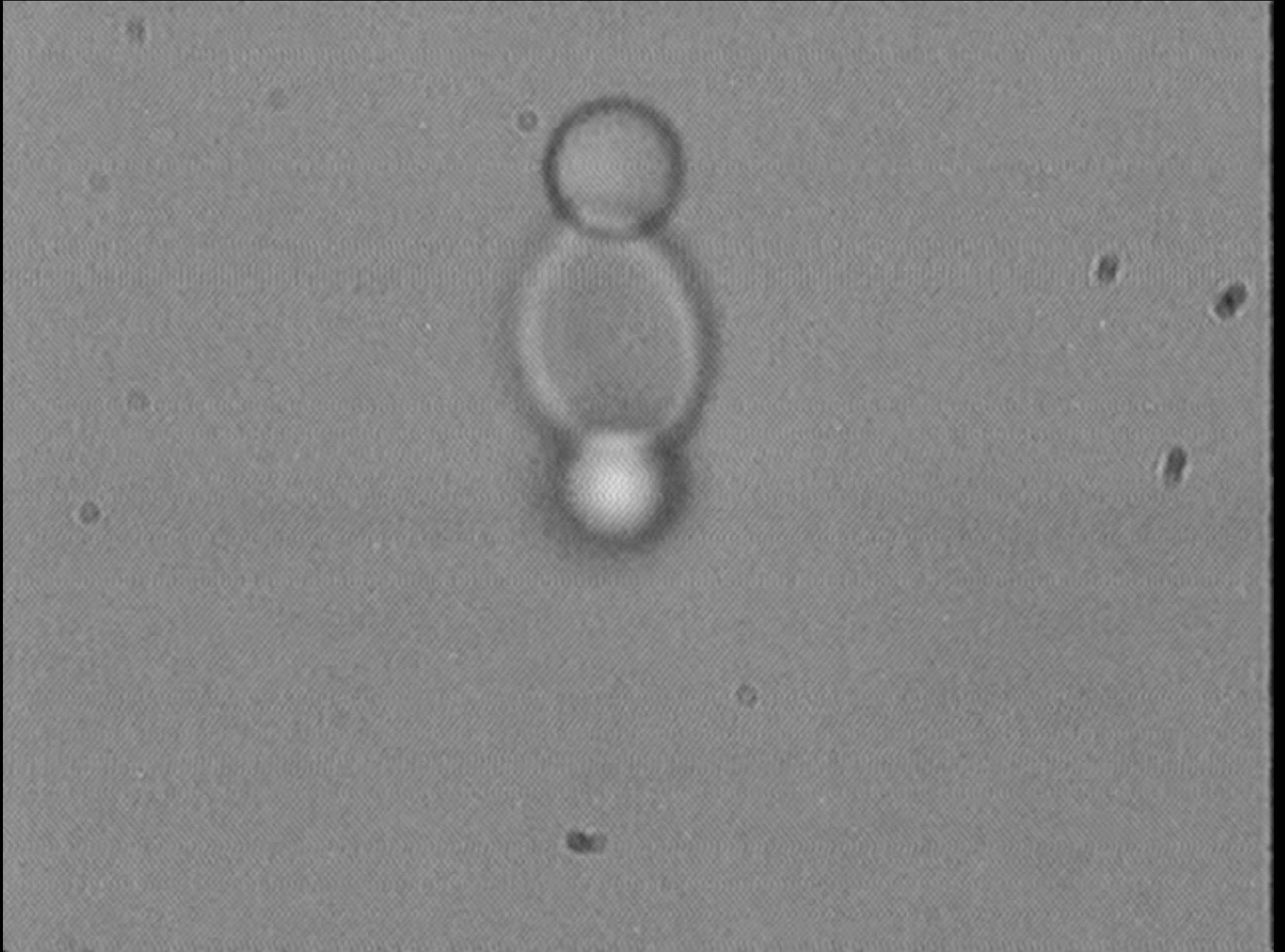


Cytoskeleton dynamics simulation of the red blood cell

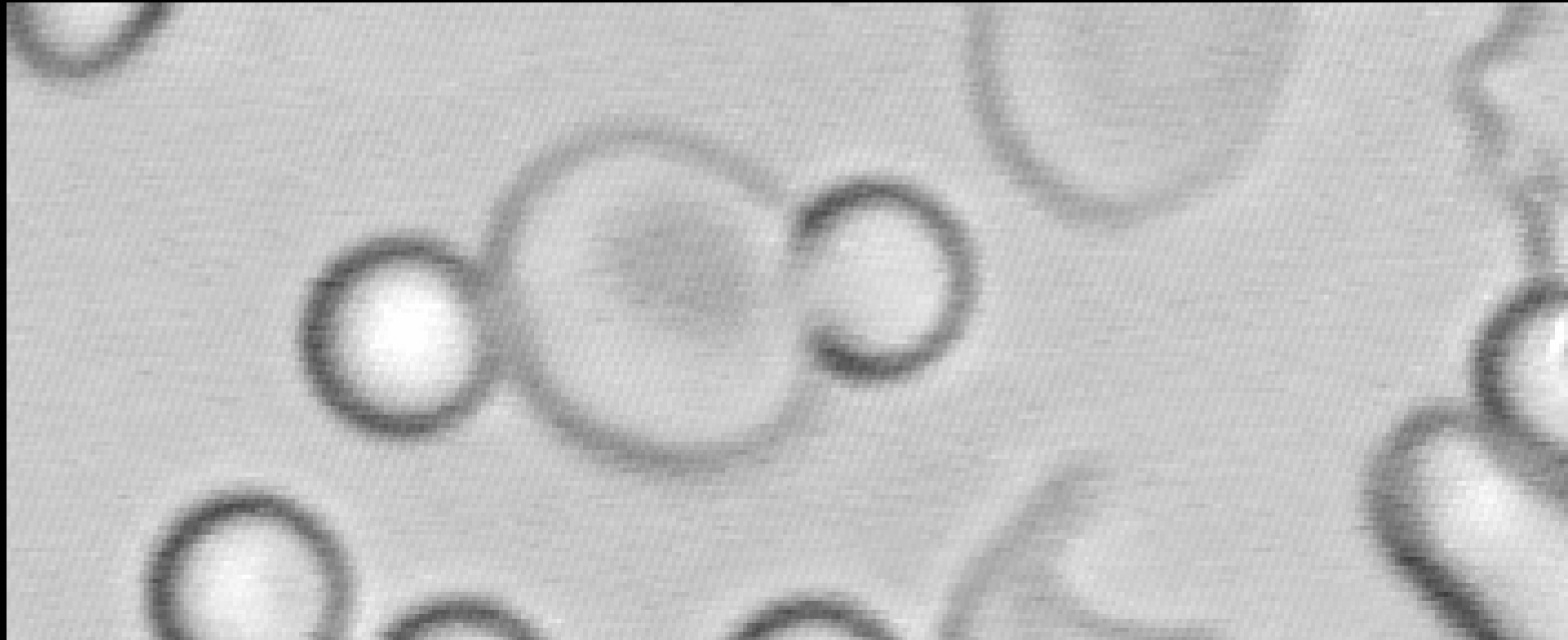
Ju Li

Collaborators: Subra Suresh, Ming Dao, George Lykotrafitis,
Chwee-Teck Lim

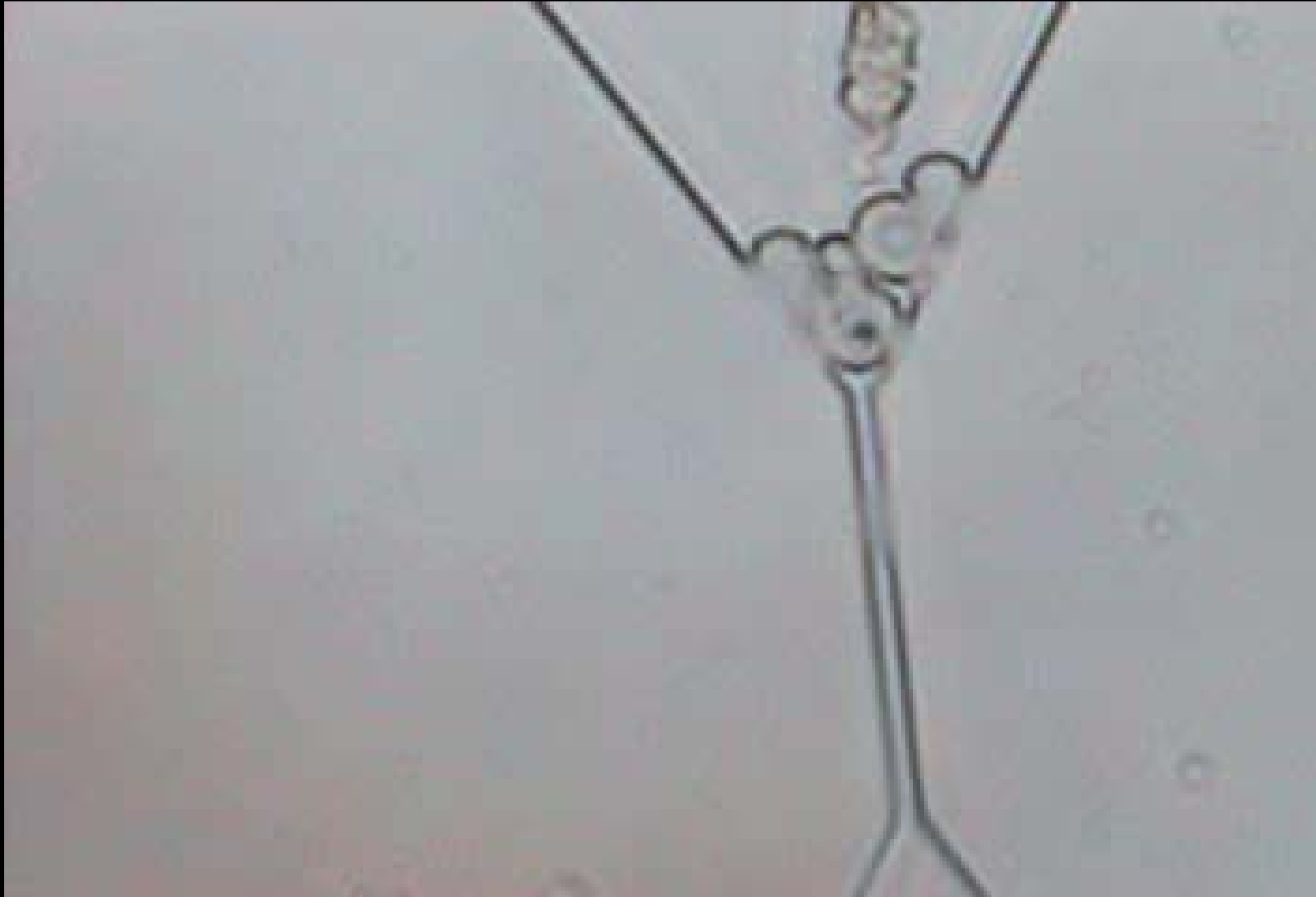
Optical tweezers stretching of healthy human red blood cell

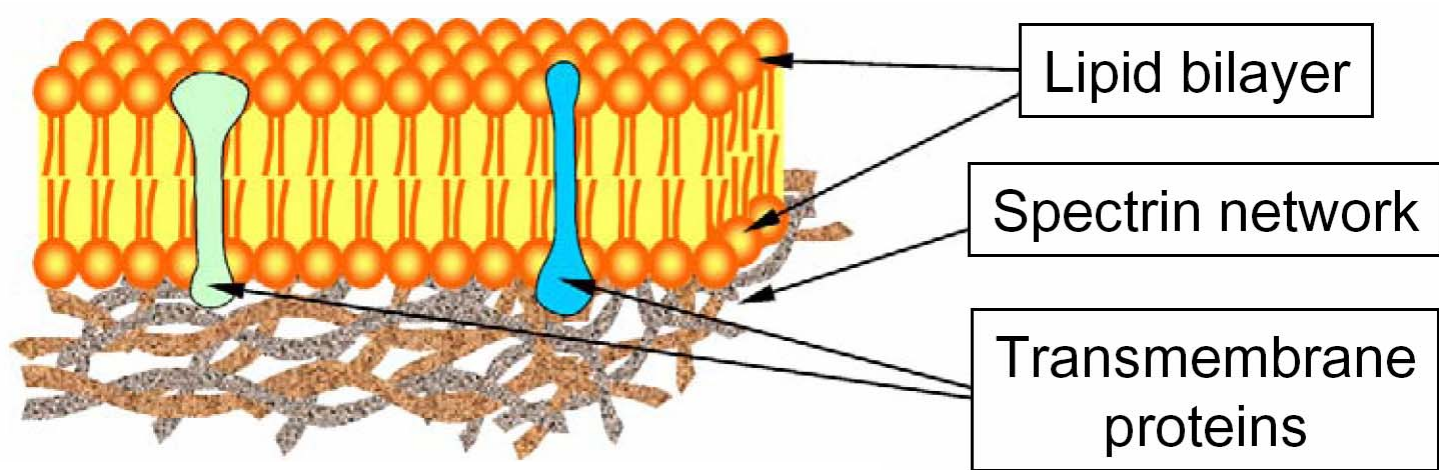


Malaria infected human red blood cell (schizont stage)



2 μ m microfluidic channel (amazing fluidity)





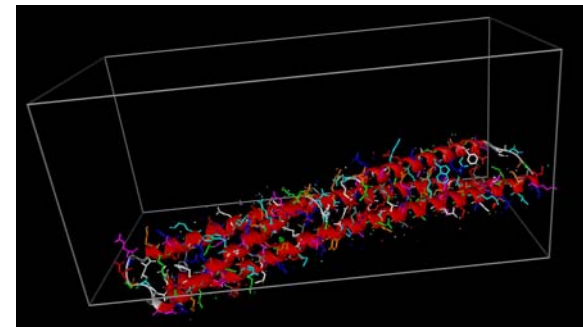
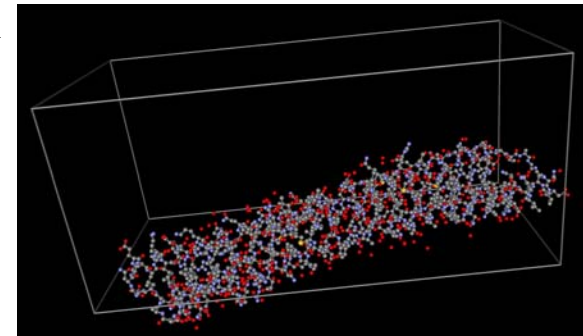
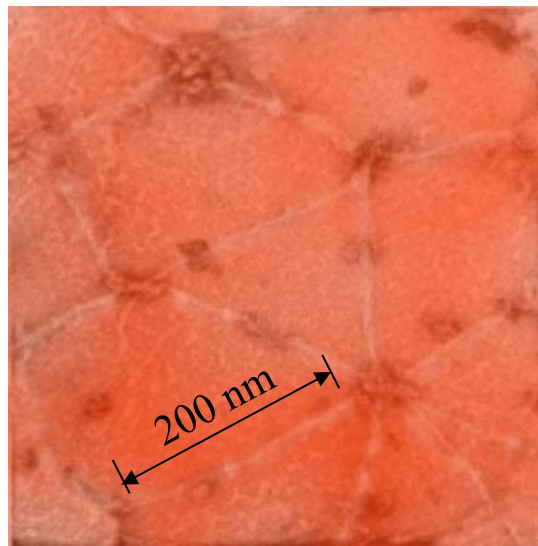
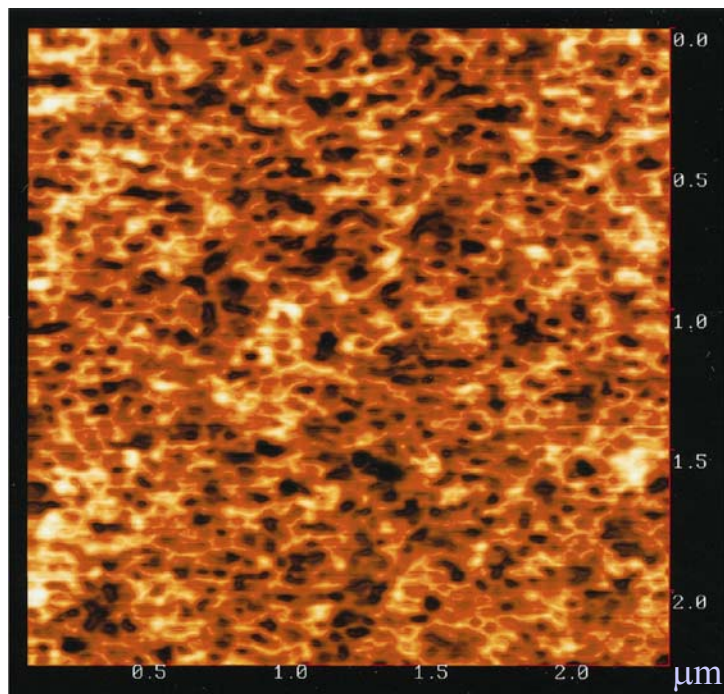
Red blood cell wall

Takeuchi *et al.*, *Biophys. J.* **74** (1998) 2171. One spectrin tetramer has 39 segments,

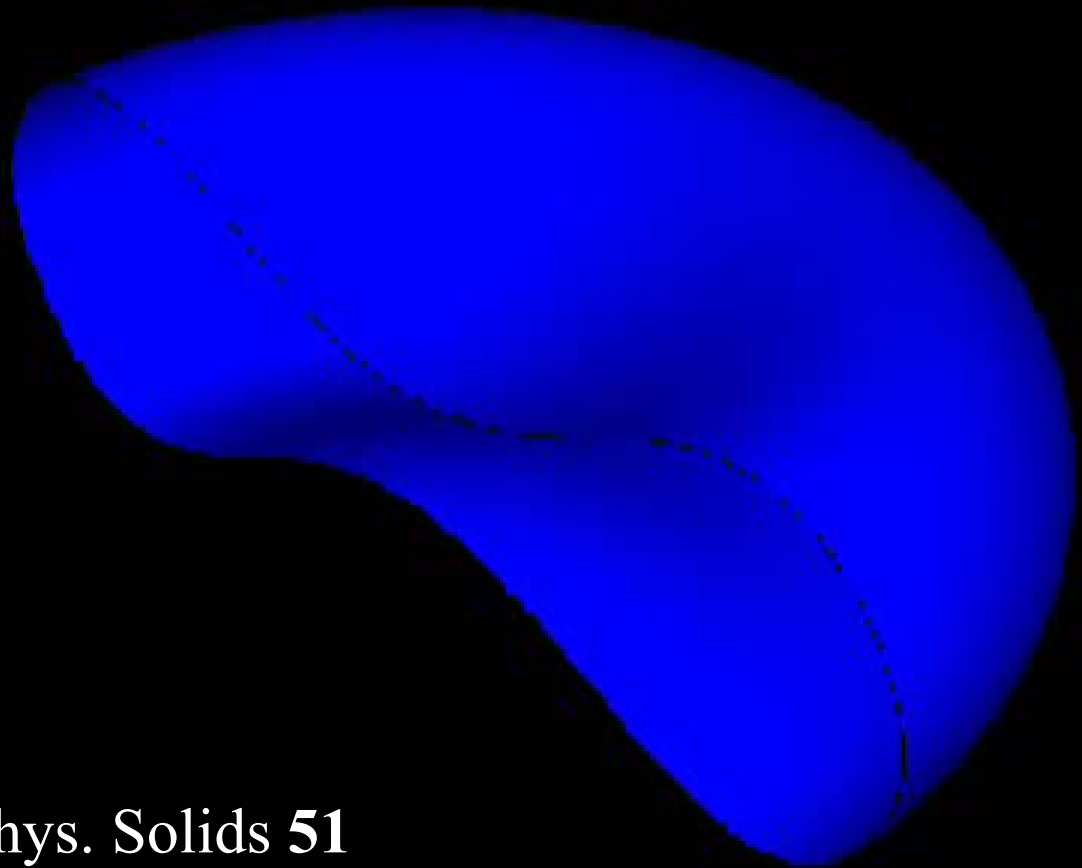
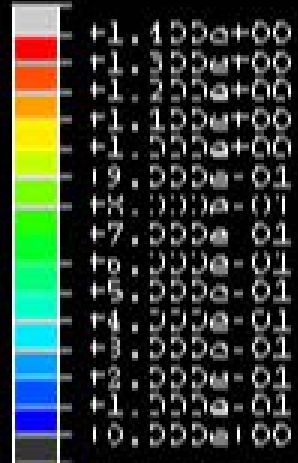
contour length ~ 200 nm.

Room-temperature length ~ 80 nm due to thermal fluctuations.

one segment ~ 5 nm

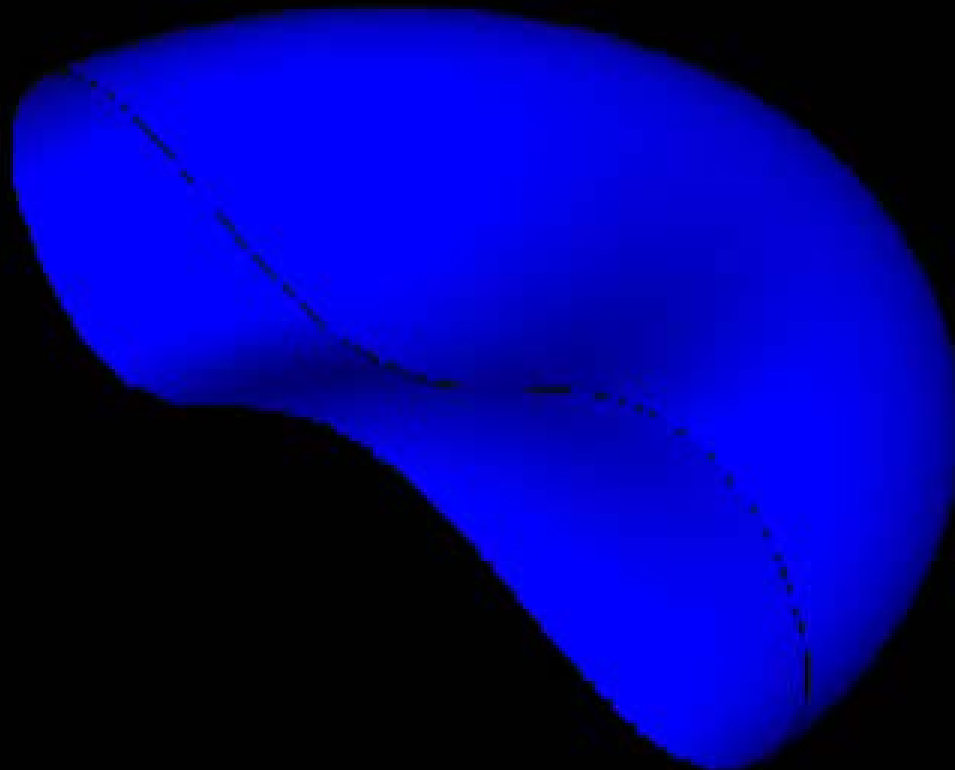
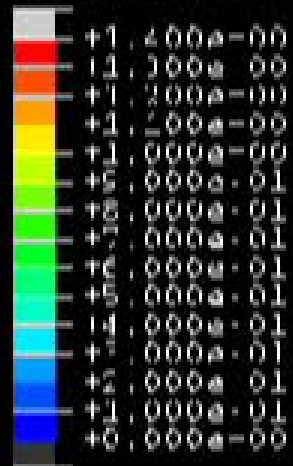


LE, Max. Principal
SNEC, (fraction = -1.5)
(Avg. Crit.: 75%)



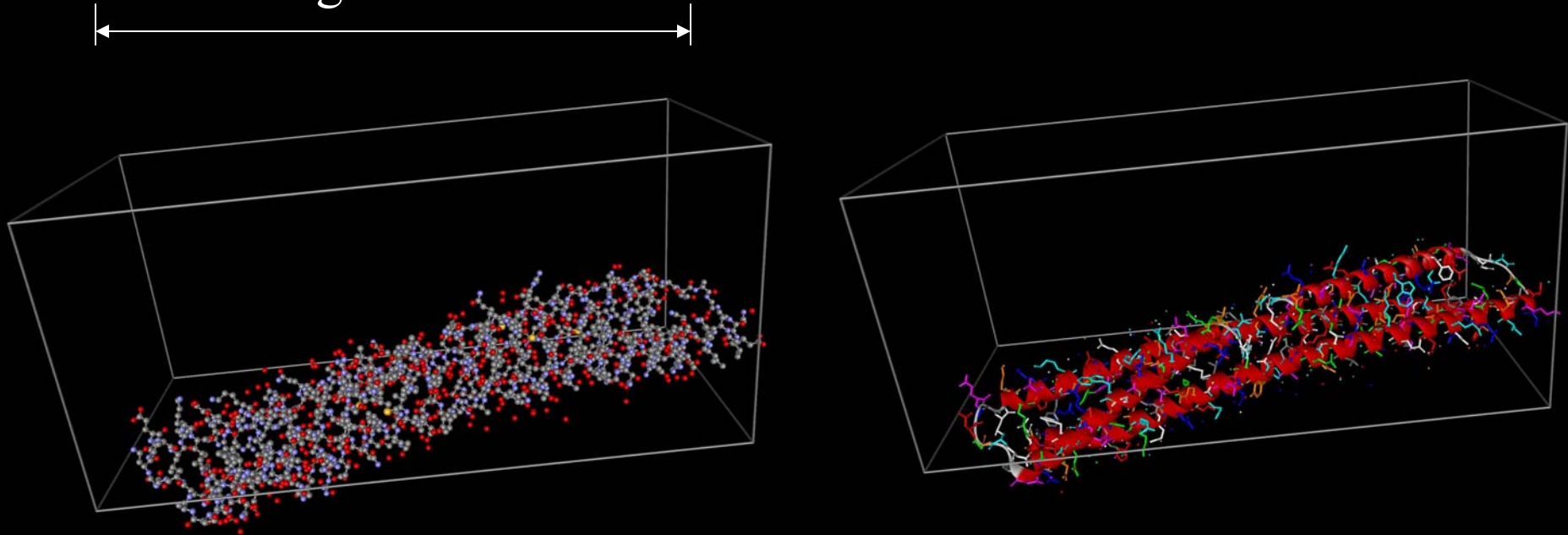
Dao *et al.*, J. Mech. Phys. Solids **51**
(2003) 2259; *ibid* **53** (2004) 493.

LE, Max. Principal
SPE3, (fraction = 1.0)
(Ave. Crit.: 75%)



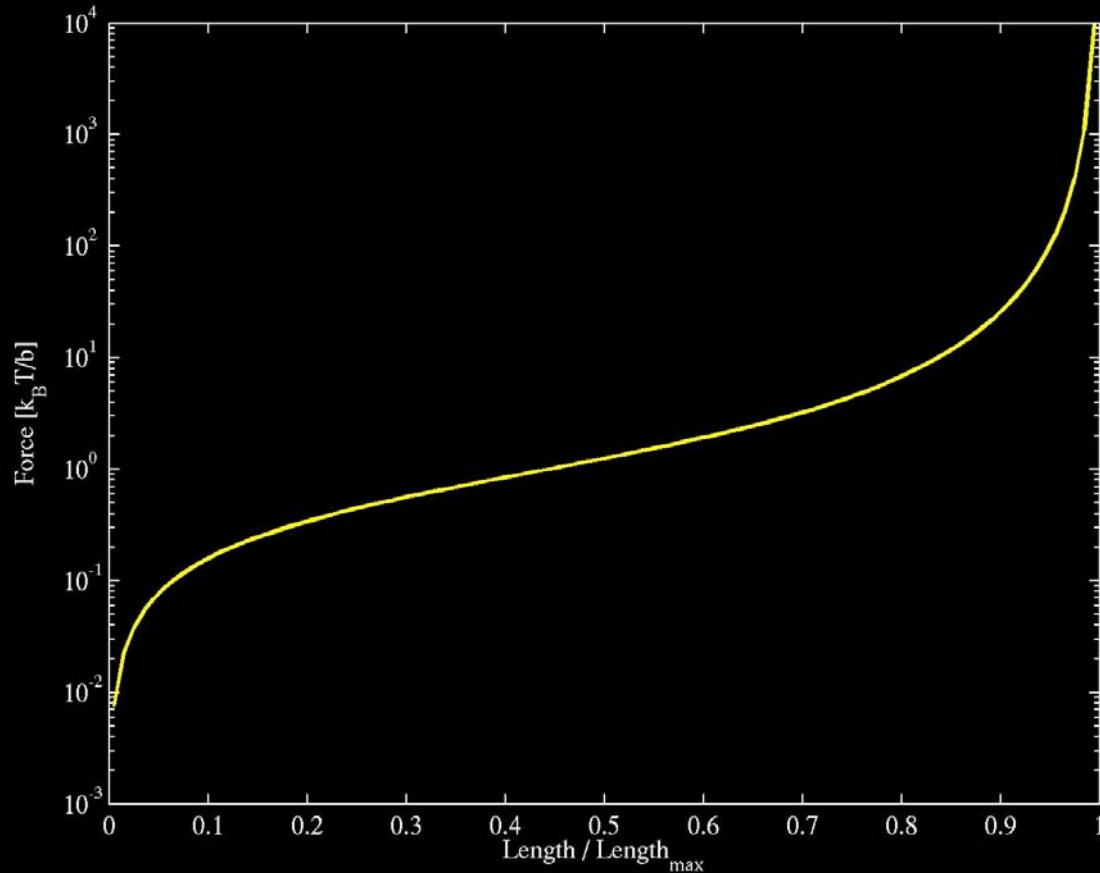
Spectrin Elasticity

one segment ~ 5 nm



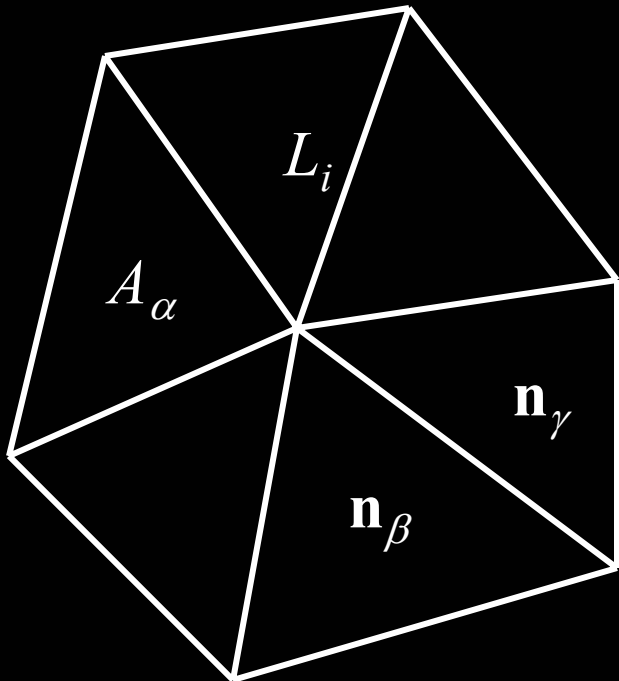
One spectrin tetramer has ~ 40 segments, contour length ~ 200 nm.
Room-temperature length ~ 80 nm due to thermal fluctuations.

Worm-like Chain Coarse-Grained Free Energy



$$V_{\text{WLC}}(L) = \frac{k_{\text{B}} T L_{\text{max}}}{4b} \cdot \frac{3x^2 - 2x^3}{1-x}, \quad x \equiv \frac{L}{L_{\text{max}}}$$

Spectrin-Net Level, Whole Red Blood Cell model (Discher, Boal, Boey, 1998)



$$V_{\text{total}} = \sum_{i \in \text{spectrin link}} V_{\text{WLC}}(L_i) +$$

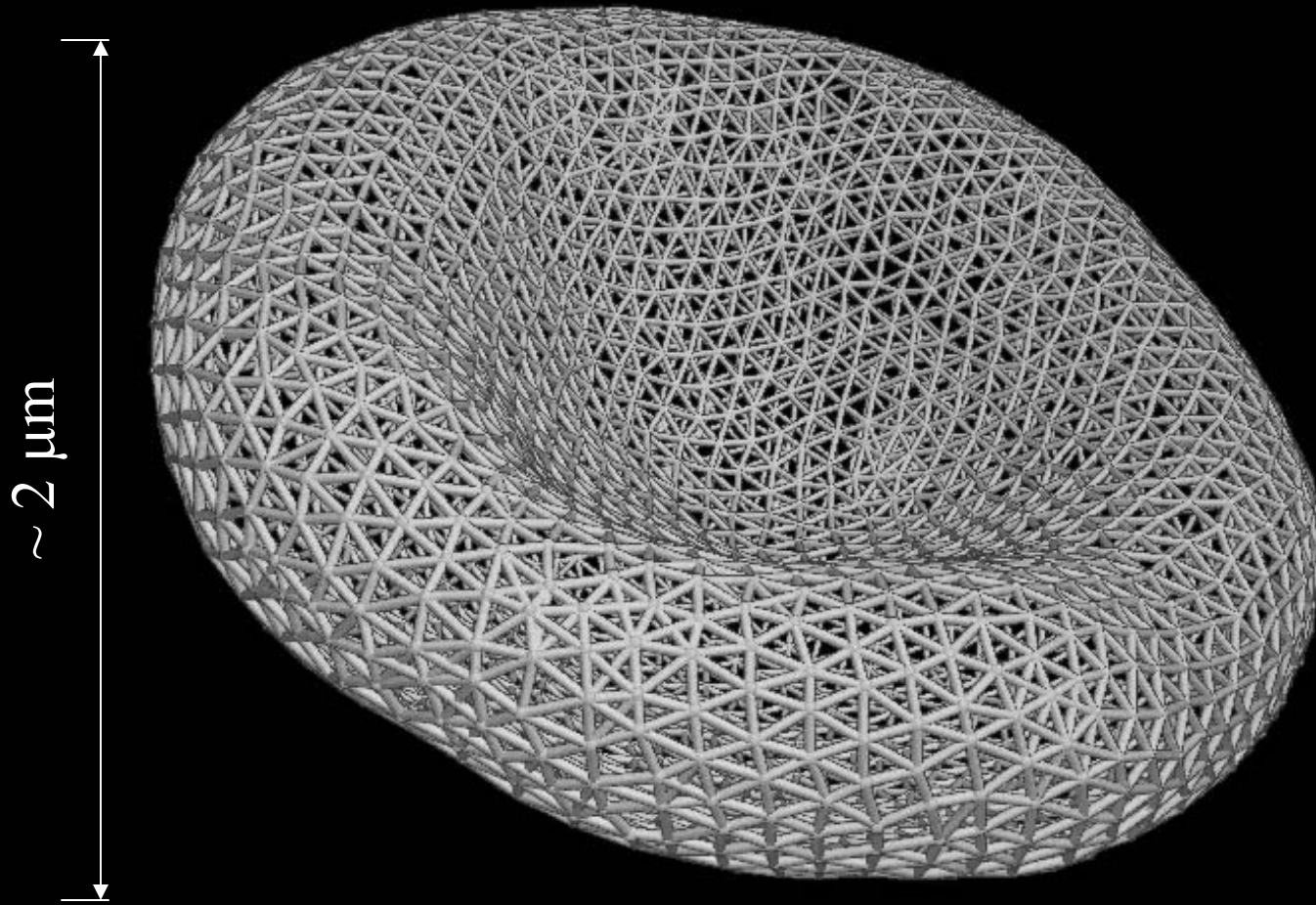
$$\sum_{\alpha \in \text{triangle}} \frac{C}{A_\alpha} +$$

$$\sum_{\beta, \gamma \in \text{triangle}} K_{\text{bend}} (1 - \mathbf{n}_\beta \cdot \mathbf{n}_\gamma)$$

+ total volume constraint + total area constraint

Small Cell Simulation

(“volume quench” to get discocyte shape)



2562 vertices

Lim, Wortis,
Mukhopadhyay,
PNAS **99** (2002)
16766

Stomatocyte -
discocyte -
echinocyte
Sequence

spontaneous
curvature
parameter

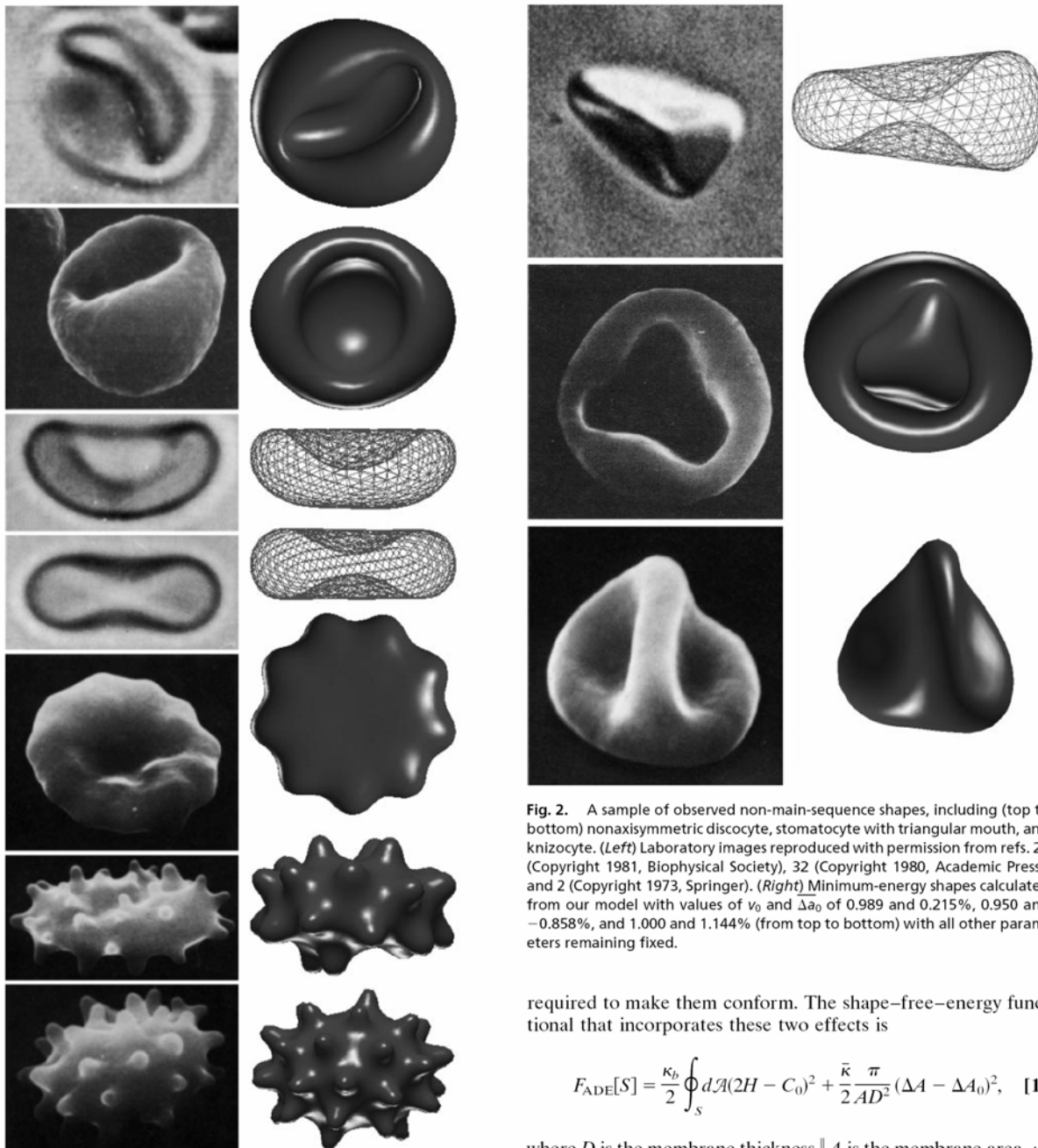


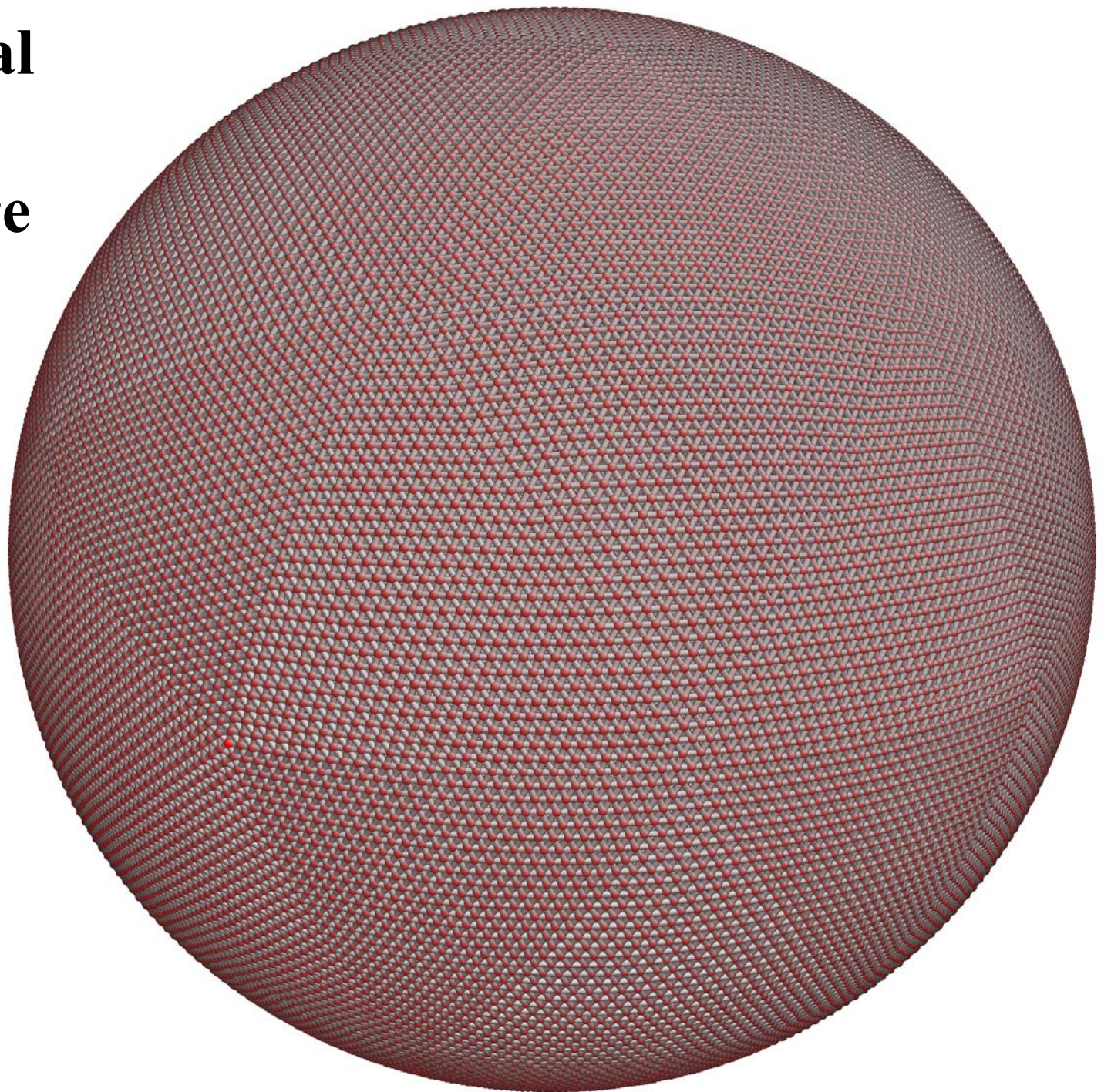
Fig. 2. A sample of observed non-main-sequence shapes, including (top to bottom) nonaxisymmetric discocyte, stomatocyte with triangular mouth, and knizocyte. (Left) Laboratory images reproduced with permission from refs. 27 (Copyright 1981, Biophysical Society), 32 (Copyright 1980, Academic Press), and 2 (Copyright 1973, Springer). (Right) Minimum-energy shapes calculated from our model with values of v_0 and Δa_0 of 0.989 and 0.215%, 0.950 and -0.858%, and 1.000 and 1.144% (from top to bottom) with all other parameters remaining fixed.

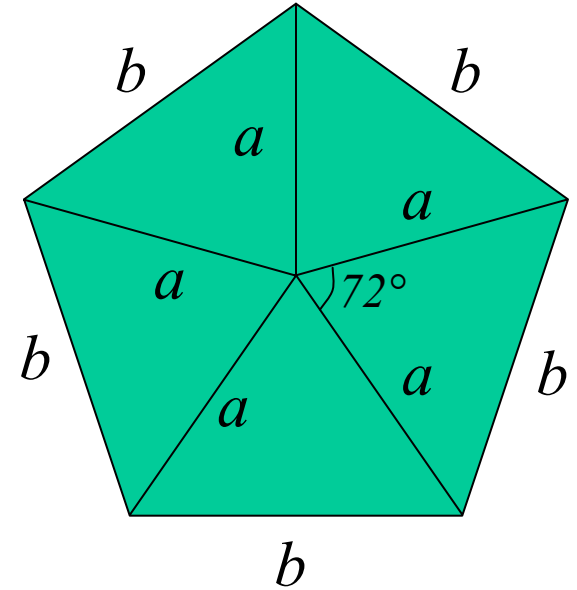
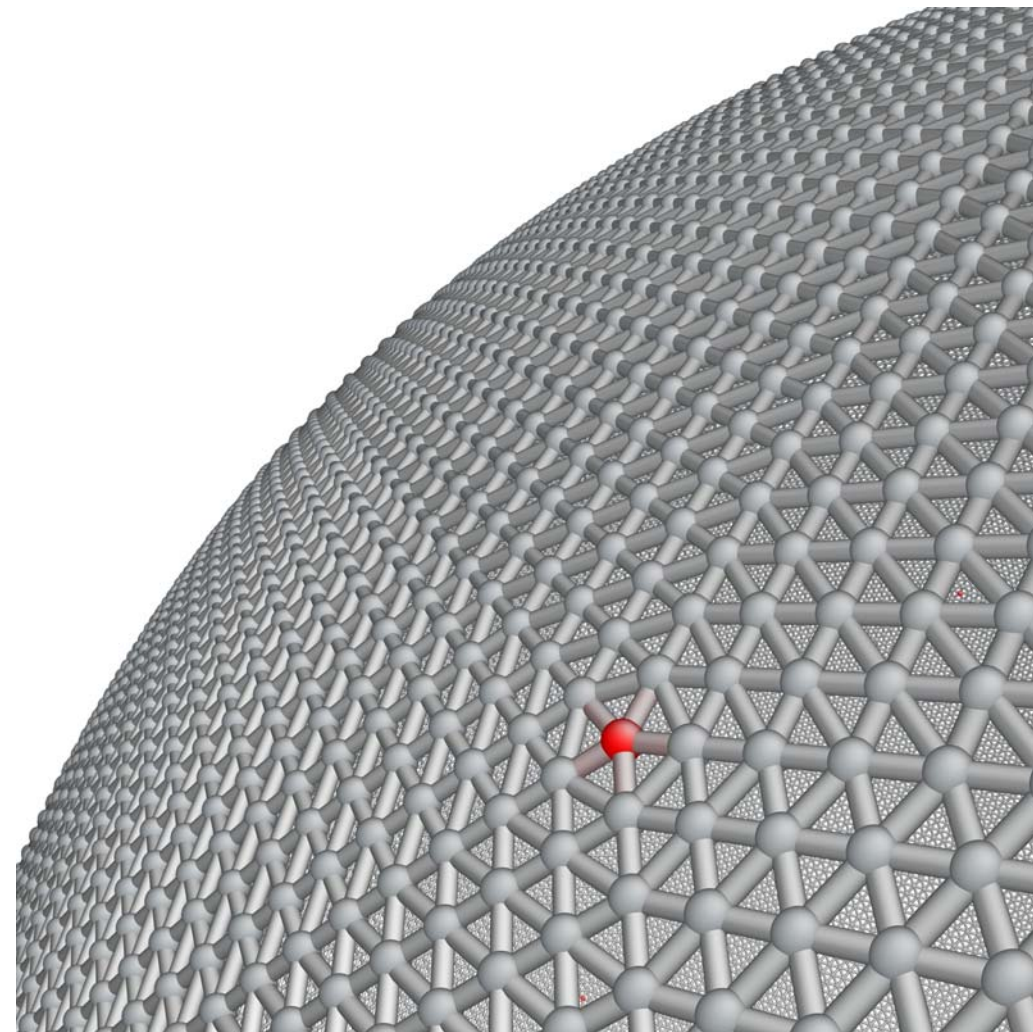
required to make them conform. The shape-free-energy functional that incorporates these two effects is

$$F_{ADE}[S] = \frac{\kappa_b}{2} \oint_S dA (2H - C_0)^2 + \frac{\bar{\kappa}}{2AD^2} \pi (\Delta A - \Delta A_0)^2, \quad [1]$$

where D is the membrane thickness, A is the membrane area, κ_b

Icosahedral network on a sphere

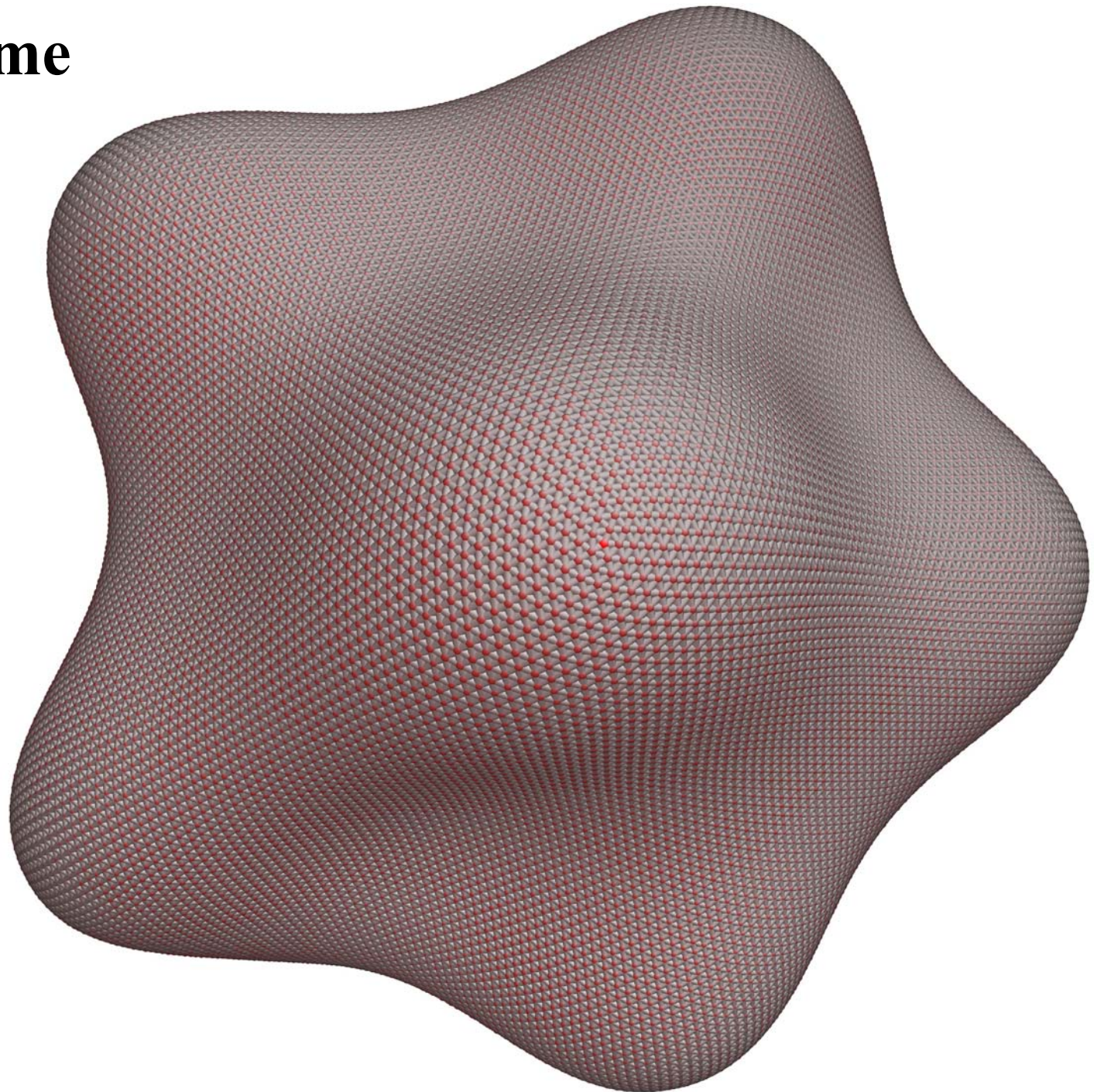




Geometrically Necessary Disinclinations

If each carries disinclination charge 60° , need 12

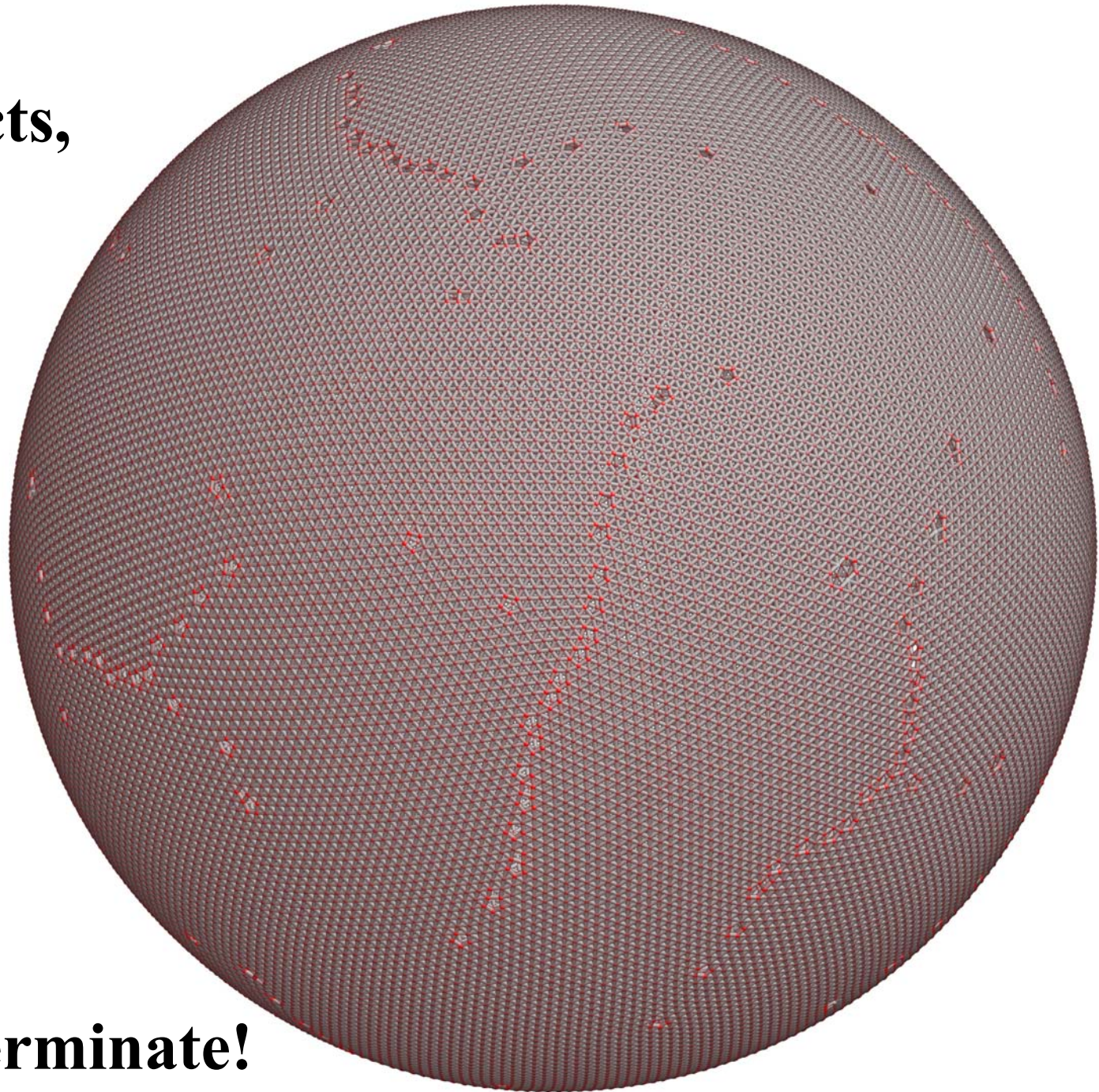
100% volume



60% volume

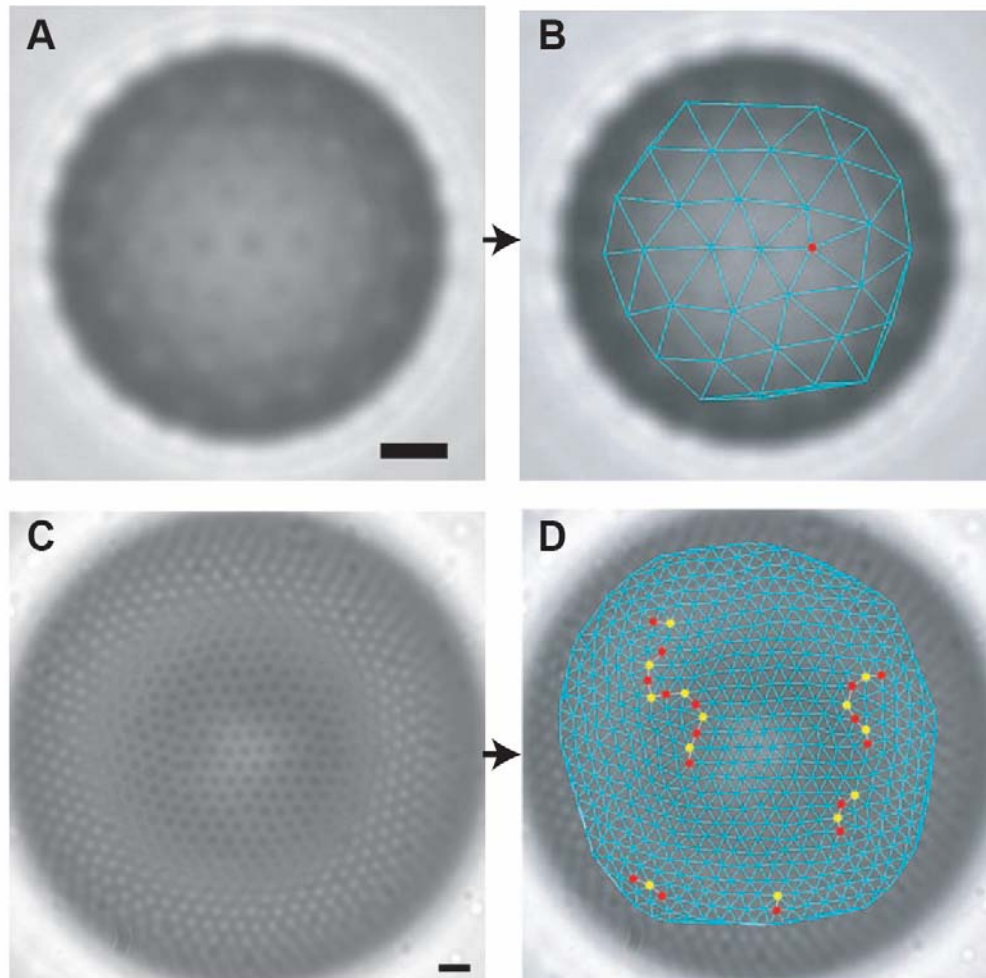


**To rid of the
shape artifacts,
melt and
quench the
network**



GBs freely terminate!

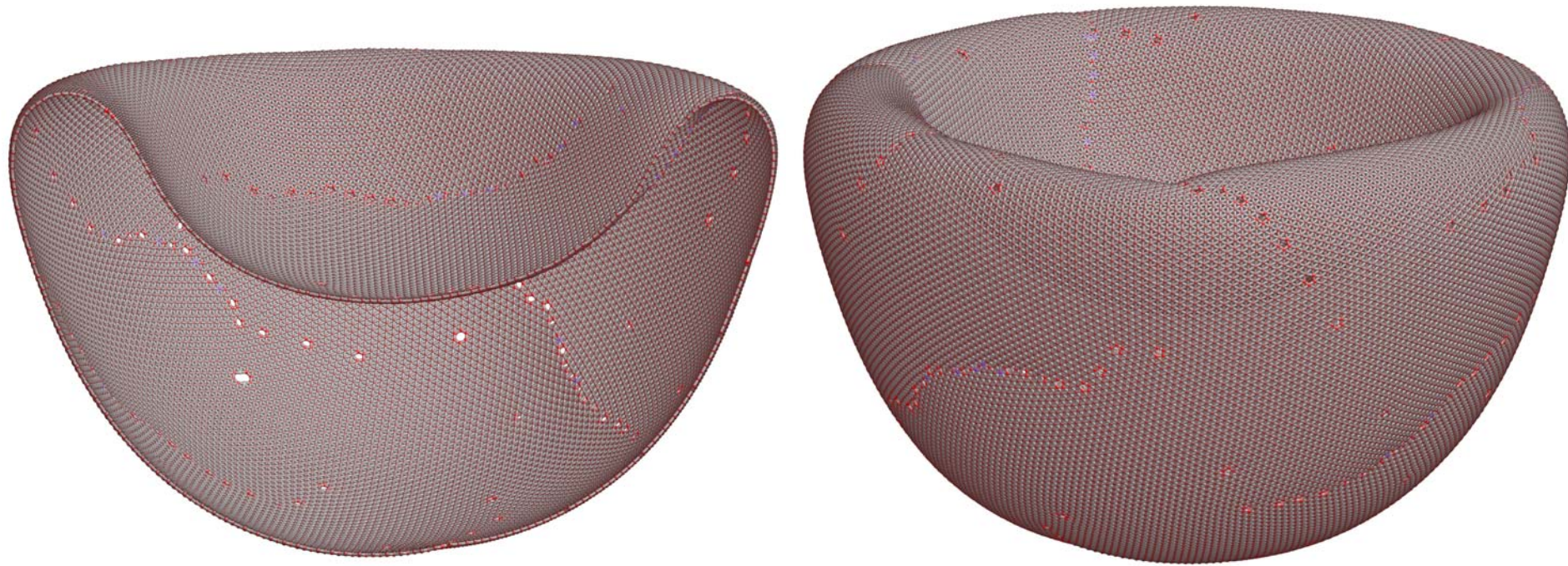
Fig. 1. Light microscope images of particle-coated droplets. Two droplets (A) and (C) are shown, together with their associated defect structures (B) and (D). (A) An $\sim 13\%$ portion of a small spherical droplet with radius $R = 12.0 \mu\text{m}$ and mean particle spacing $a = 2.9 \mu\text{m}$ ($R/a = 4.2$), along with the associated triangulation (B). Charge $+1(-1)$ disclinations are shown in red and yellow, respectively. Only one $+1$ disclination is seen. (C) A cap of spherical colloidal crystal on a water droplet of radius $R = 43.9 \mu\text{m}$ with mean particle spacing $a = 3.1 \mu\text{m}$ ($R/a = 14.3$), along with the associated triangulation (D). In this case the imaged crystal covers $\sim 17\%$ of the surface area of the sphere. Bars [(A) and (C)], $5 \mu\text{m}$.



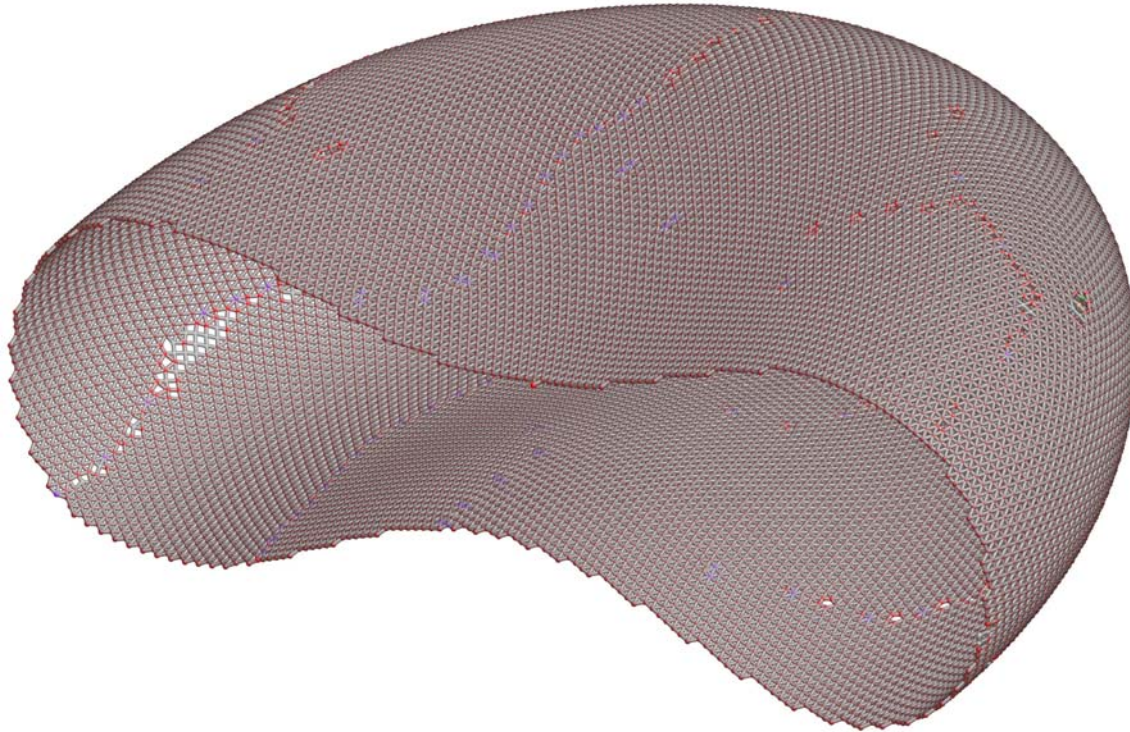
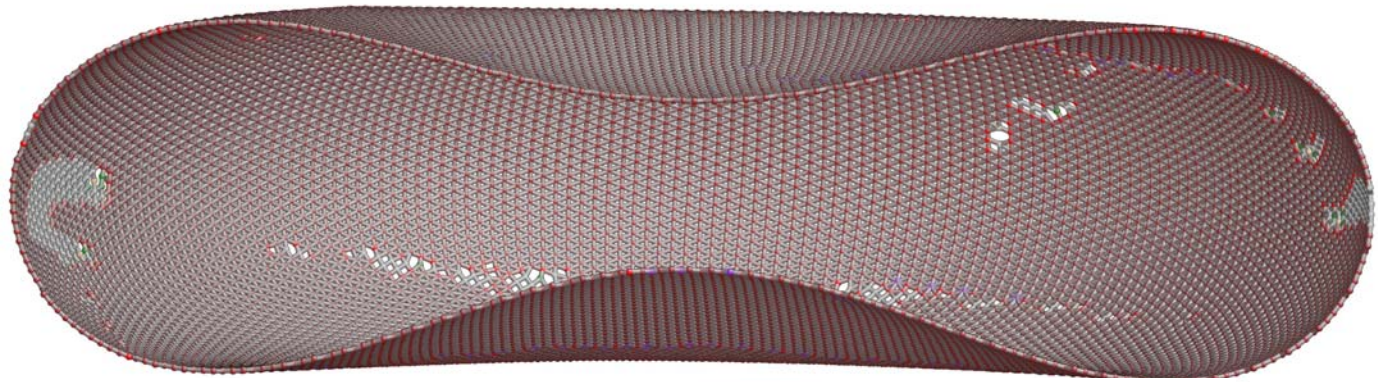
Bausch *et al.*,
Science **299**
 (2003) 1716.

These GBs should be widespread in nature: large viral protein capsids, giant spherical fullerenes, spherical bacterial surface layers, siliceous skeletons of spherical radiolaria (aulosphaera), etc. Sites for chemical reactions, initiation points for bacterial cell division, influence the mechanical response.

Material reference state for the in-plane shear energy E_{shear}



60% volume: **spherical state** as stress-free reference.



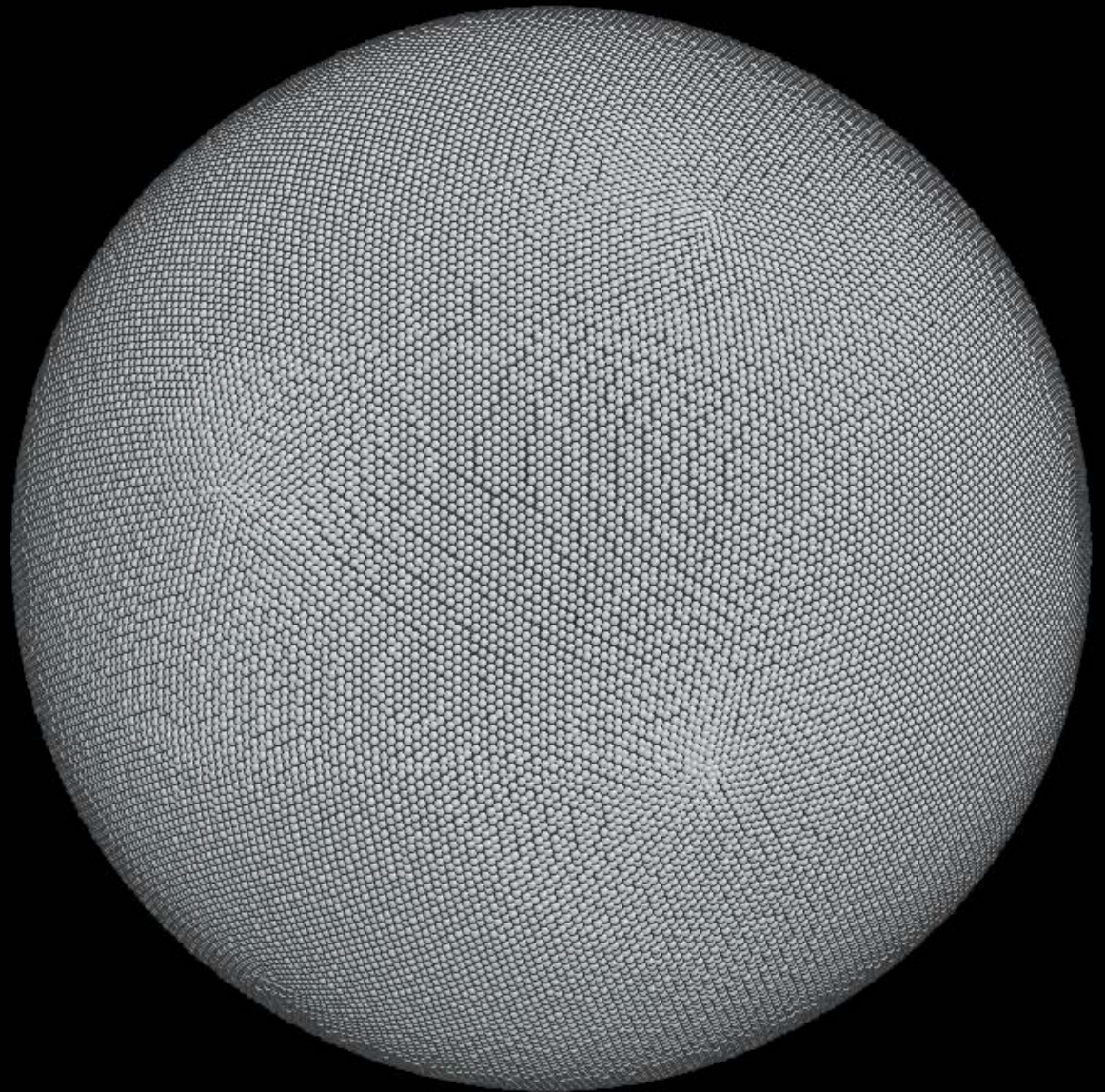
W/ experimental range of parameters and sphere as stress-free reference state, the biconcave shape is only metastable at 60% volume.

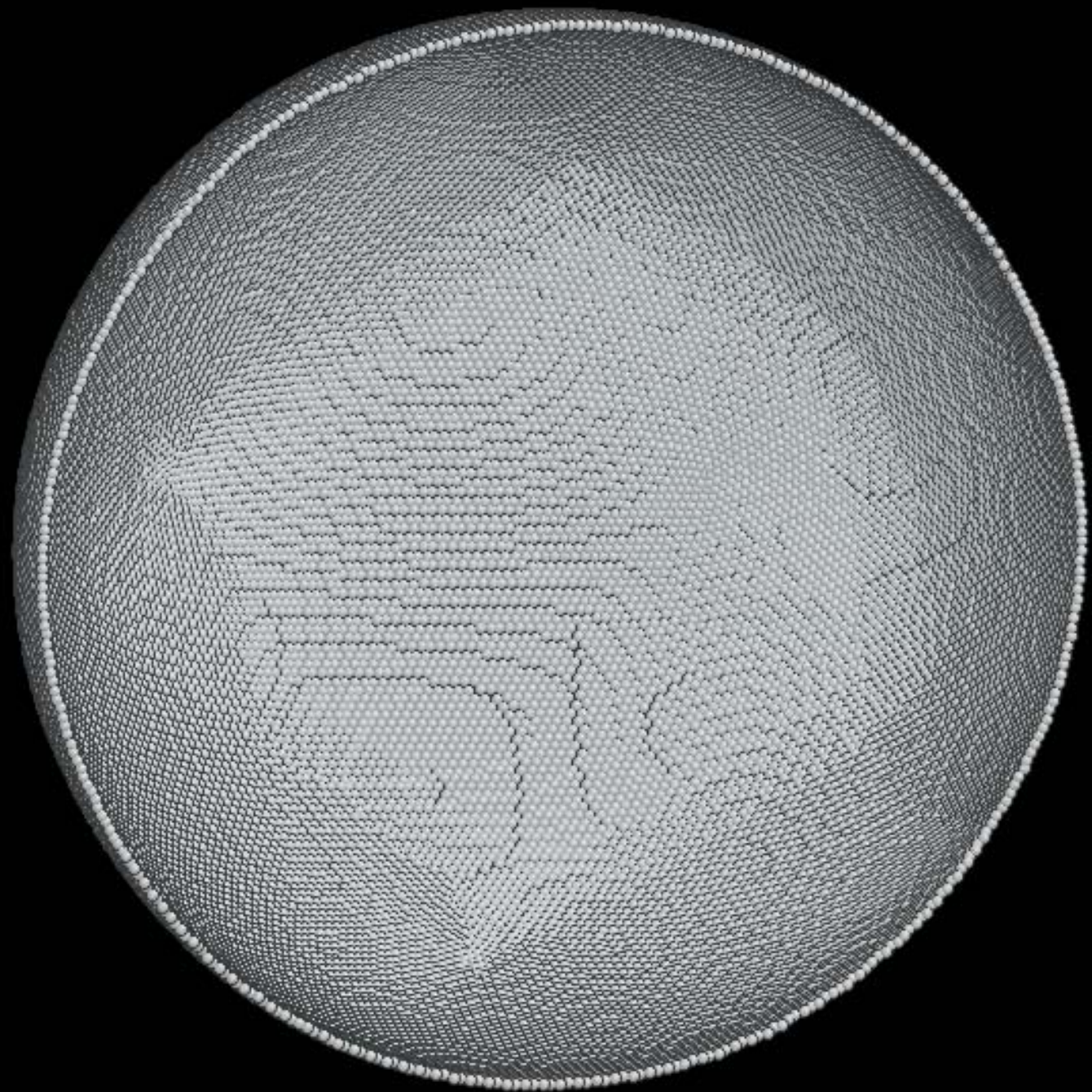
**With bending
energy**

E_{bend} only

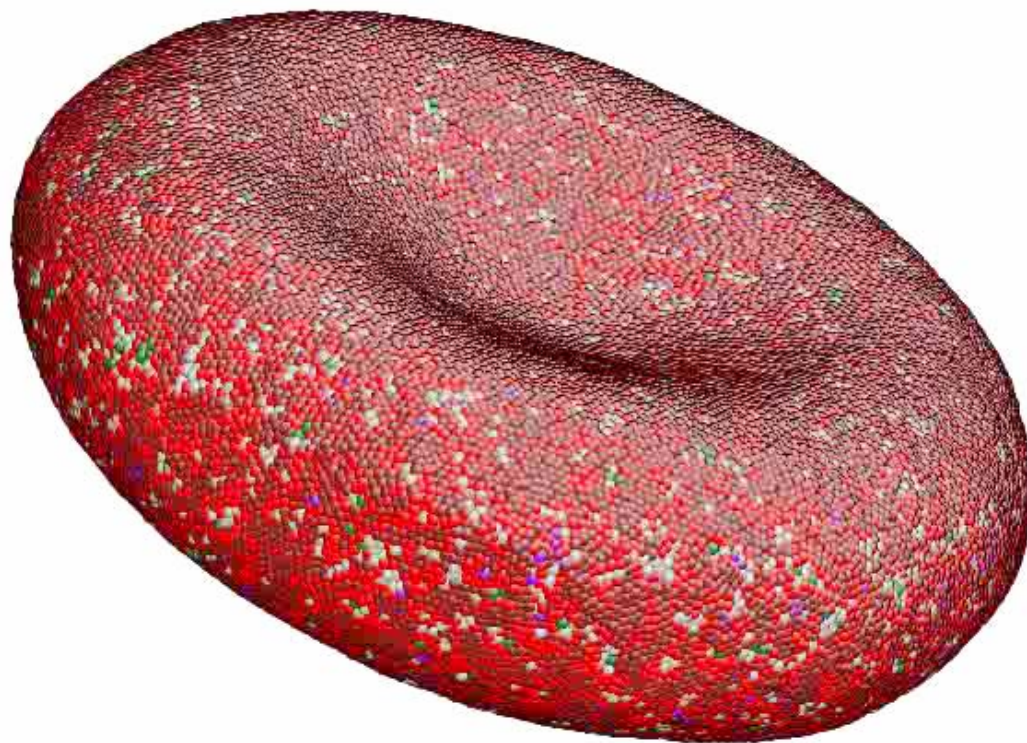
Canham (1970)

Helfrich (1973)

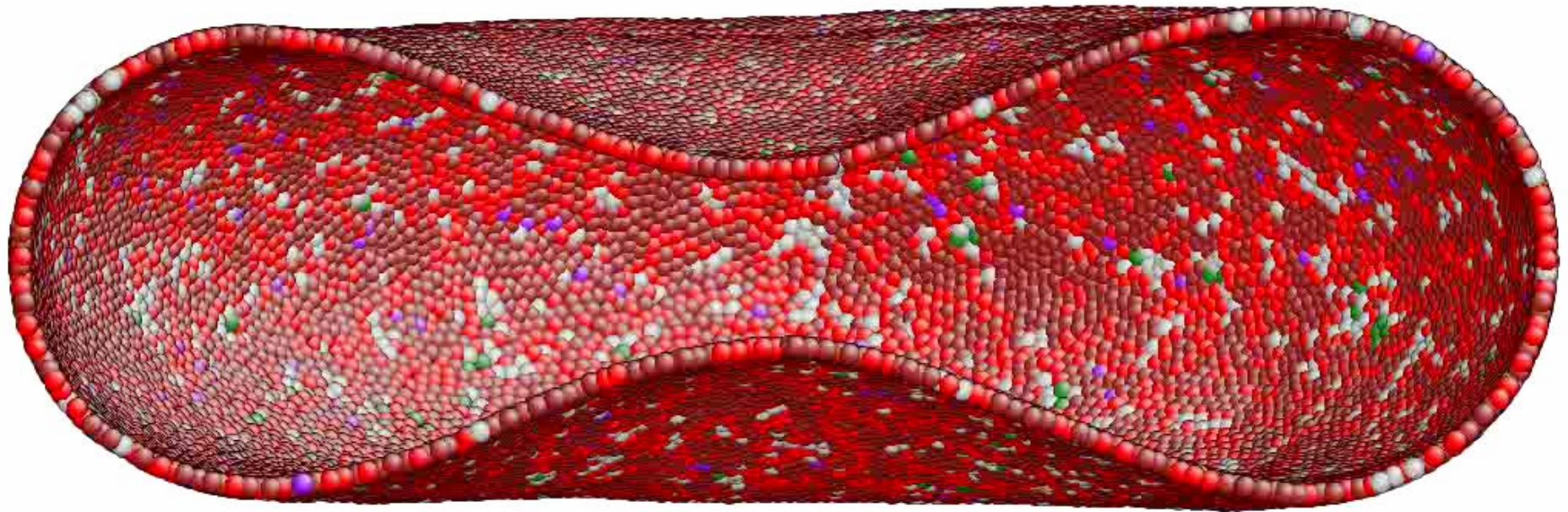


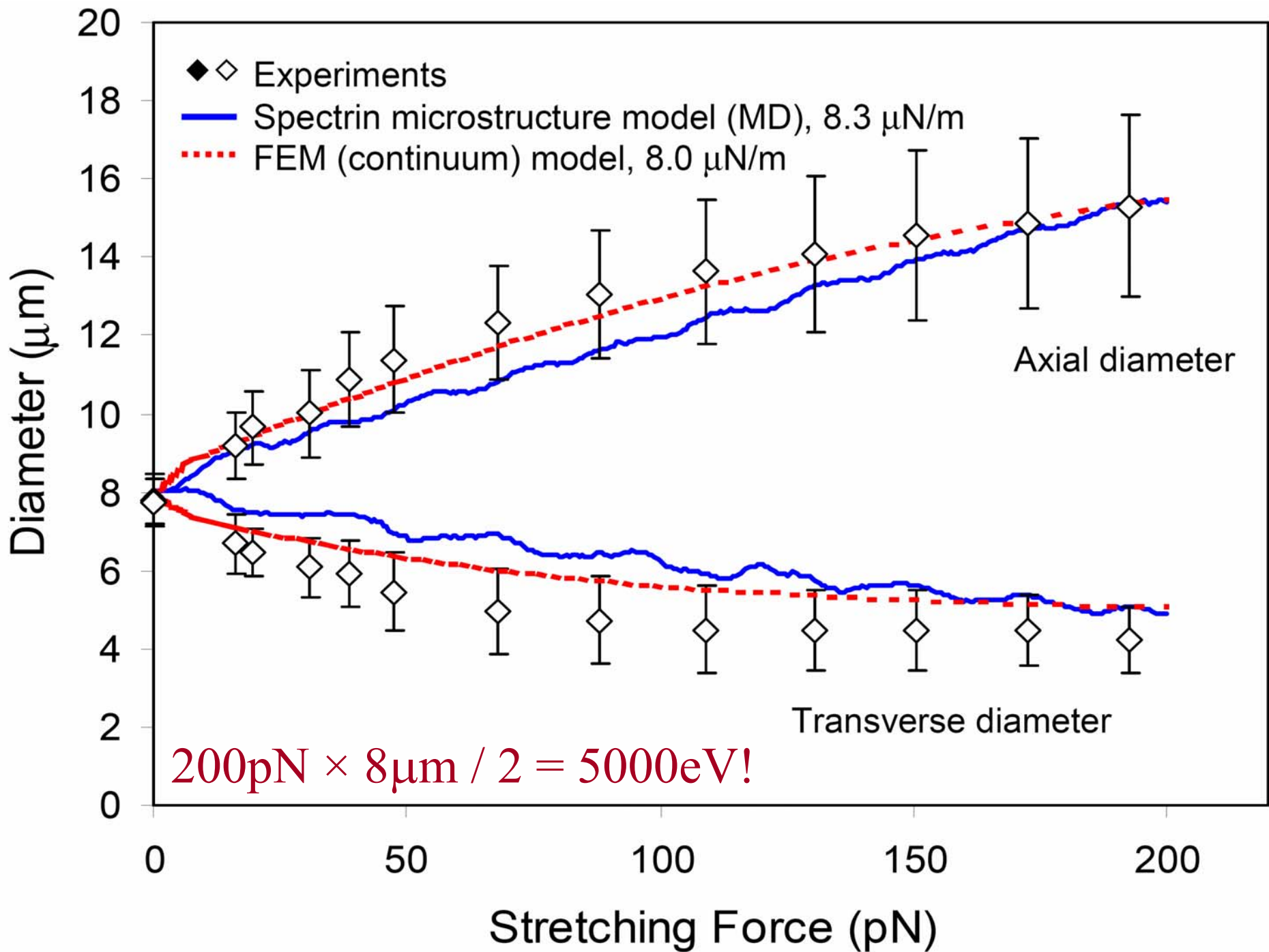


Optical Tweezers Stretching Simulation

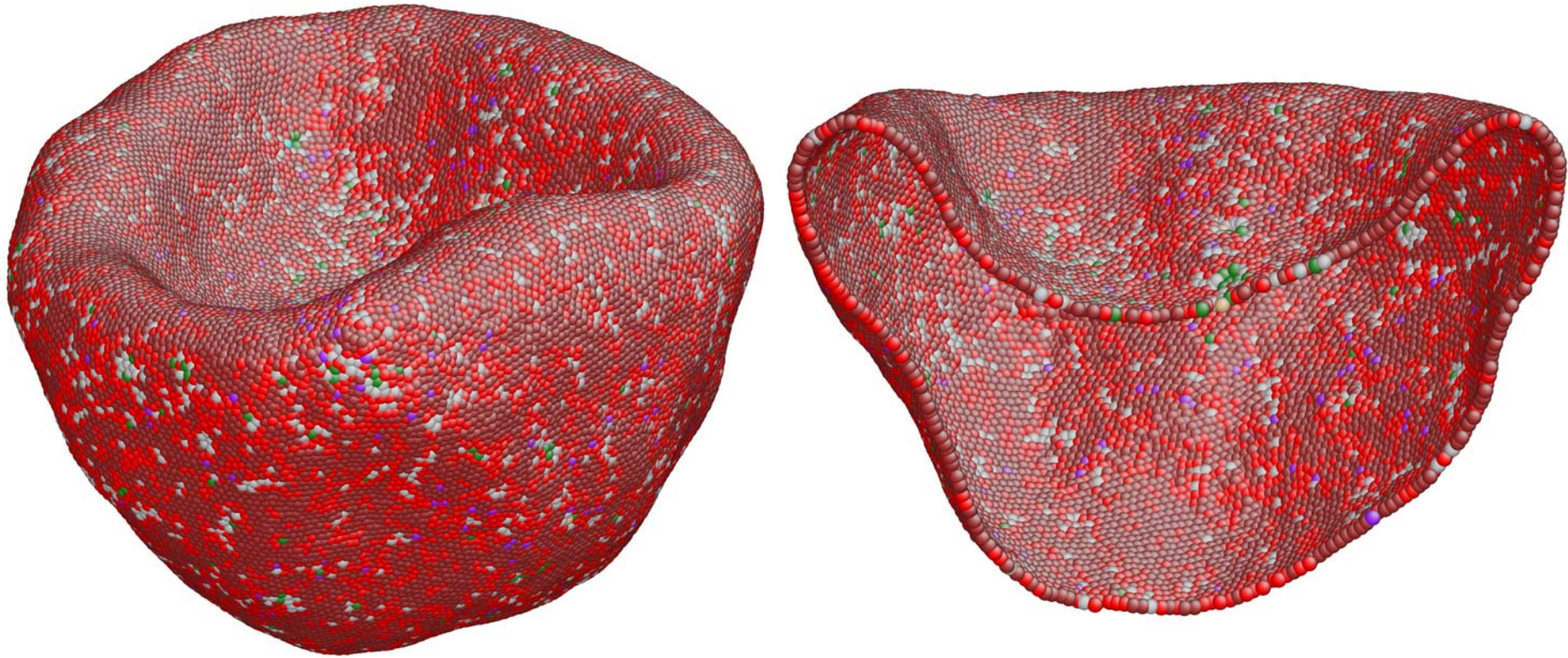


Cross Sectional View





Why is biconcave the stable equilibrium shape?



$$E_{\text{bend}} \sim 8\pi\kappa: \kappa \sim 2 \times 10^{-19} \text{ J} \rightarrow E_{\text{bend}} \sim 30 \text{ eV}$$

$$E_{\text{shear}} \sim \mu\varepsilon^2 A: \mu \sim 8\mu\text{N/m}, \varepsilon \sim 0.1, A \sim 140\mu\text{m}^2 \\ \rightarrow E_{\text{shear}} \sim 70 \text{ eV}$$

Material Concept Hypothesis

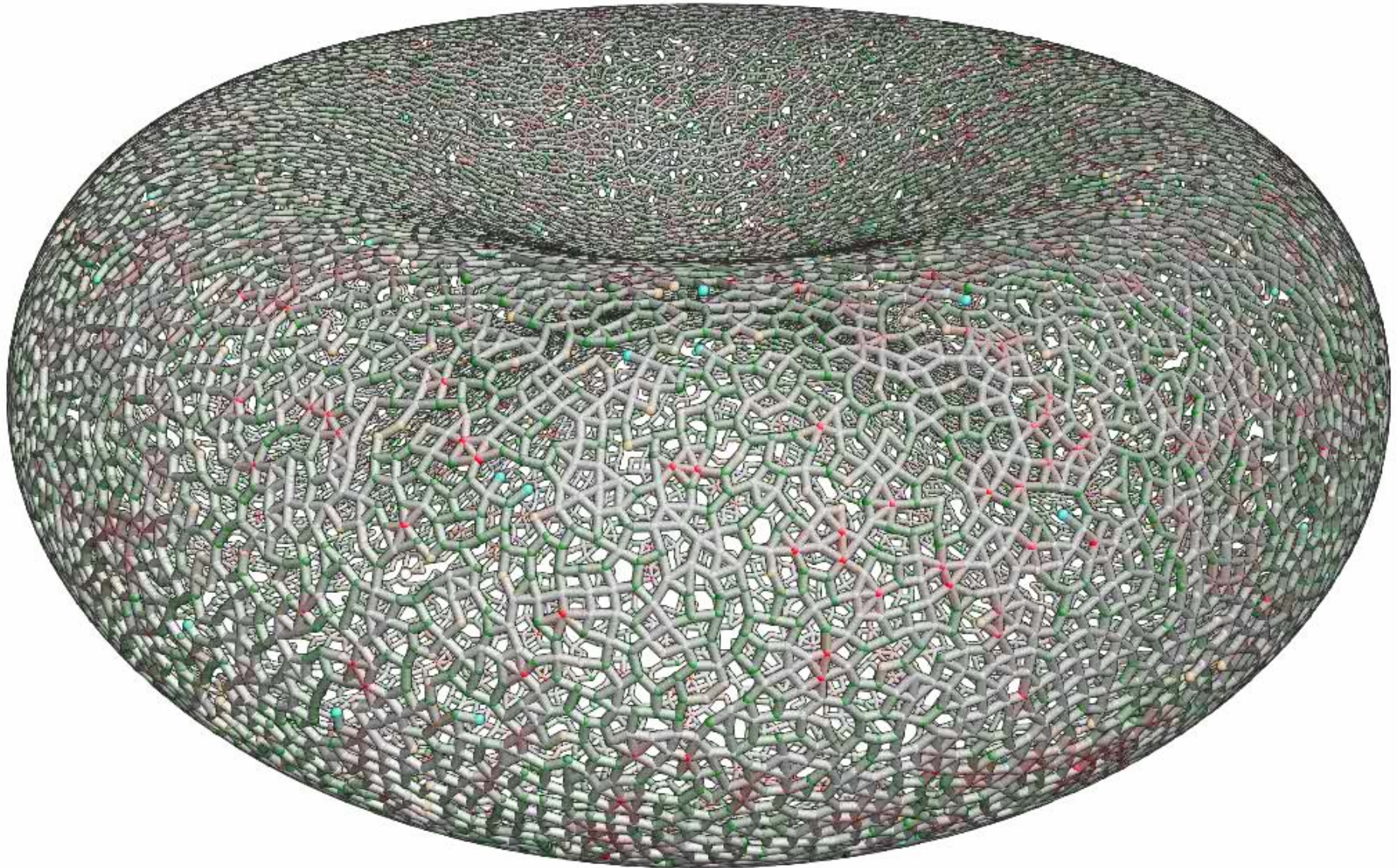
Li, Dao, Lim & Suresh, *Biophys. J.* **88** (2005) 3707.

- In an ideal limit, for any RBC shape, the cytoskeleton will always undergo remodeling in topological connectivity at a slow rate to relax its in-plane *shear* elastic energy to *zero*.

“liquefaction”, “slow-flowing glass”

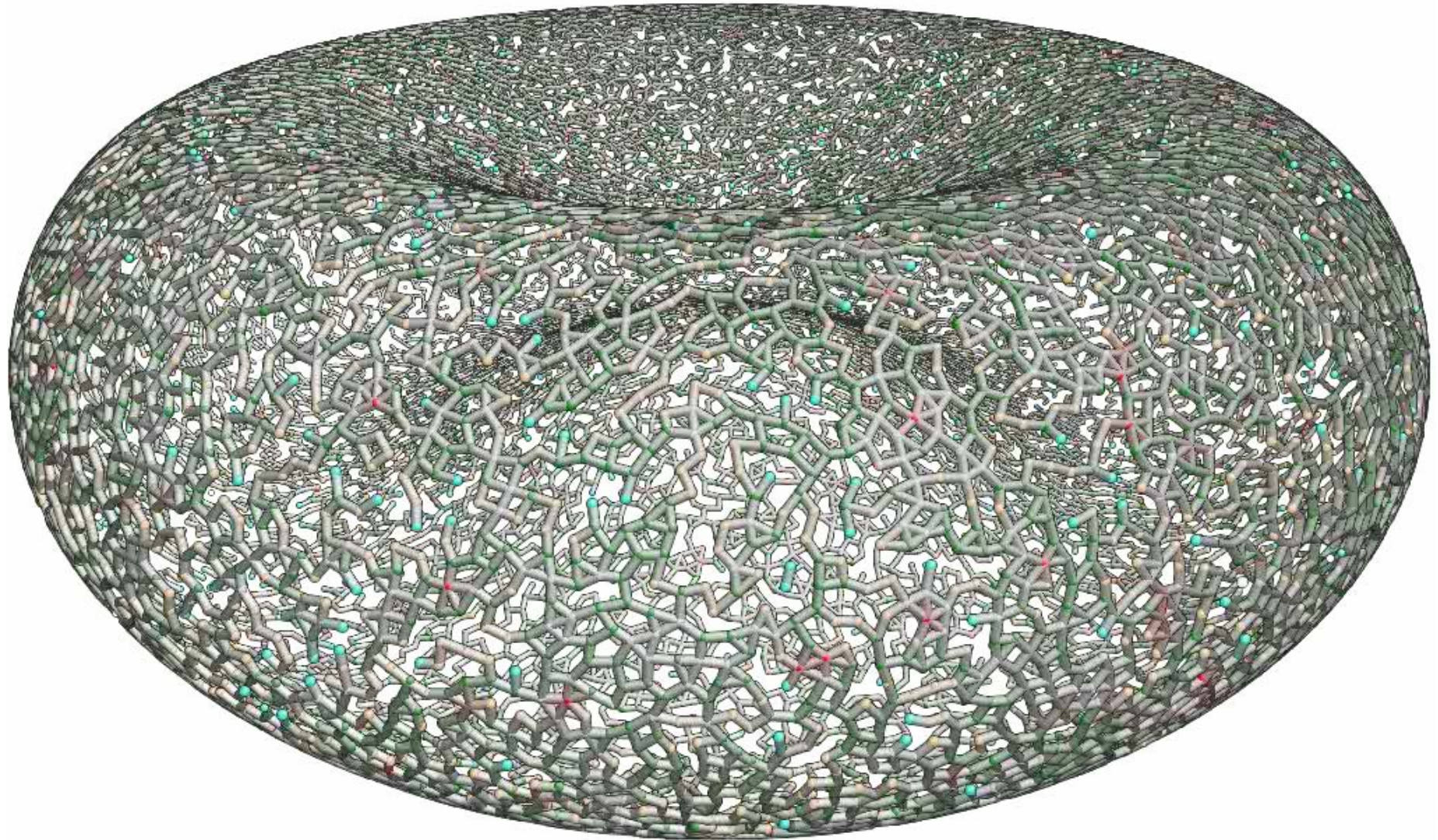
- At the timescale of optical tweezers stretching, the above relaxation is not significant, so large shear energy can be injected temporarily.

Stillinger-Weber liquid on curved surface:



no shear energy can survive long!

RBC cytoskeleton at reduced spectrin density



very large holes start to percolate ...

Extreme Statistics of Cytoskeletal Defects in RBC

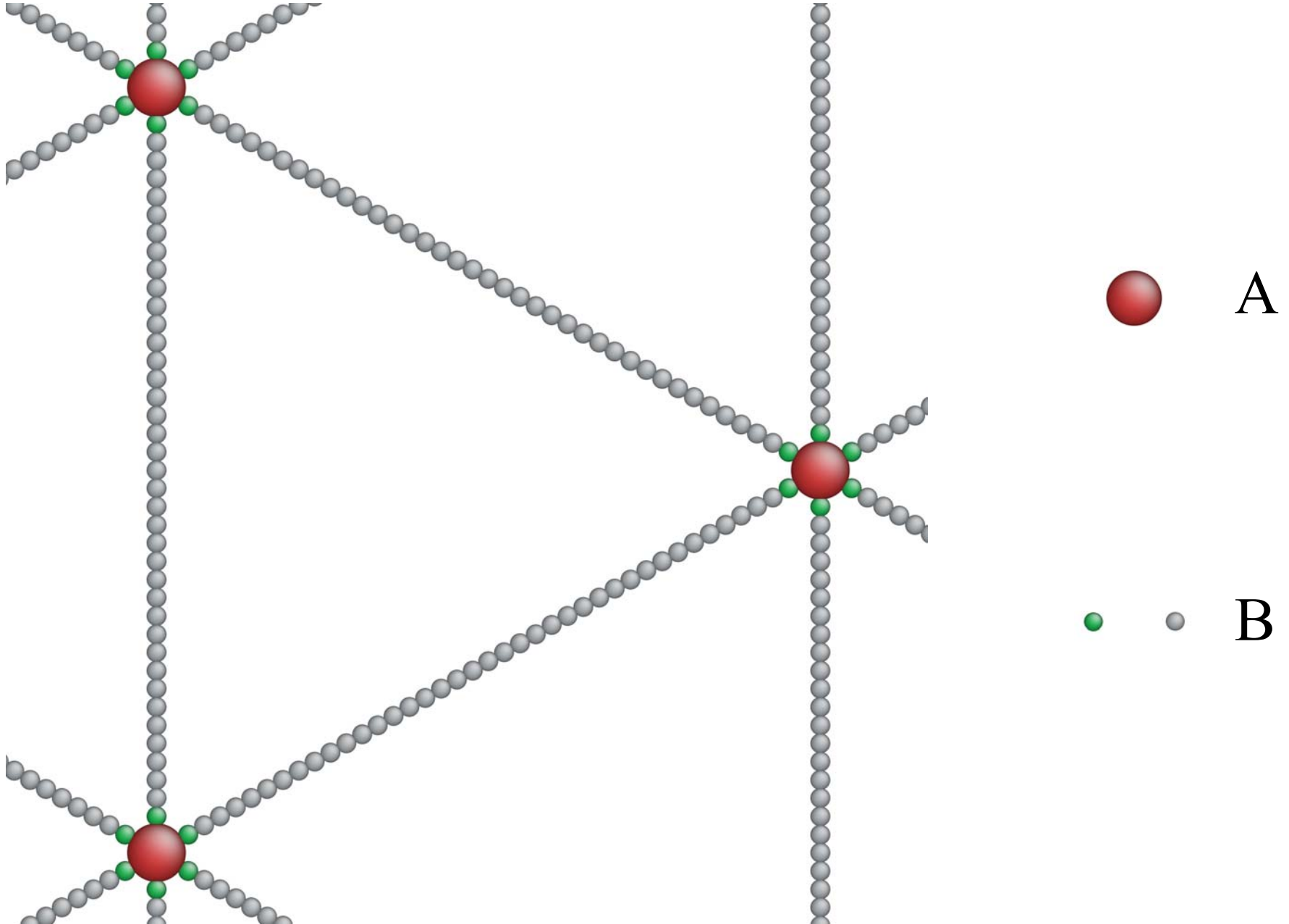
	actin#	spectrin#	largest polygon hole
normal	28673	81718	6
degree-4.5	26880	57523	8
degree-4	24372	48012	11
degree-3.5	21504	37416	22
degree-3	18637	26837	35

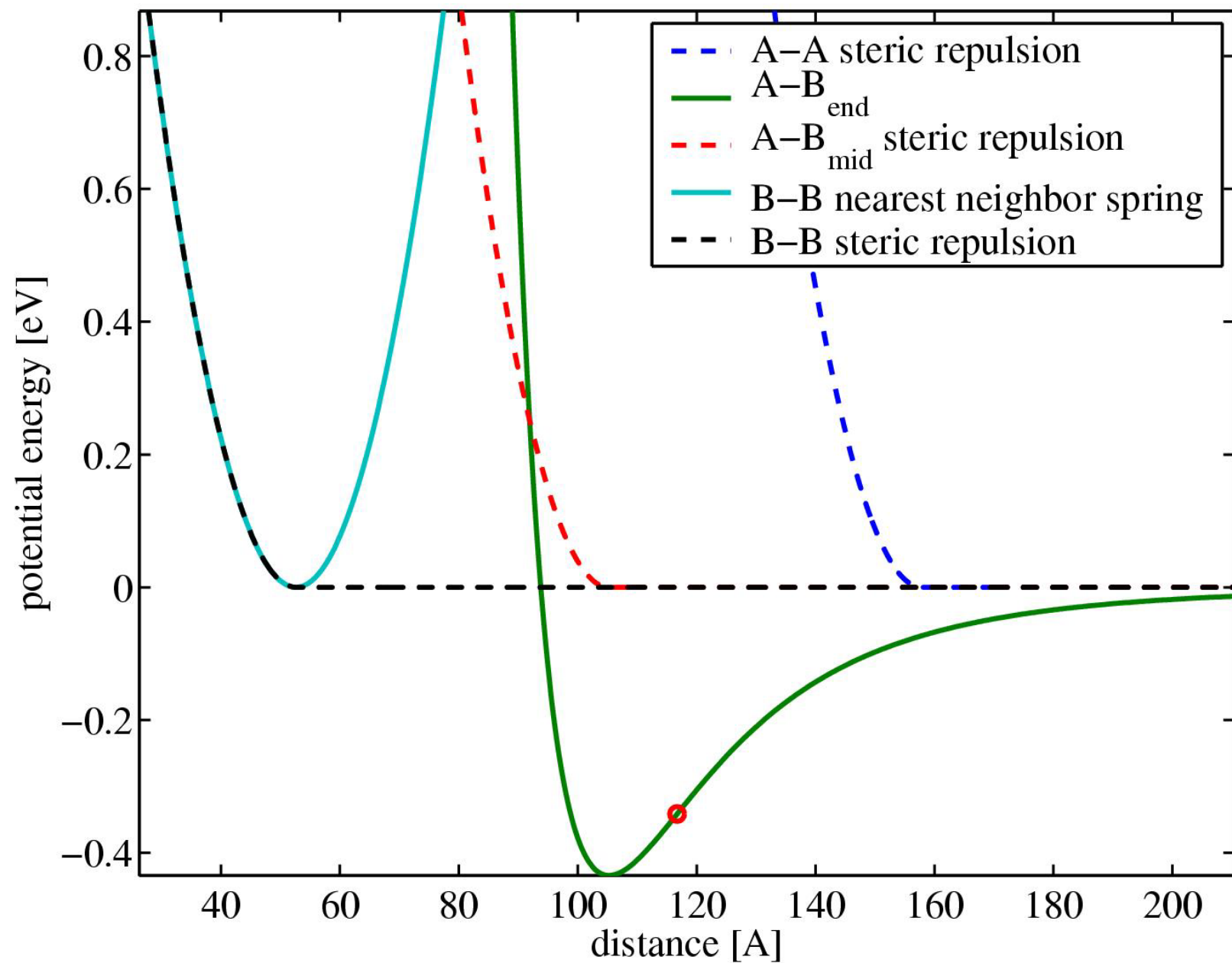
But this is basically from a “geometrical” simulation
no biophysical basis, yet.

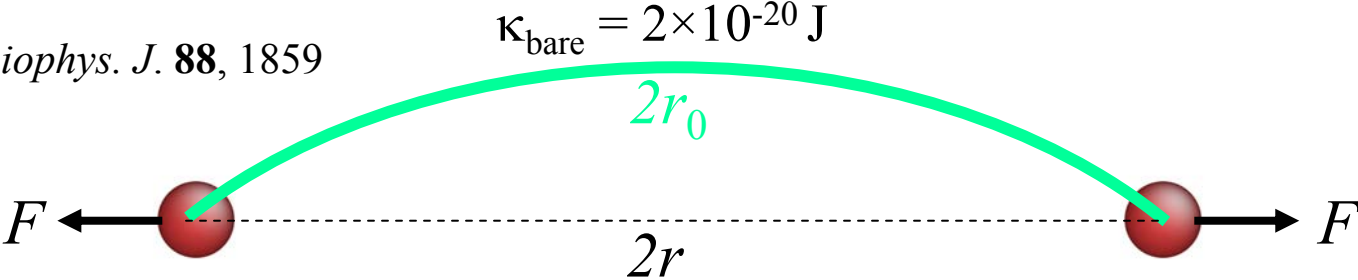
Intermediate Summary

- Spectrin-level and continuum FEM analyses indicate our optical tweezers experiments give approximately the same in-plane shear modulus as micropipette aspiration experiments: $\mu = 5 \text{ to } 10 \times 10^{-6} \text{ N/m}$.
- Stabilization of biconcave equilibrium shape strongly suggests the cytoskeleton undergoes slow but constant remodeling topologically to always relax the in-plane shear elastic energy to zero.
- Connection to single-molecule stretching experiments (“intermolecular potential development”).

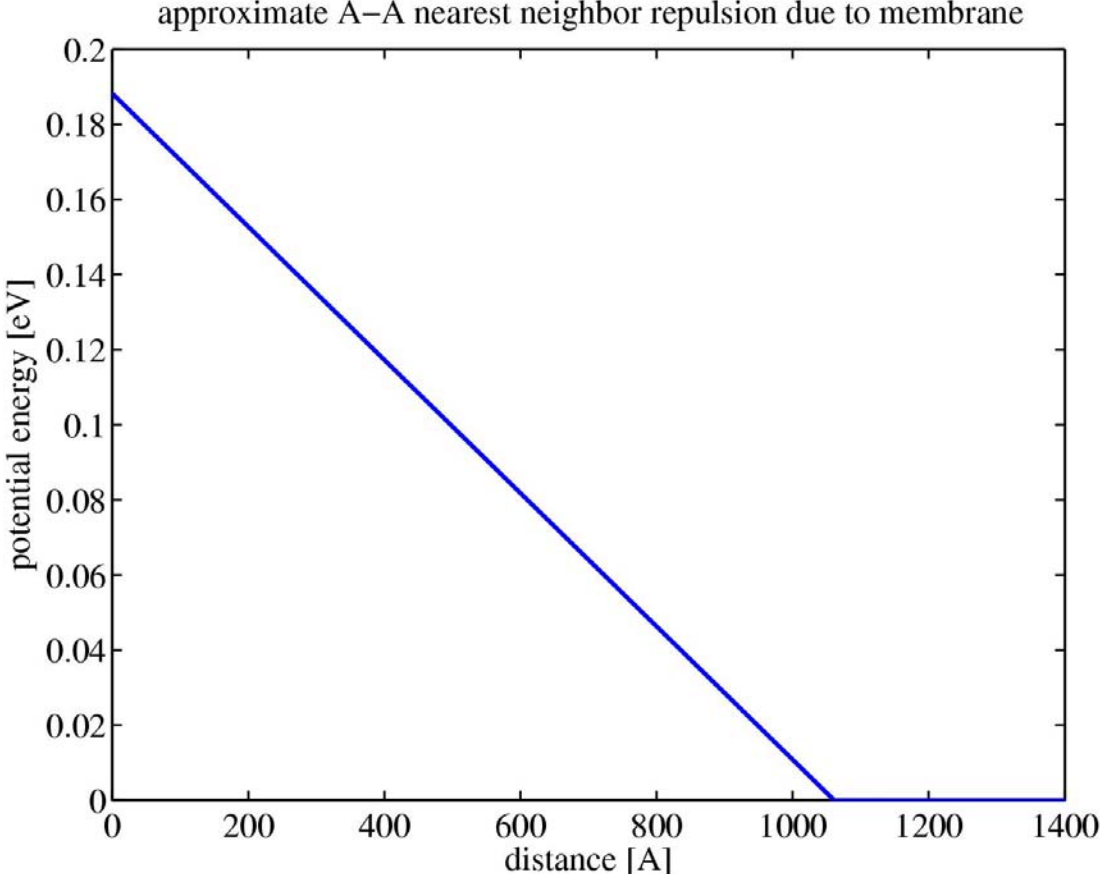
CGMD model with *breakable* actin-spectrin junction



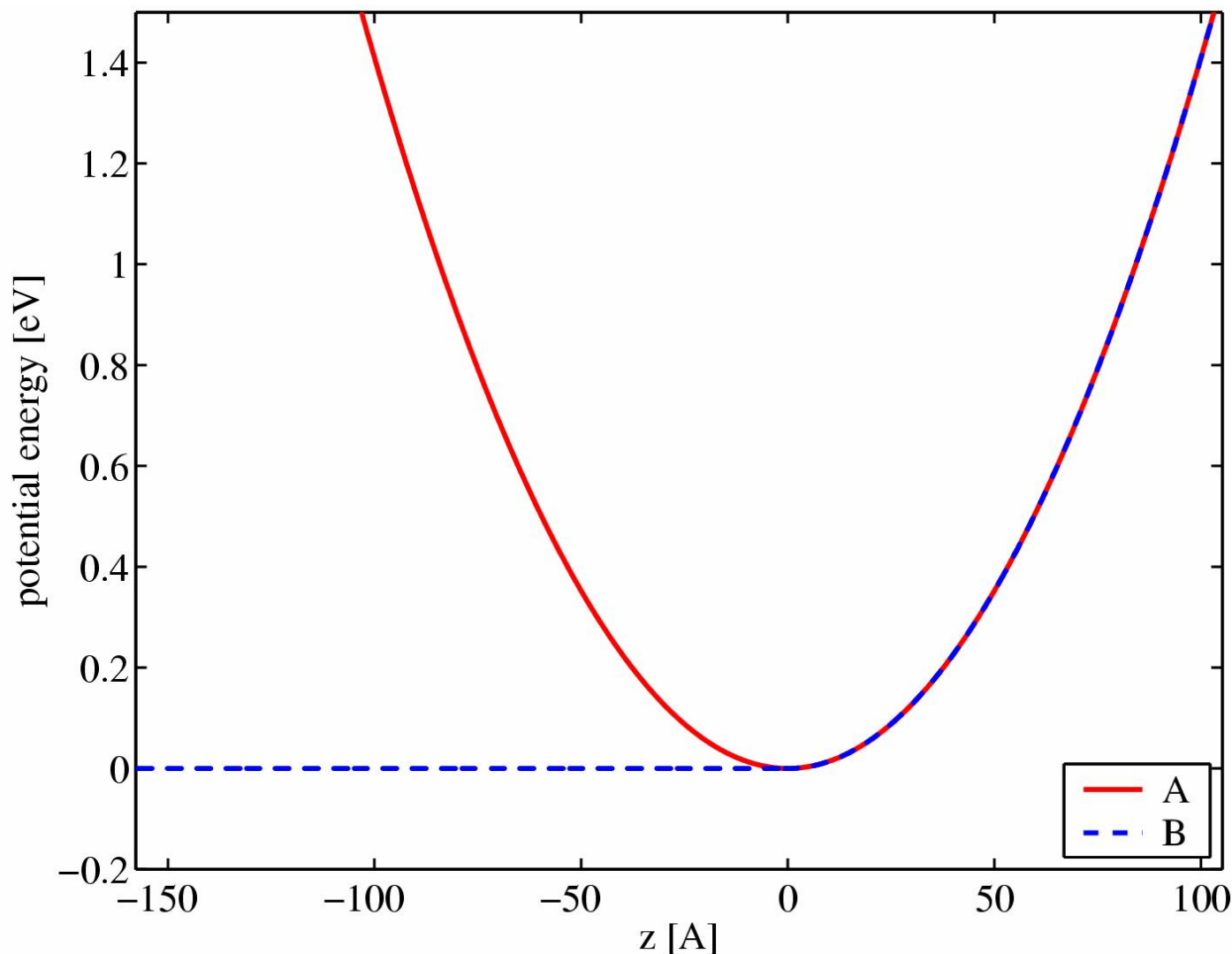


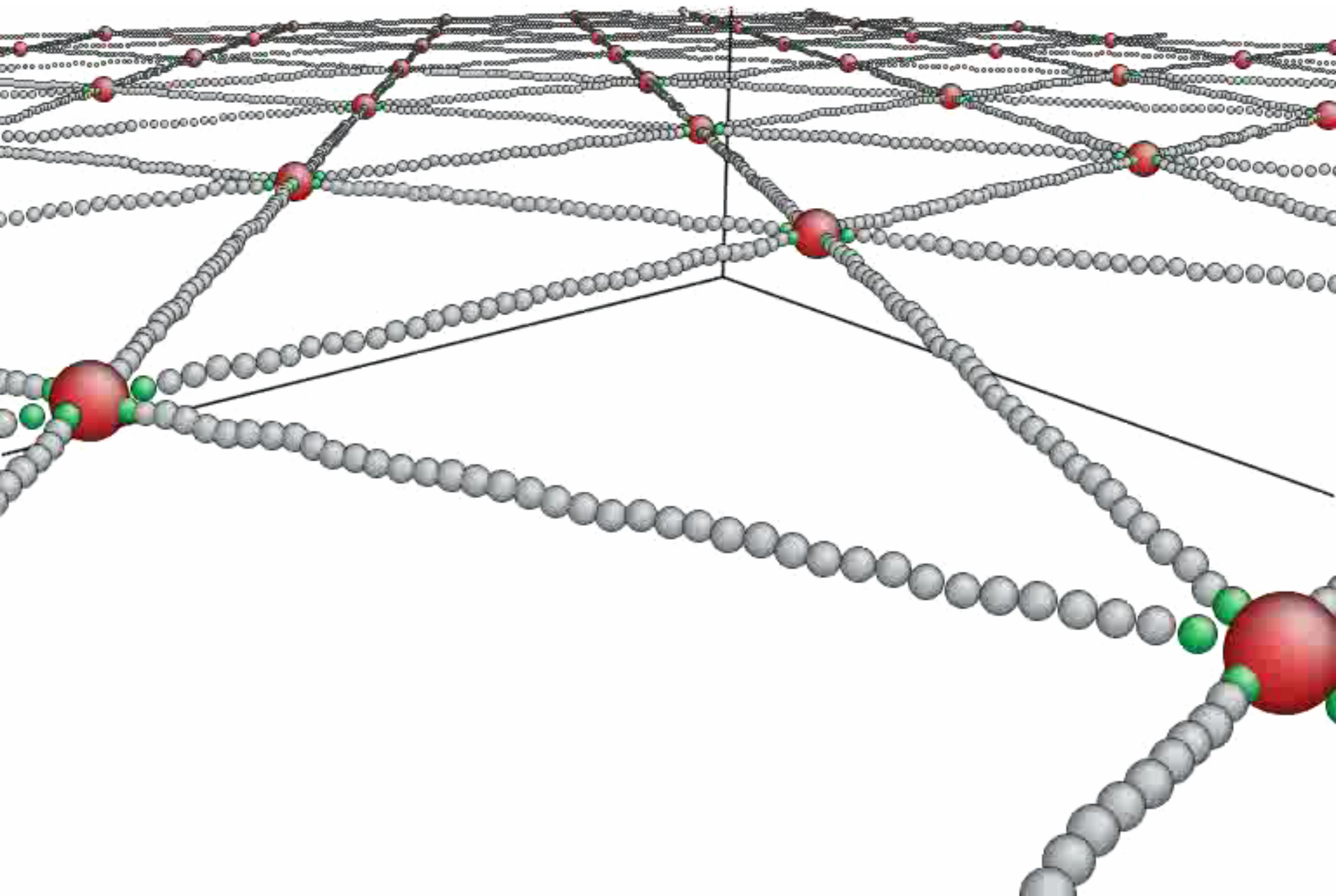


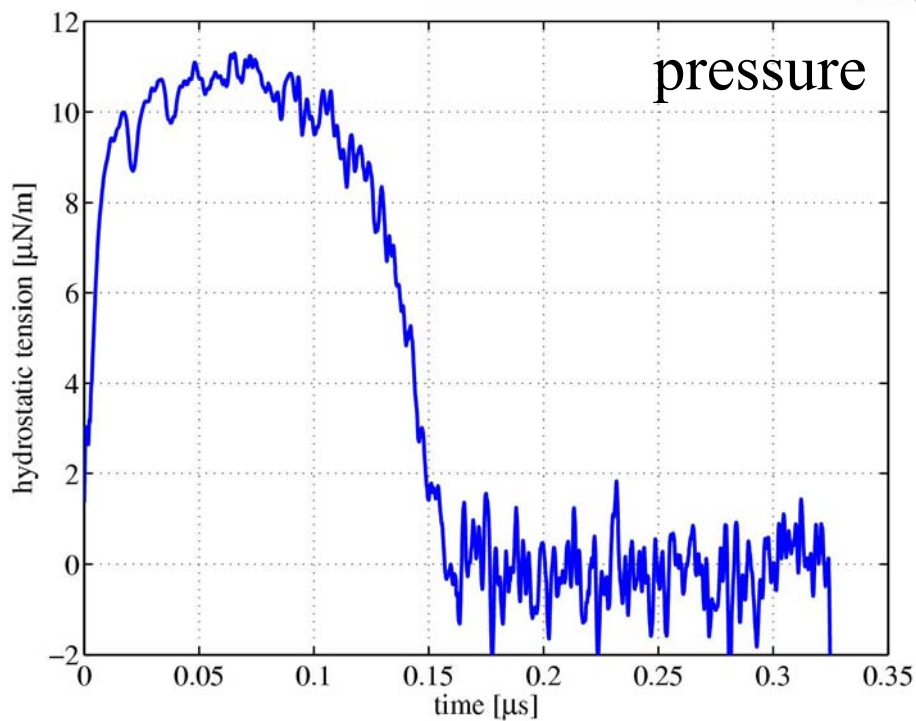
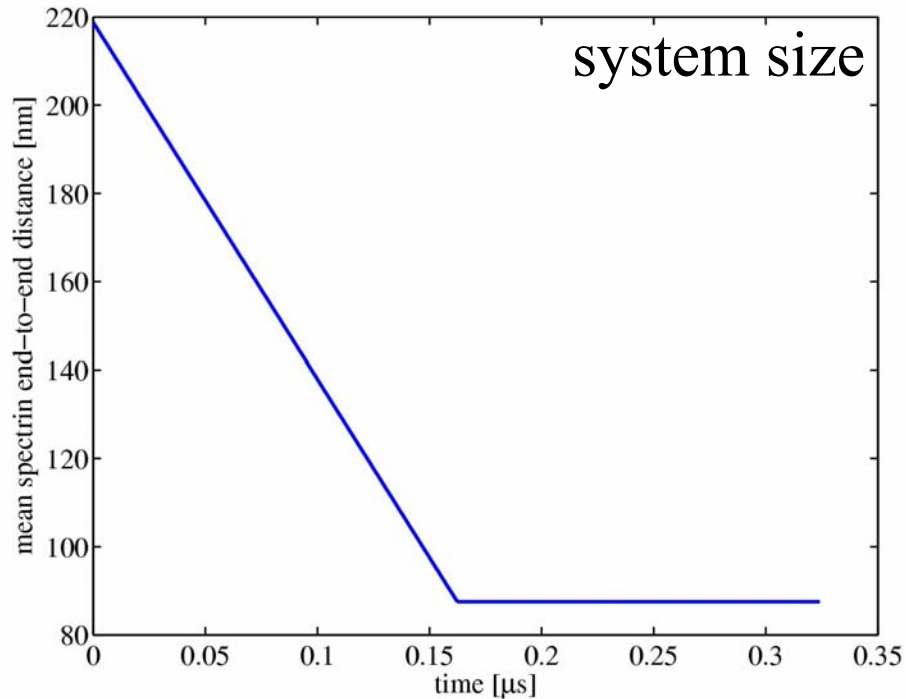
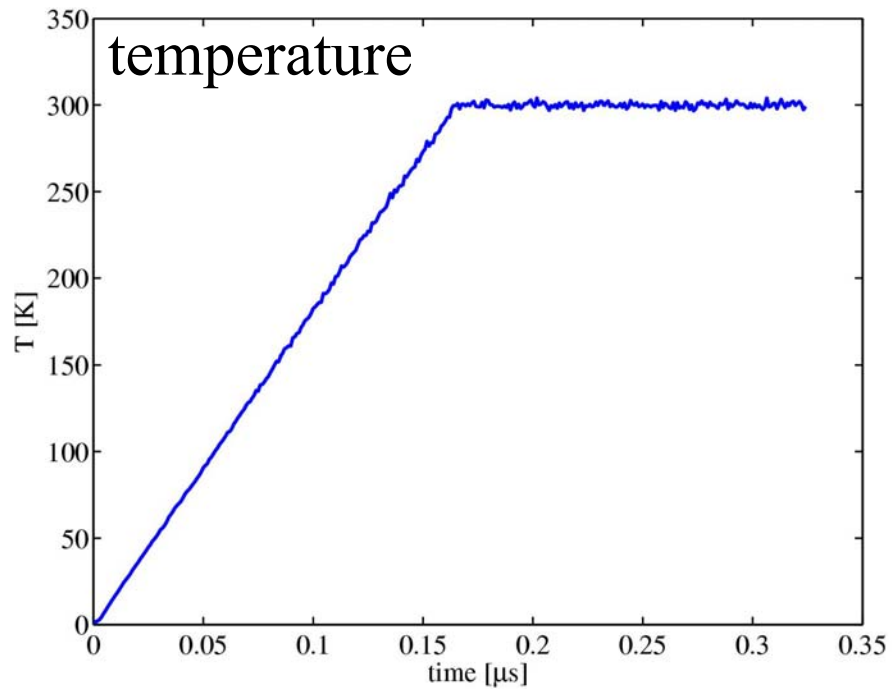
$$F = \alpha \frac{4\pi\kappa_{\text{bare}}}{3r_0}, \quad \alpha \text{ chosen to be } 0.36$$



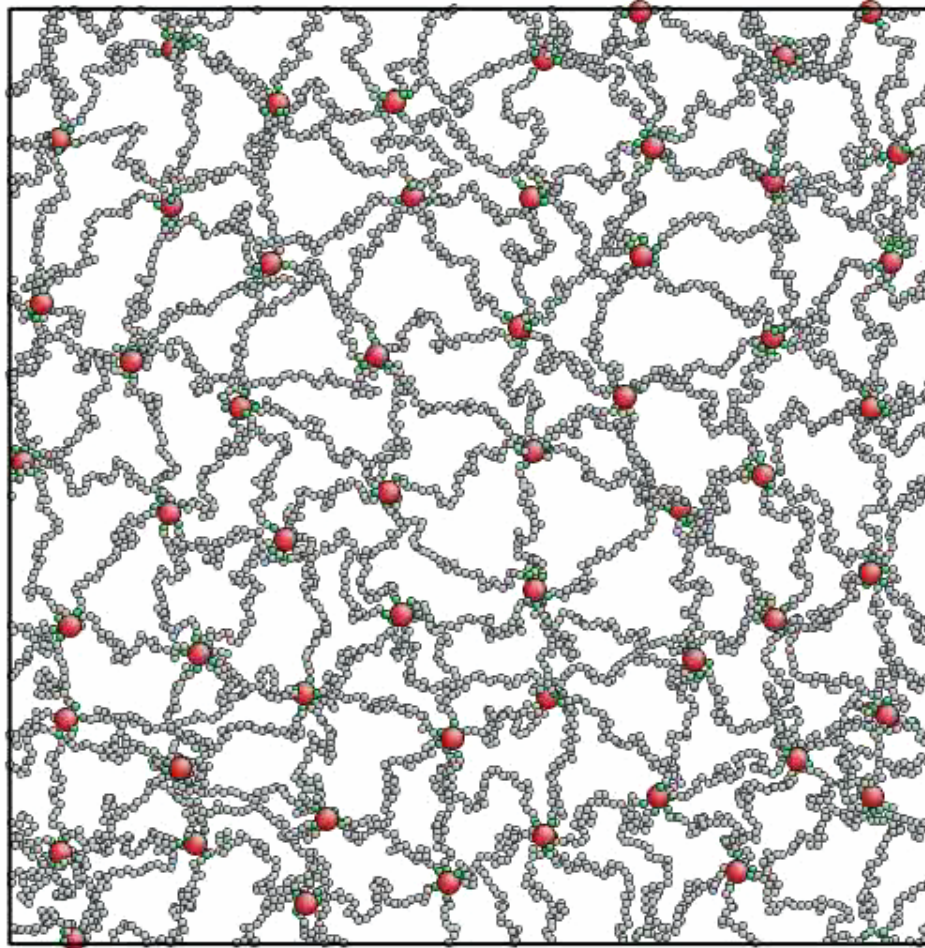
We also put soft ($0.1k_{\text{BB}}$) confinement potential on A and B in z to mimic interaction with the membrane without actually simulating the membrane.



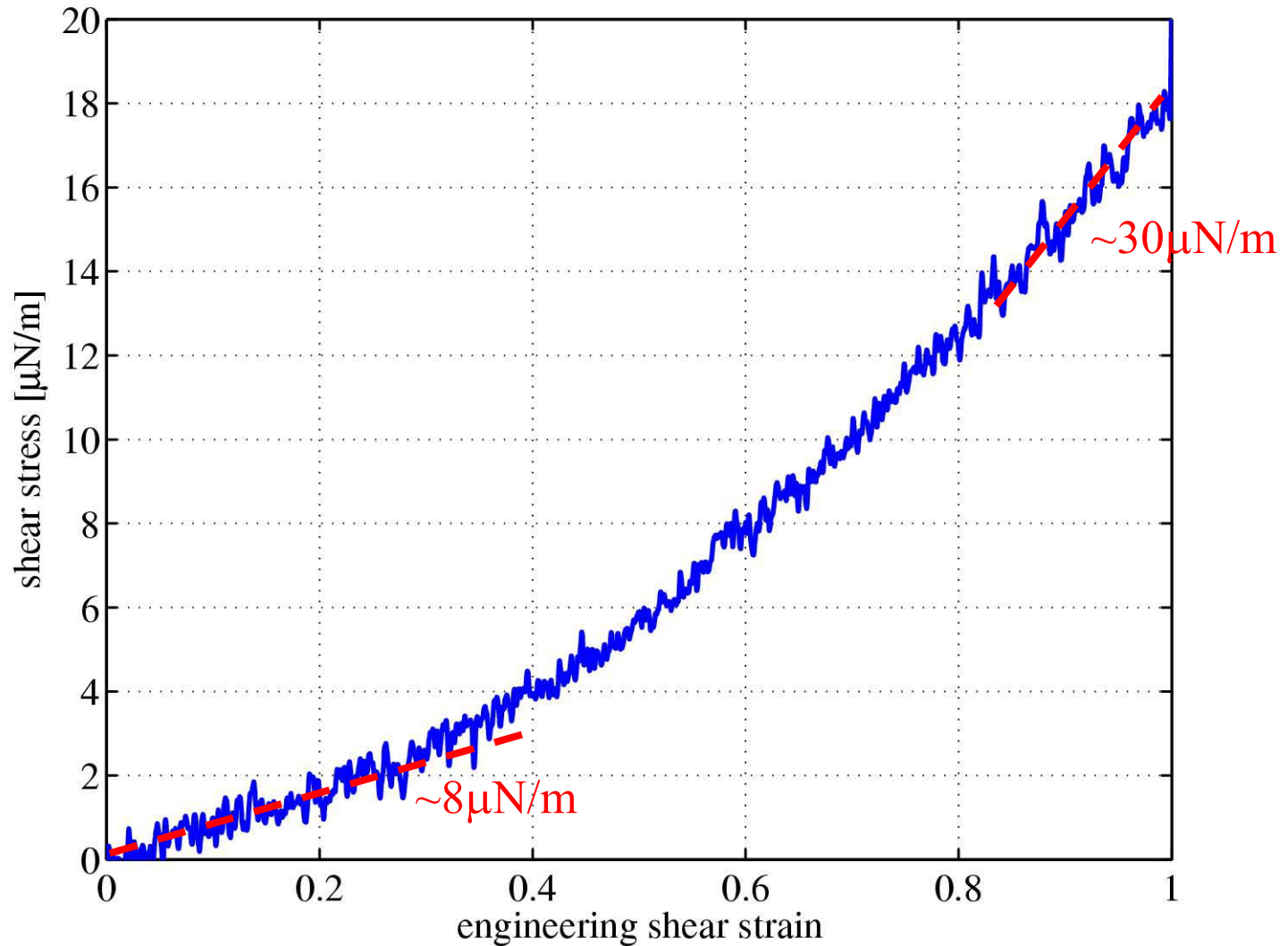




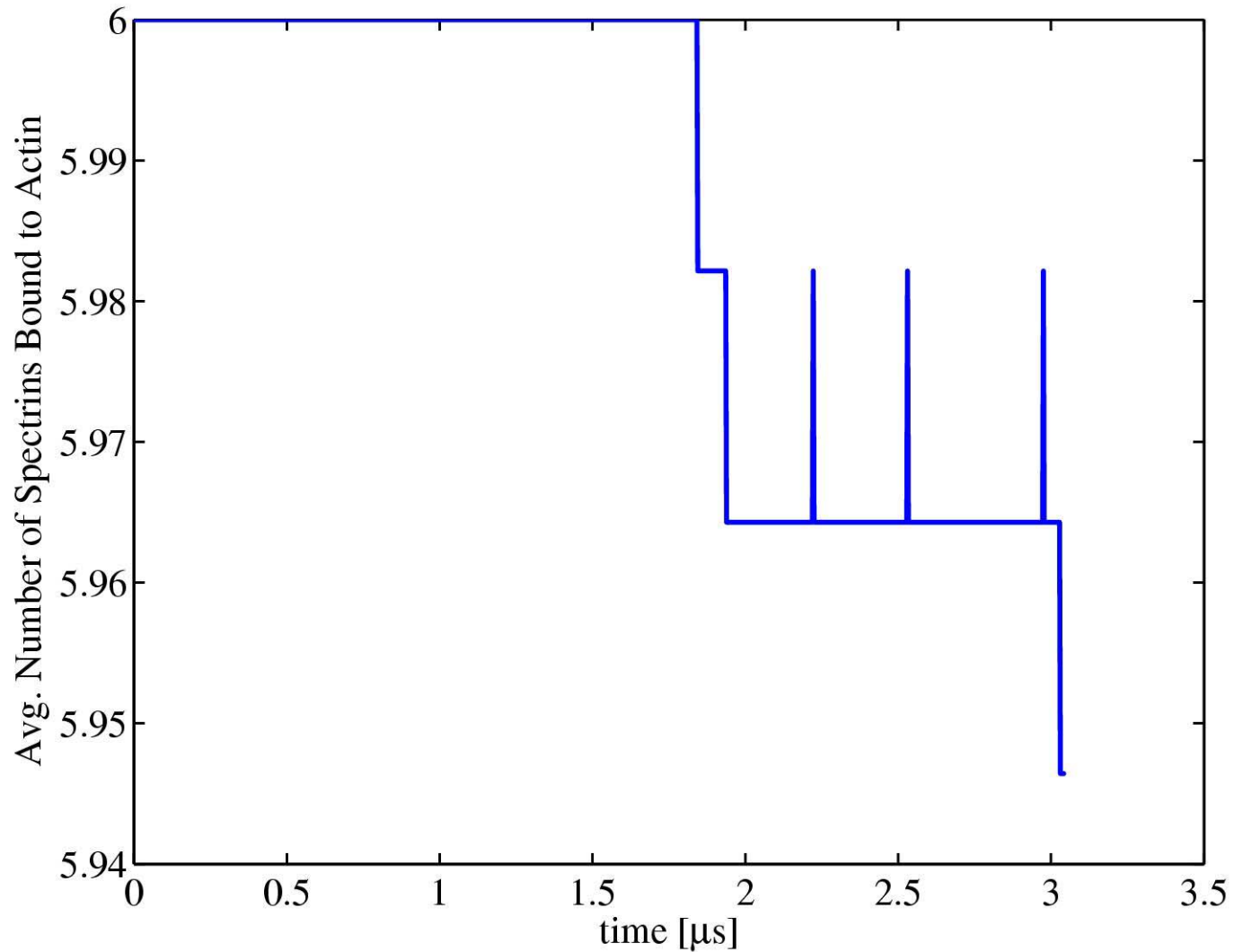
Pure shear deformation
at 300K and strain rate $3 \times 10^5/s$



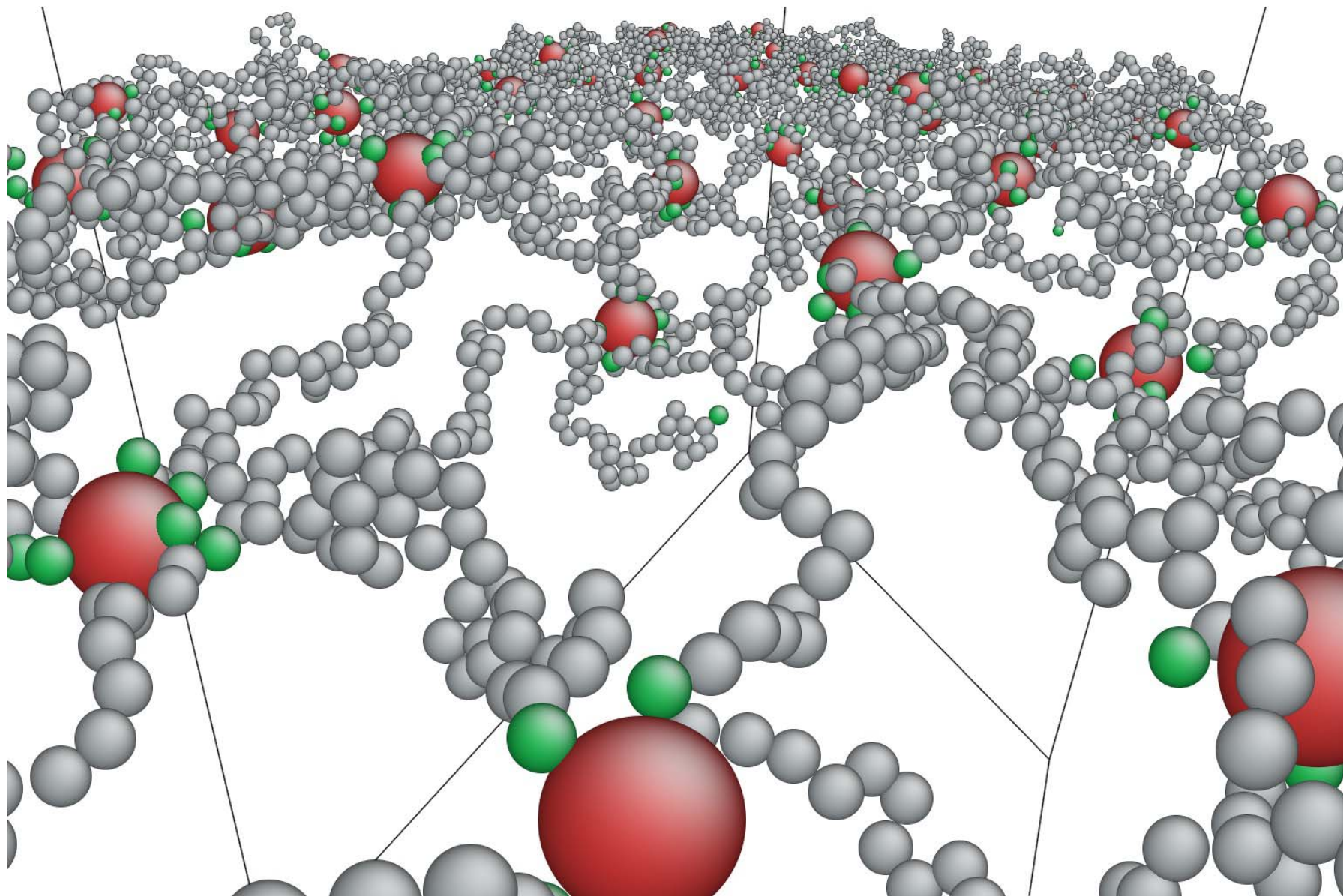
Stress-strain curve at 300K and no ATP

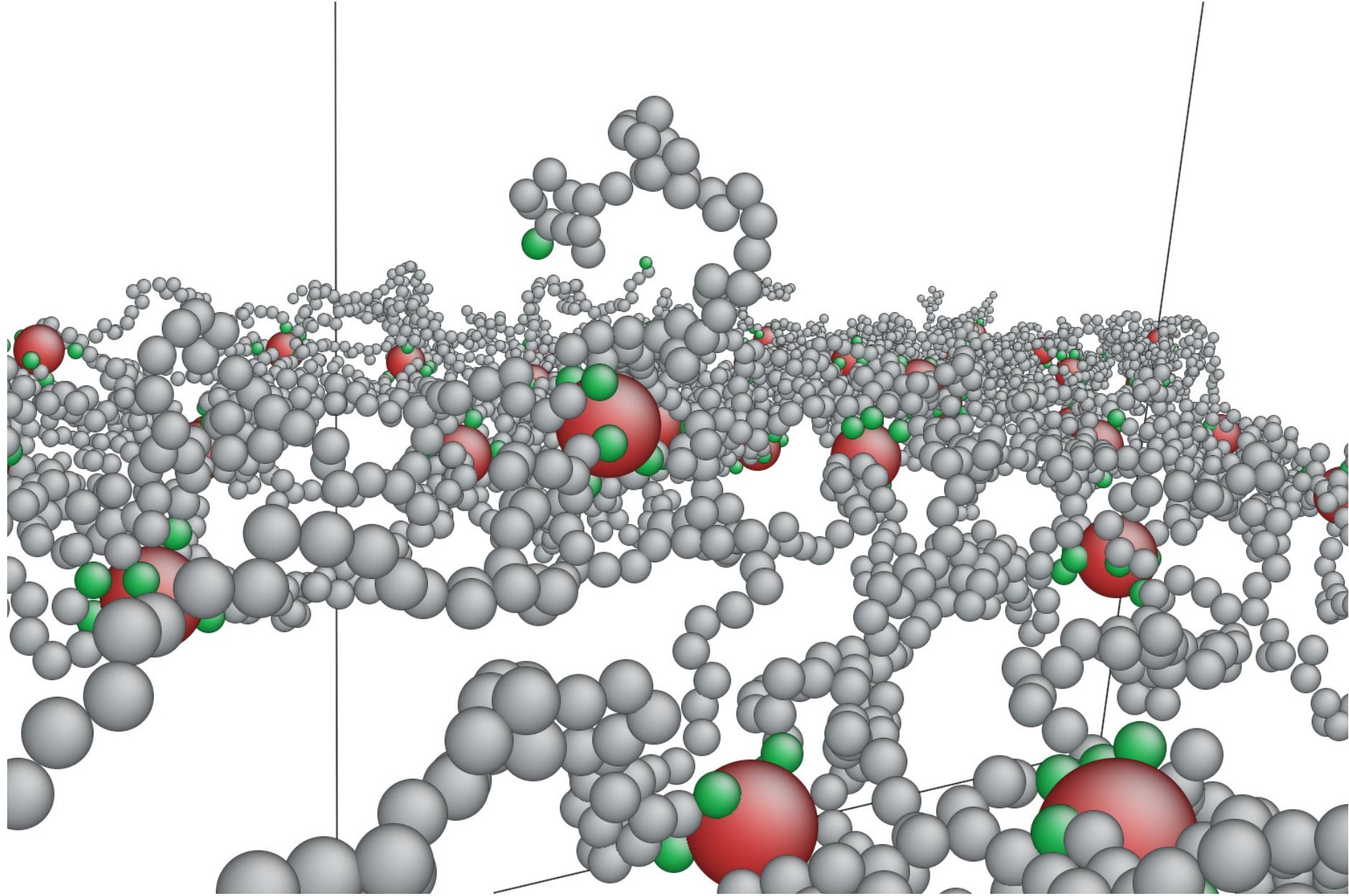


Defect statistics at 300K with no ATP



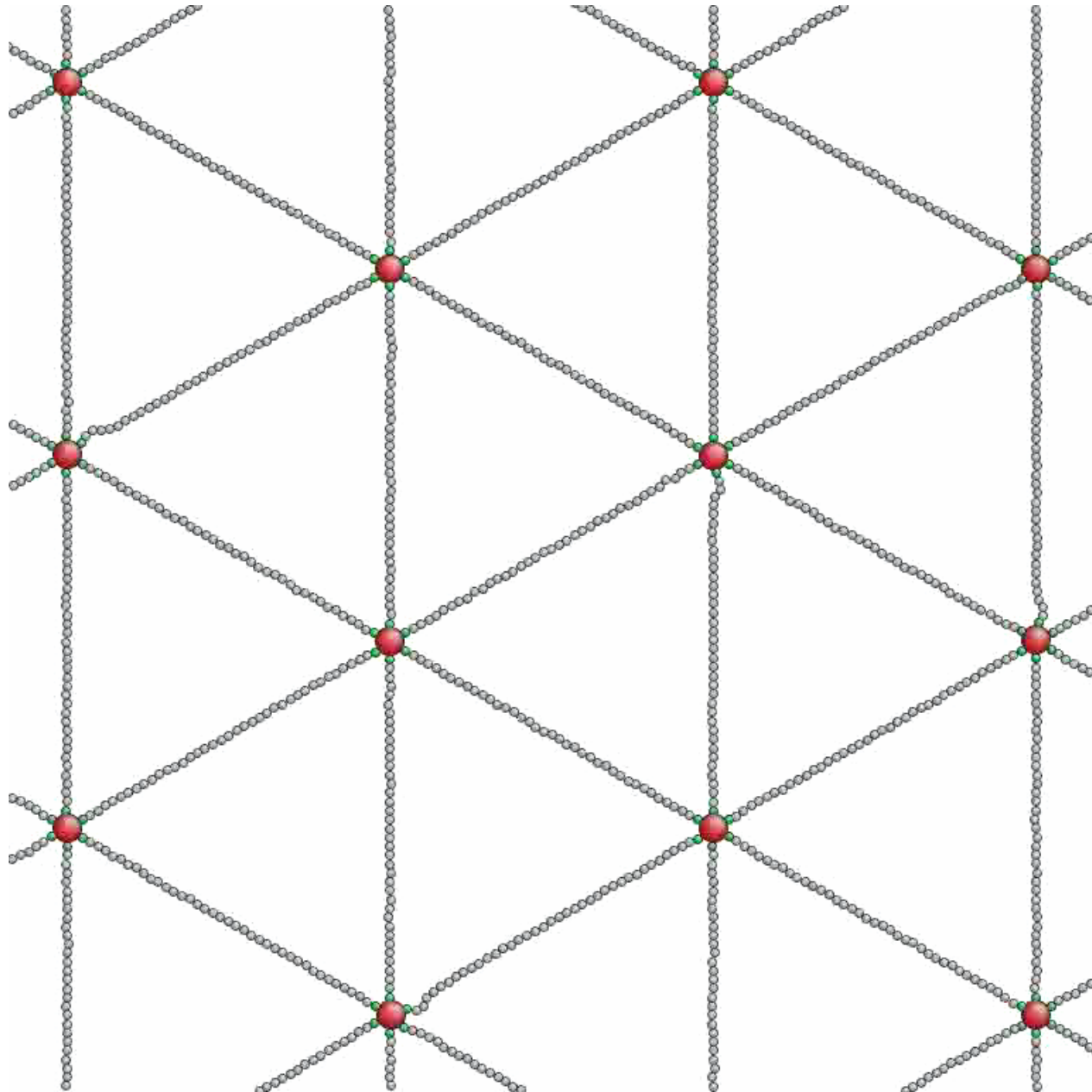
A broken link 5-fold defect



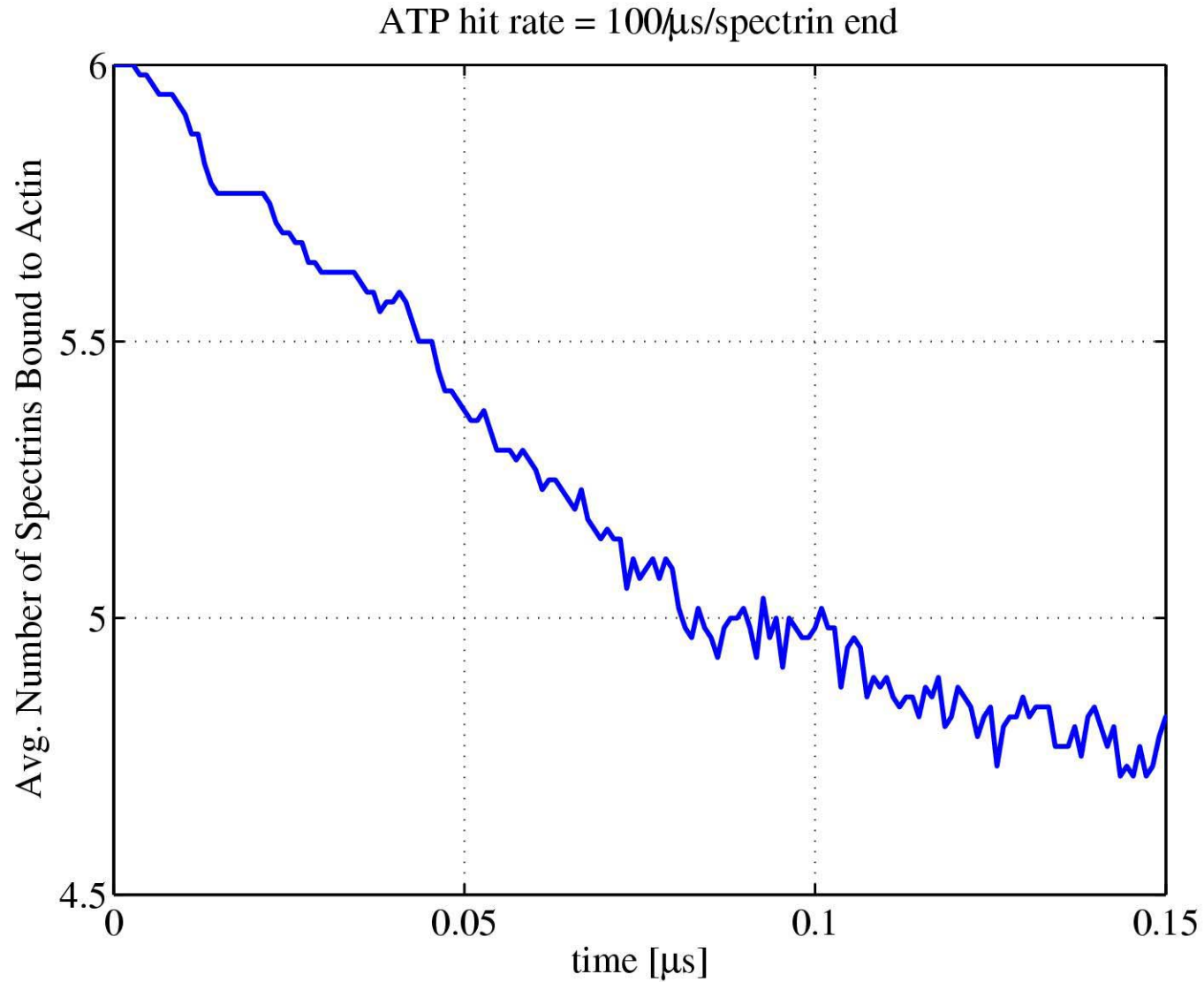


Corrugation due to buckling: elevated / depressed in height

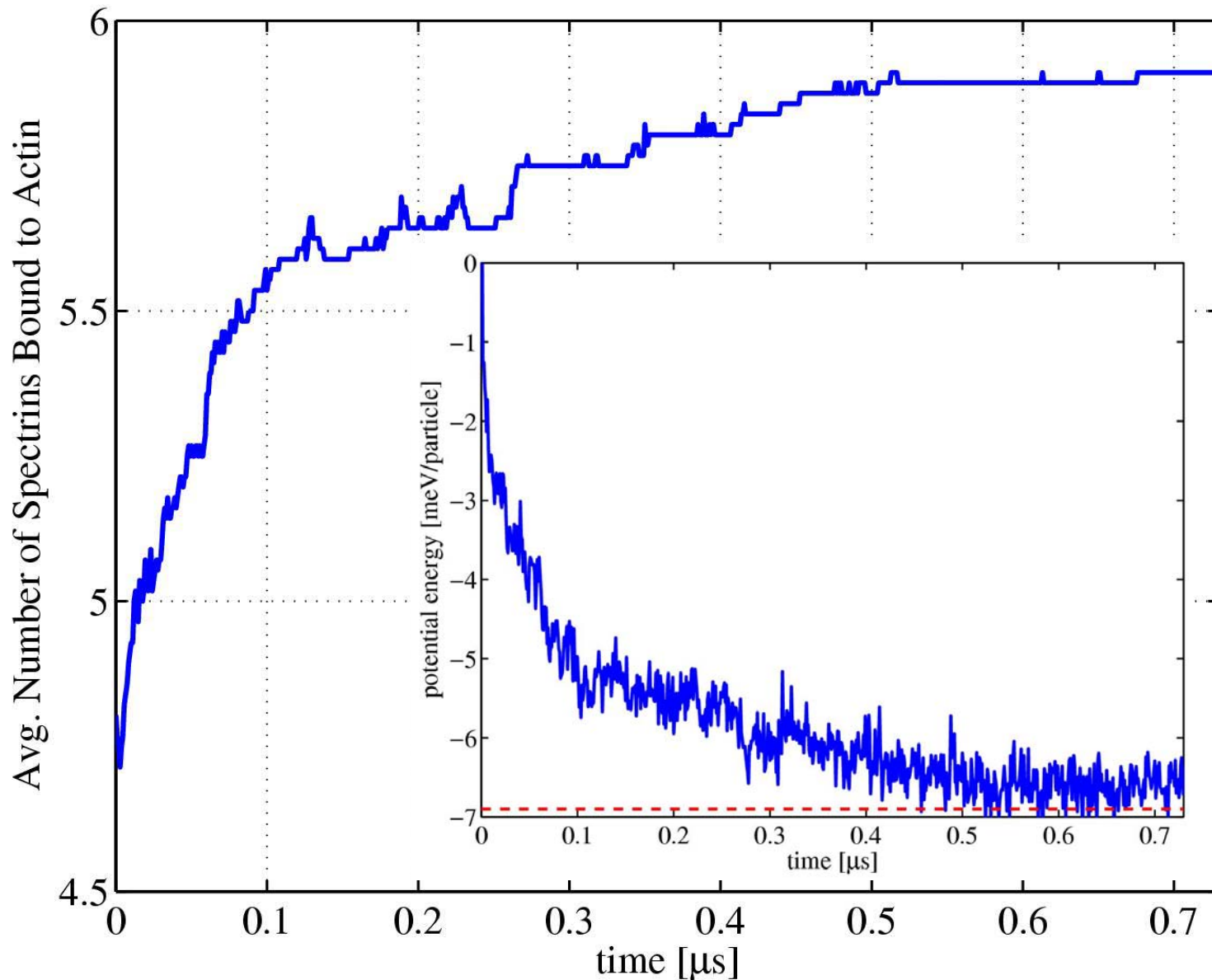
Now add ATP (0.5eV random kinetic energy to green ball):
hit rate = 100/ μ s per spectrin end



Defect statistics at 300K, ATP hit rate $100/\mu\text{s}$



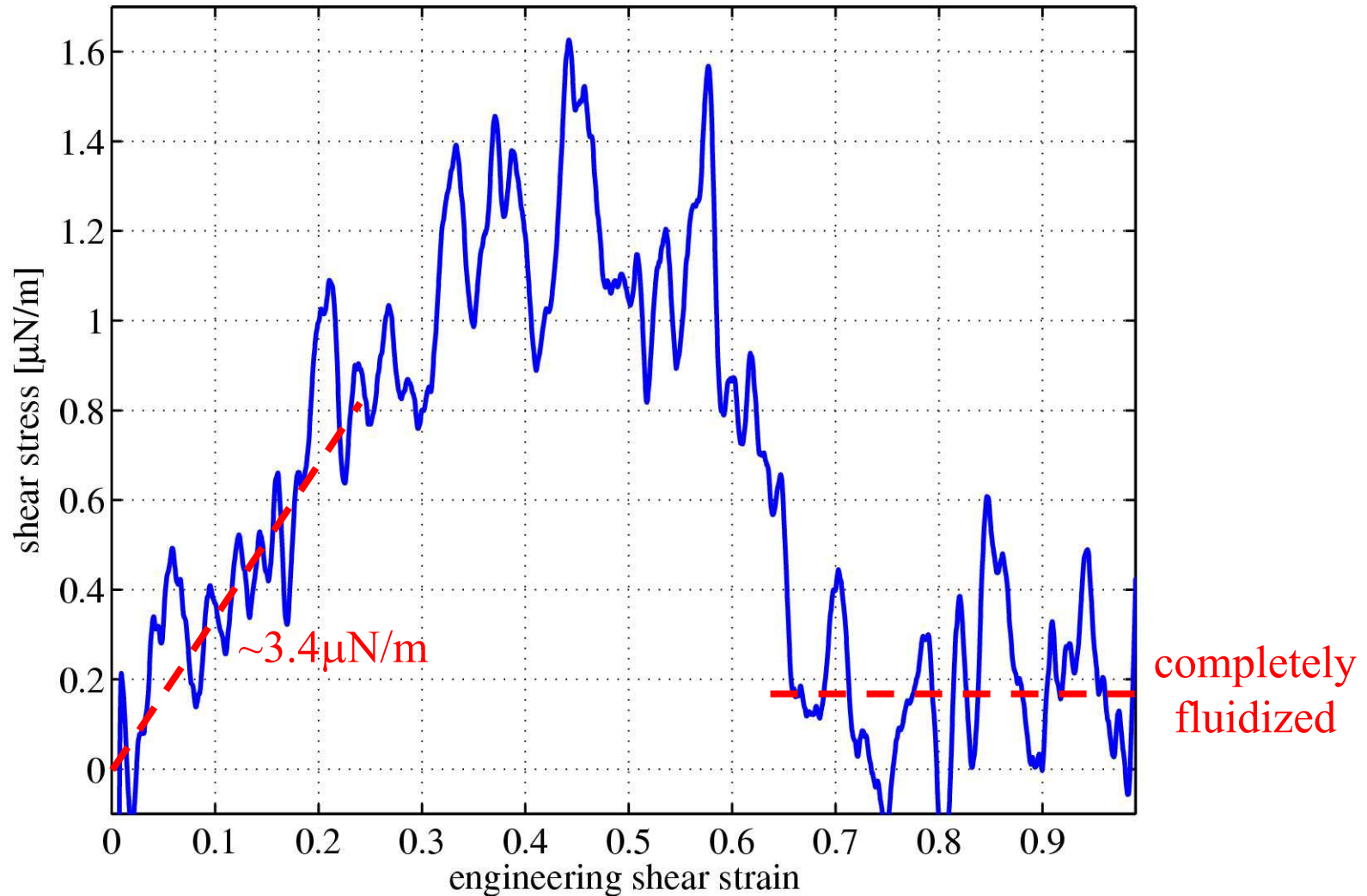
Now turn off ATP hits, “anneal” at 300K...



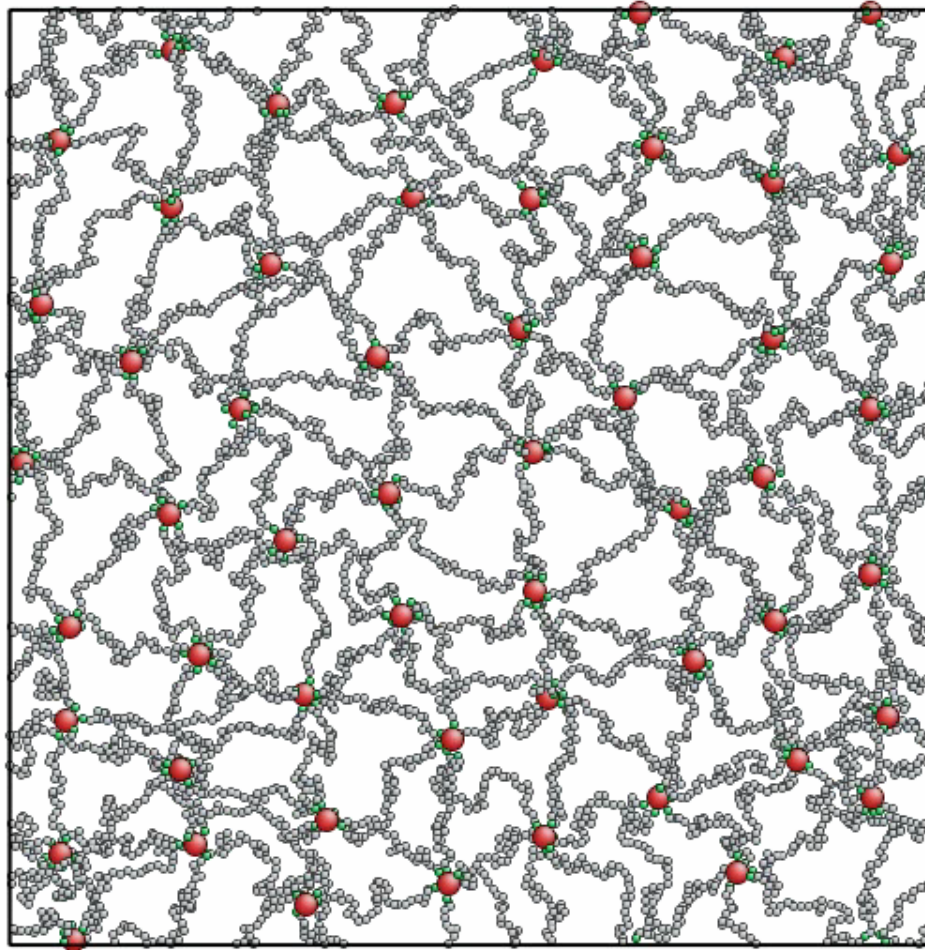
Miraculously, the system recovers, within CGMD simulation timescale.

A more reasonable ATP hit rate: $10/\mu\text{s}$.
Simultaneously, also shear deform.

ATP hit rate = $10/\mu\text{s}$ /spectrin, strain rate = $3 \times 10^5/\text{s}$

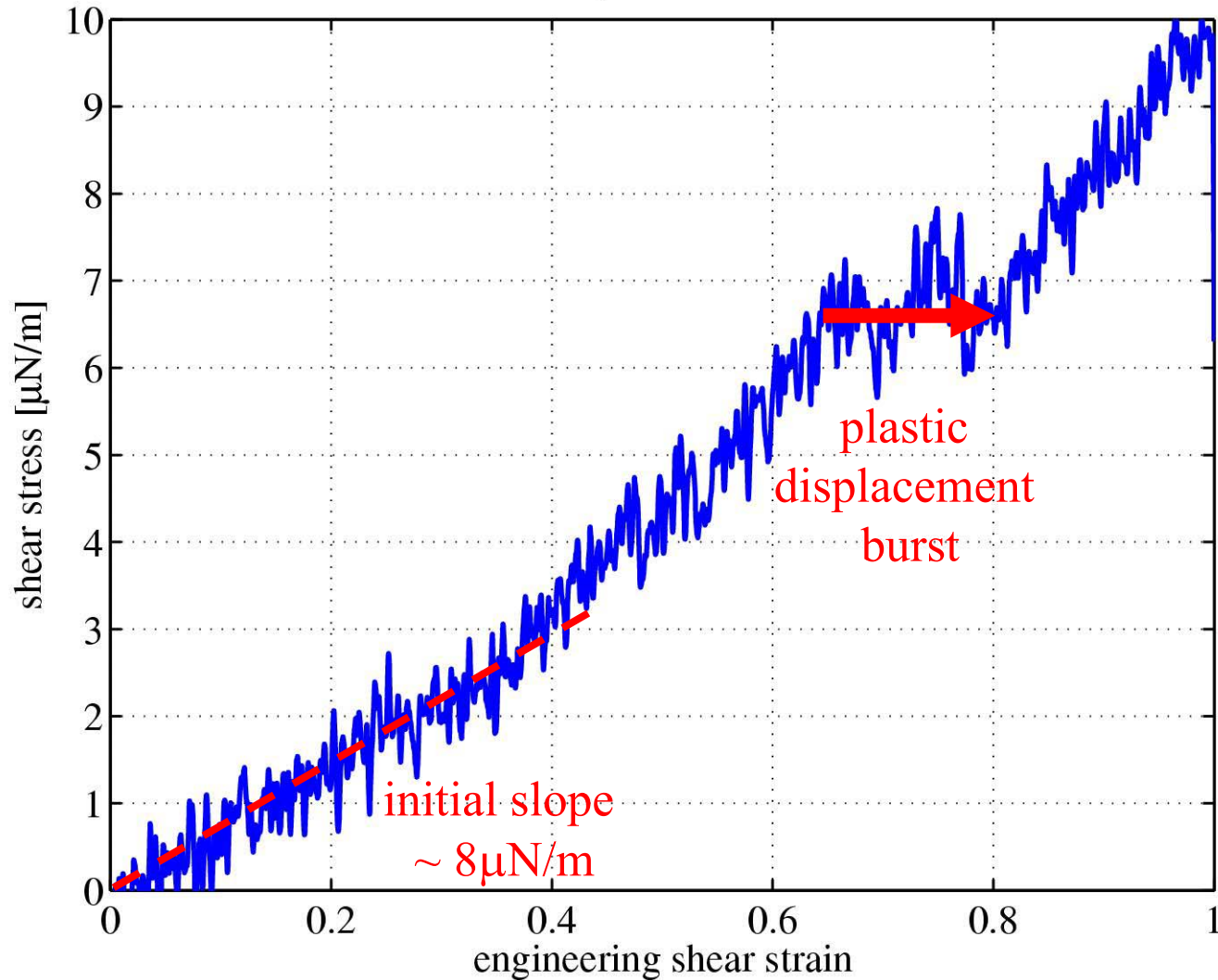


ATP hit rate = $10/\mu\text{s}$

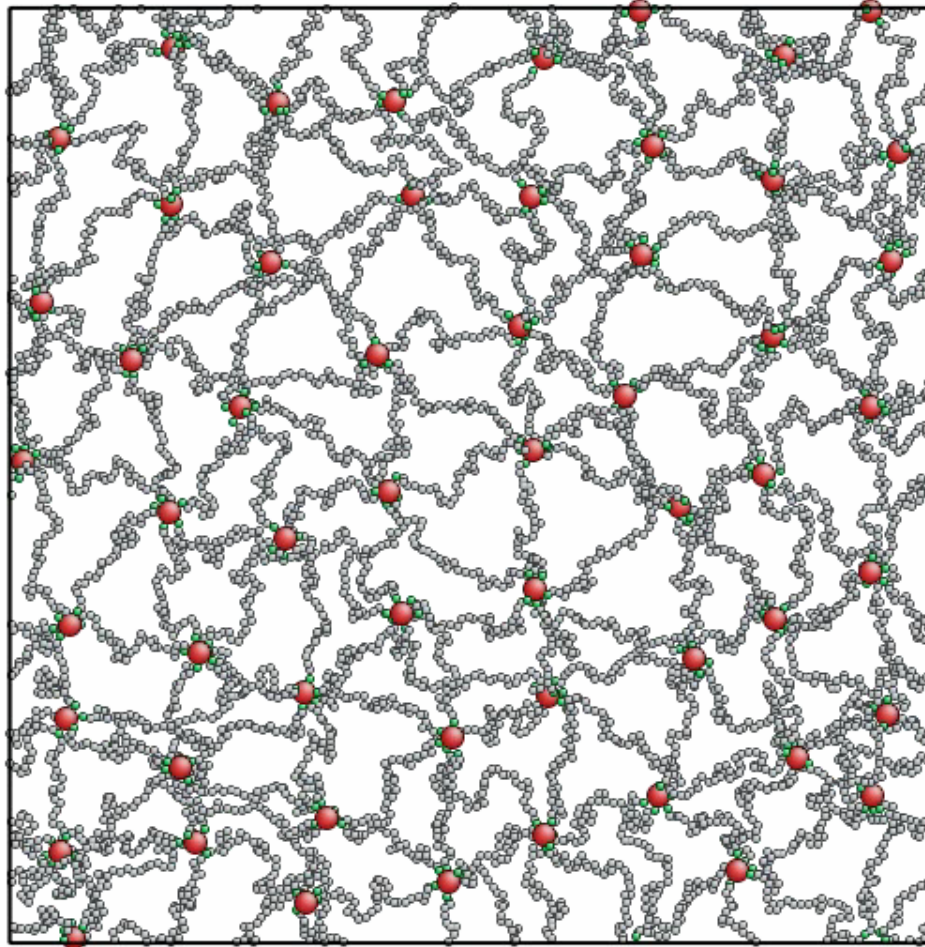


ATP hit rate = $1/\mu\text{s}$:

ATP hit rate = $1/\mu\text{s}$, strain rate = $3 \times 10^5/\text{s}$

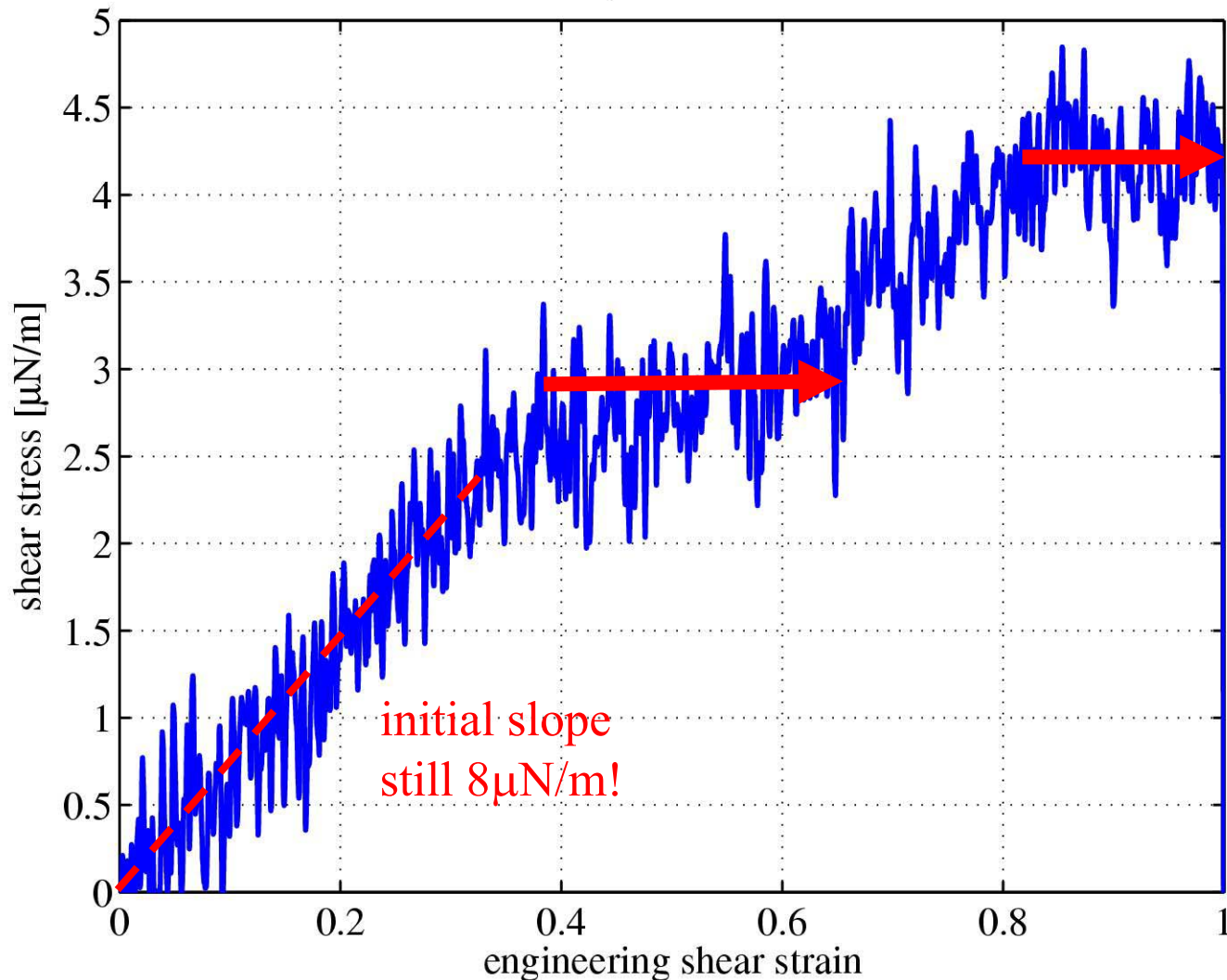


ATP hit rate = $1/\mu s$

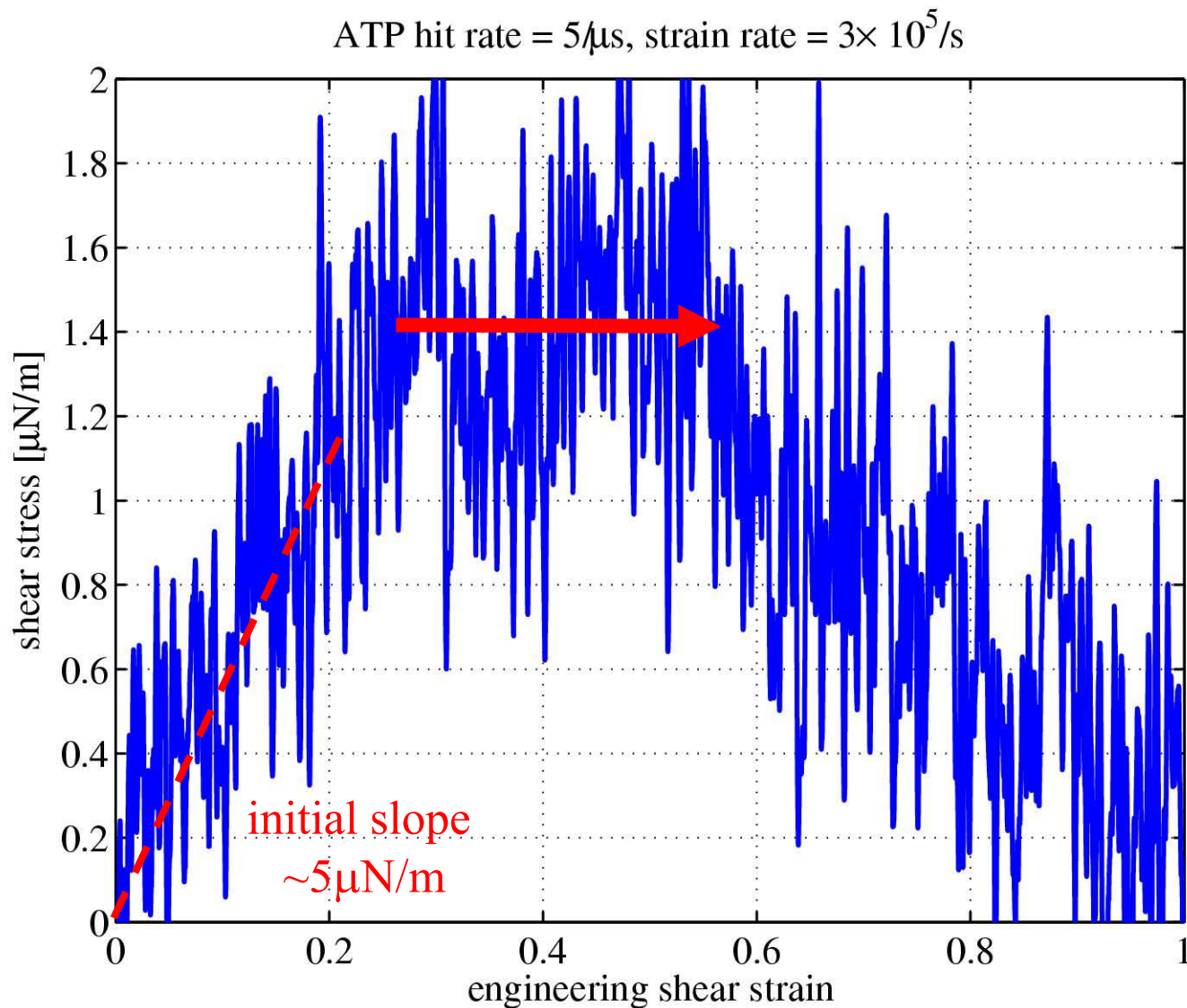


ATP hit rate = $2/\mu\text{s}$:
two plastic displacements... also longer

ATP hit rate = $2/\mu\text{s}$, strain rate = $3 \times 10^5/\text{s}$



ATP hit rate = $5/\mu\text{s}$: large-strain resistance collapses, manifest global yield



Schematic Model of the RBC Membrane

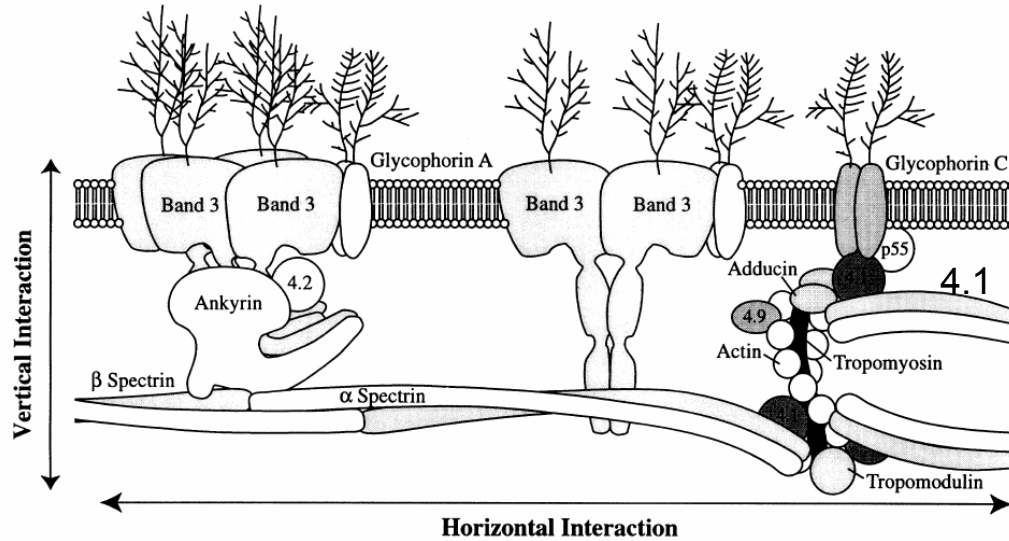
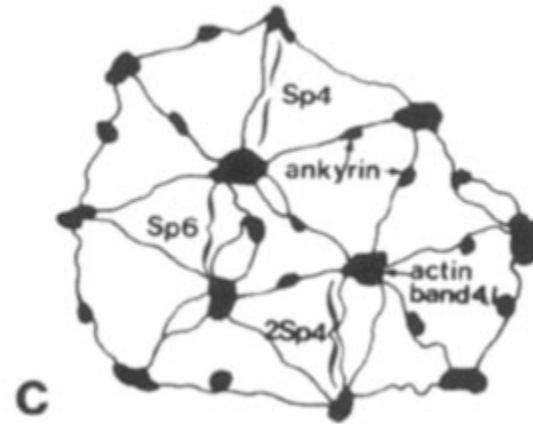
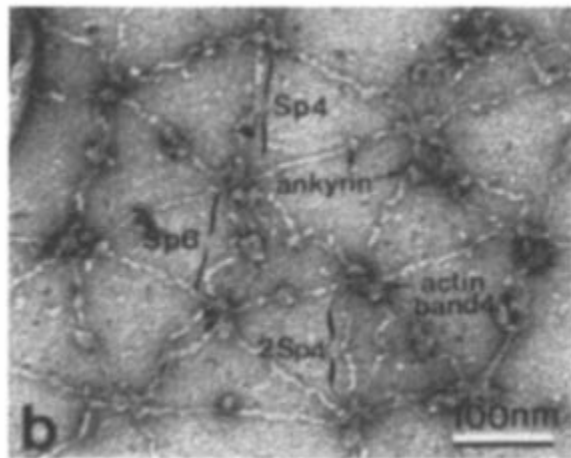


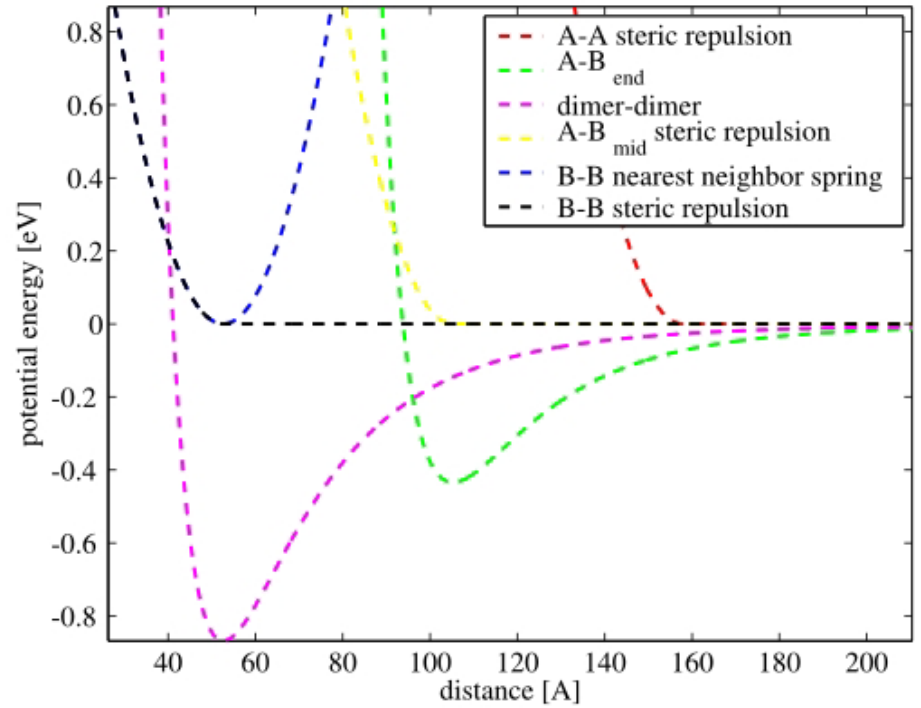
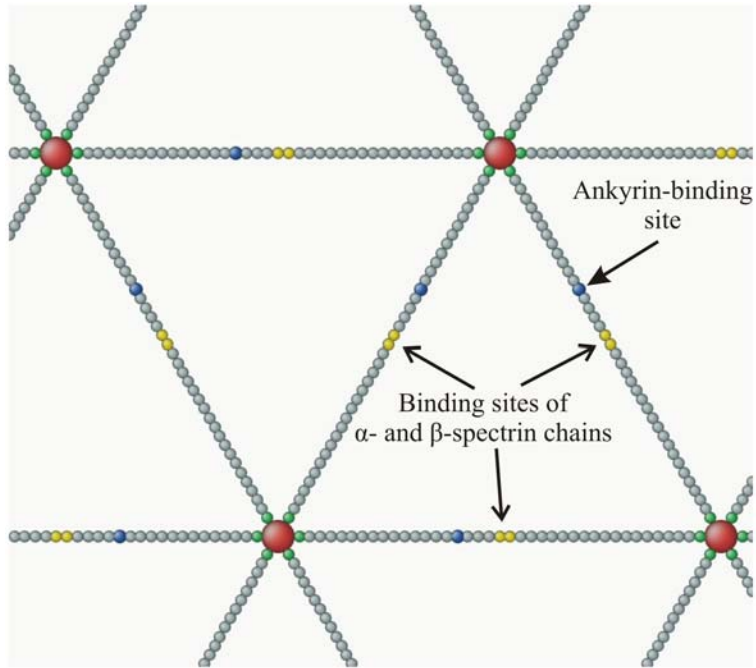
Fig 1. Schematic model of the red cell membrane, with the vertical and horizontal interaction of its components indicated. Estimated frequencies of mutations in different membrane proteins in HS and HE/HPP are as follows. Vertical interaction: hereditary spherocytosis: band 3, ~20%; protein 4.2, ~5%; ankyrin, ~45%; β spectrin, ~30%. Horizontal interaction: hereditary elliptocytosis/hereditary pyropoikilocytosis: β spectrin, ~5%; α spectrin, ~80%; protein 4.1, ~15%. The relative position of the various proteins is correct, but the proteins and lipids are not drawn to scale. Adapted from Lux & Palek (1995).

Tse et al. 1999

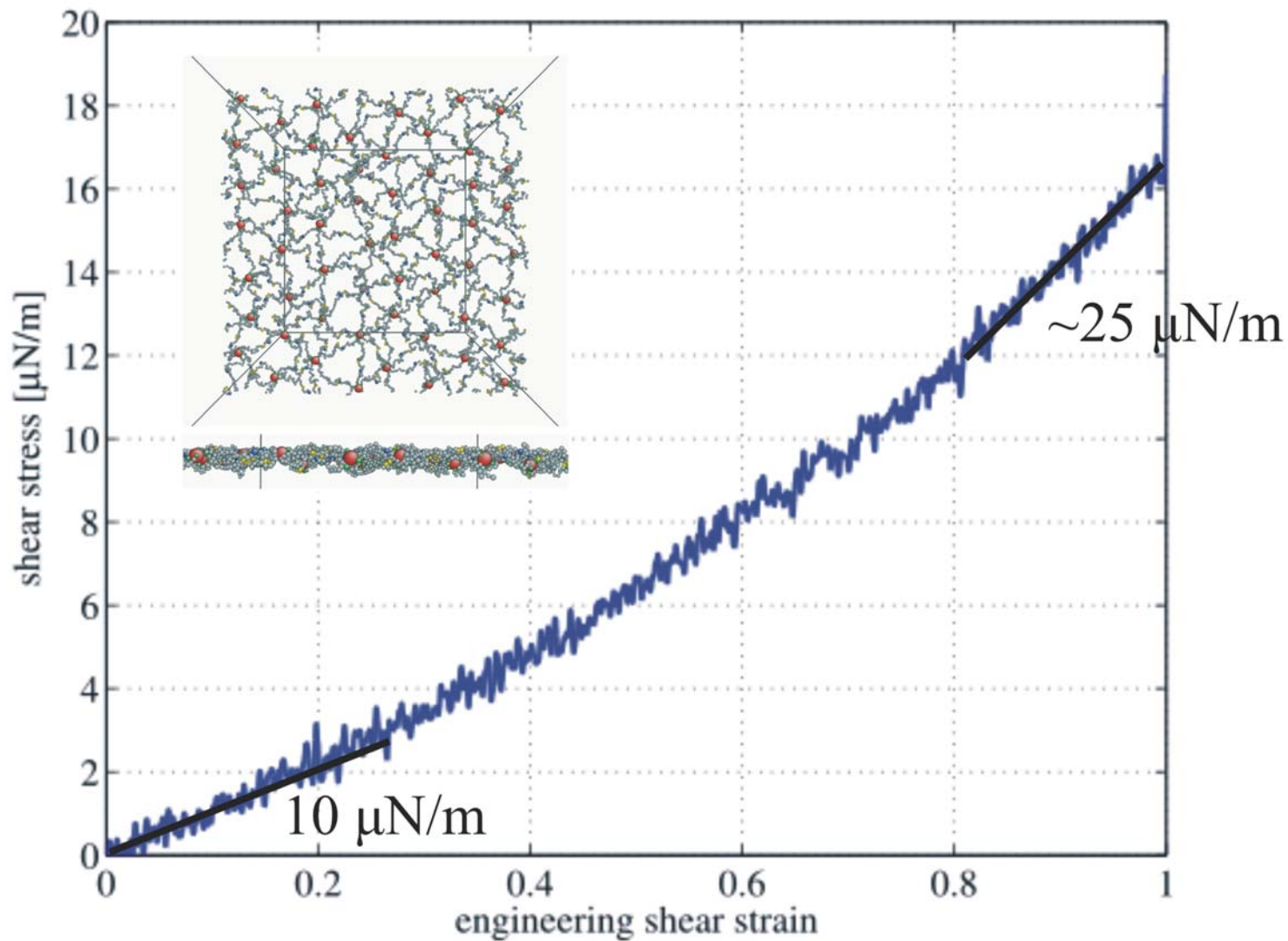


Mohandas and Evans 1994

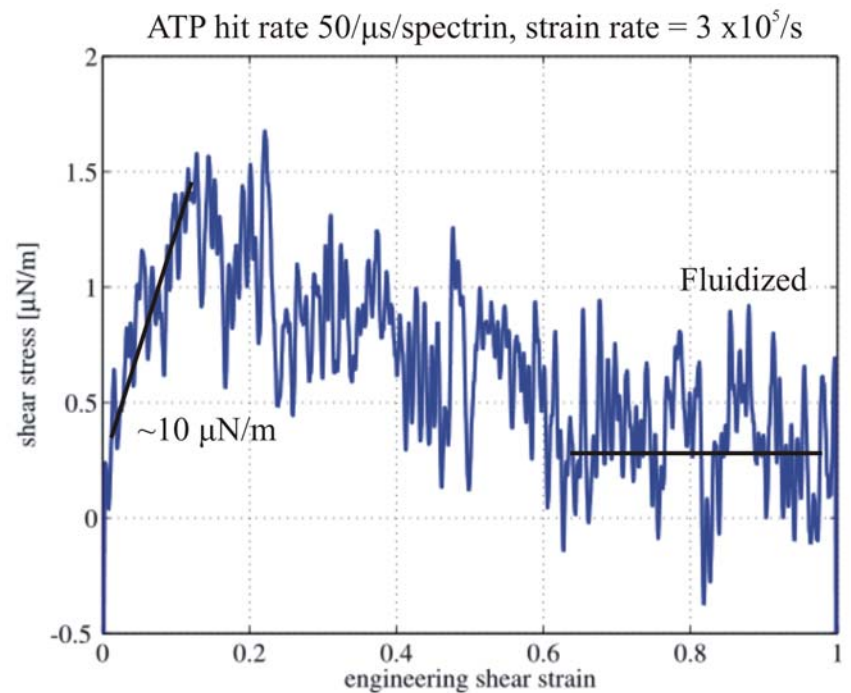
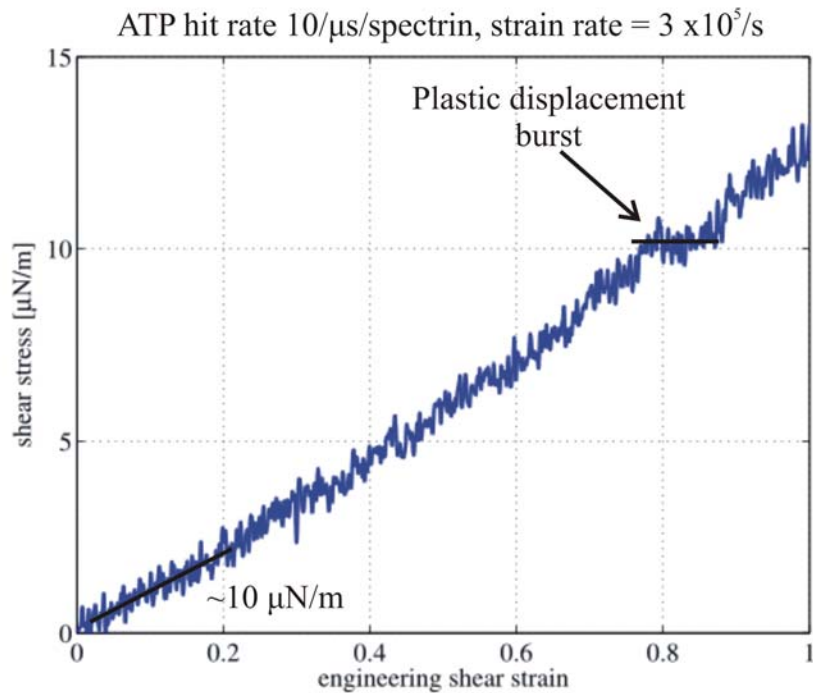
Coarse Grain Molecular Dynamic Modeling



Shear Deformation



Shear Deformation and Promoted Dimer – Dimer Dissociation



Summary

- A minimal CGMD model with *breakable* actin-spectrin junction has been developed, with physically reasonable parameters and behavior.
- ATP hydrolysis is modeled as stochastic kinetic energy transfer. As ATP hit rate rises, we see initiation of plastic displacement excursions, followed by macroscopic yield, and eventually, complete fluidization.
- Practical timescale of CGMD able to simulate recovery.