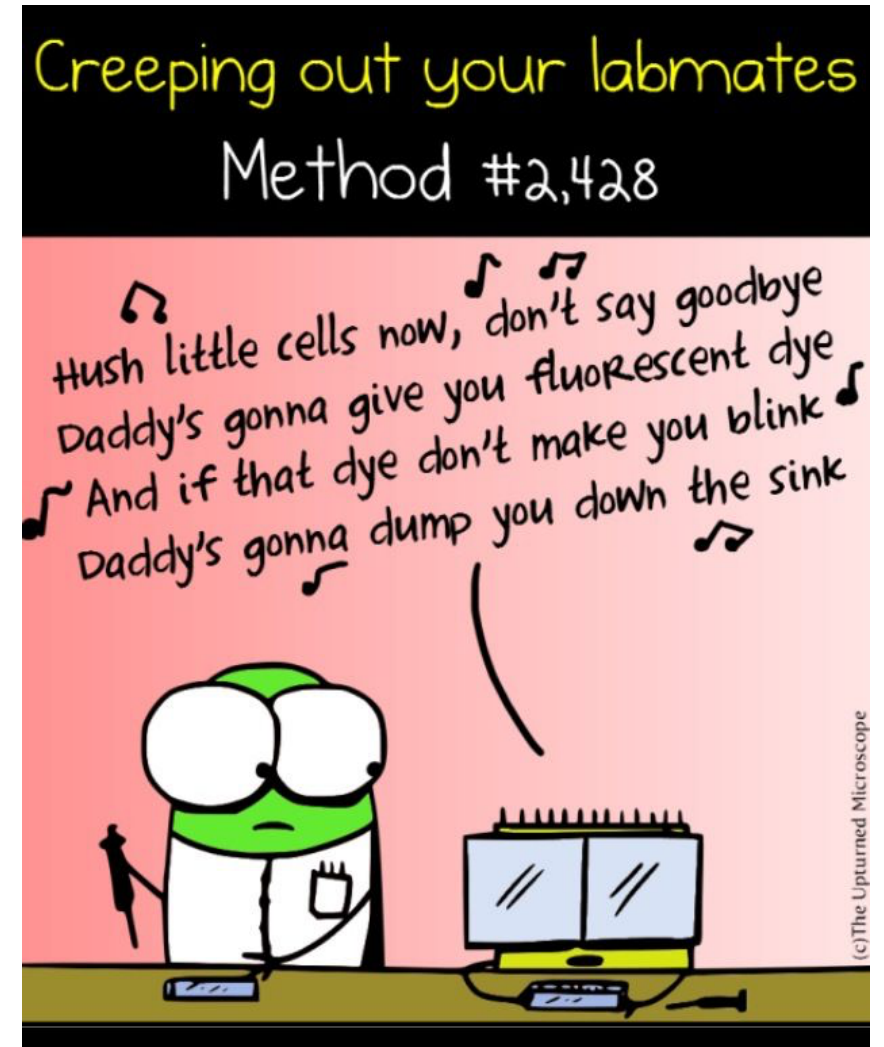
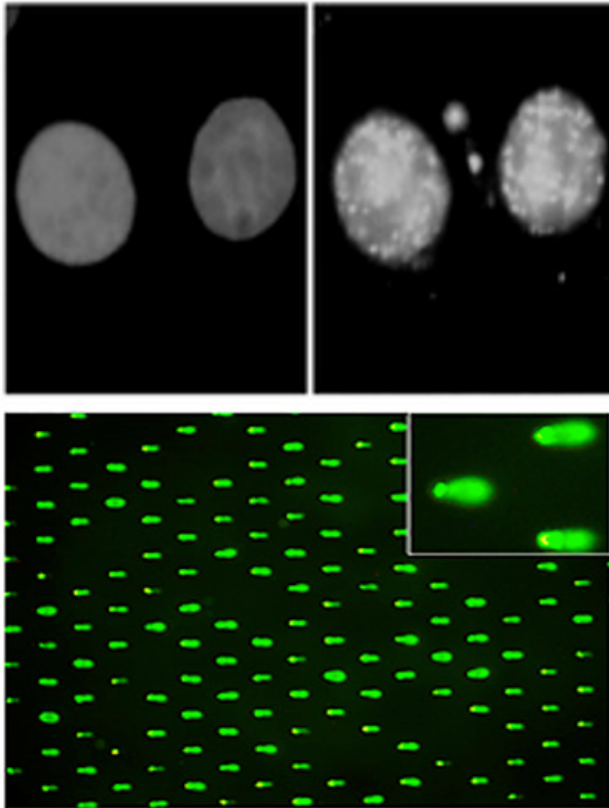


# M1D2: Prepare and treat cells for foci experiment

1. Prelab
2. Experiments for today:
  - Demo of coverslip coating
  - Treat and fix MCL-5 cells for H2AX assay



# Mod1 Overview



## 1. Use repair foci experiment to measure DNA breaks

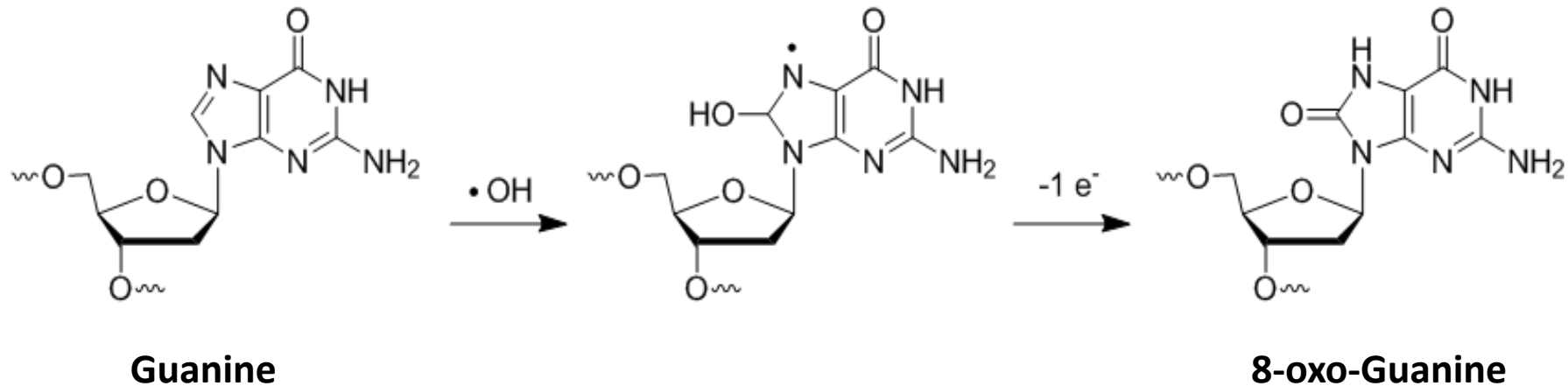
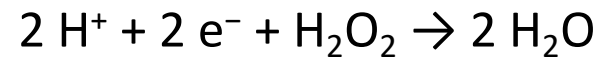
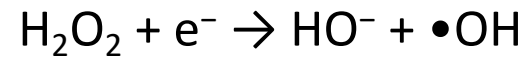
- Examine effect of  $\text{H}_2\text{O}_2$  +/- As on double strand DNA breaks by measuring  $\gamma\text{H2AX}$  foci formation

## 2. Use high-throughput genome damage assay to measure DNA damage

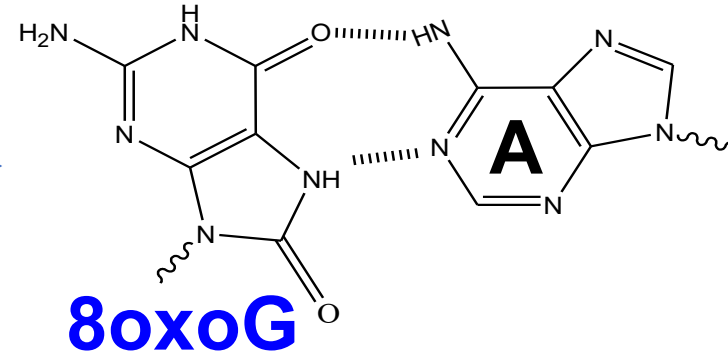
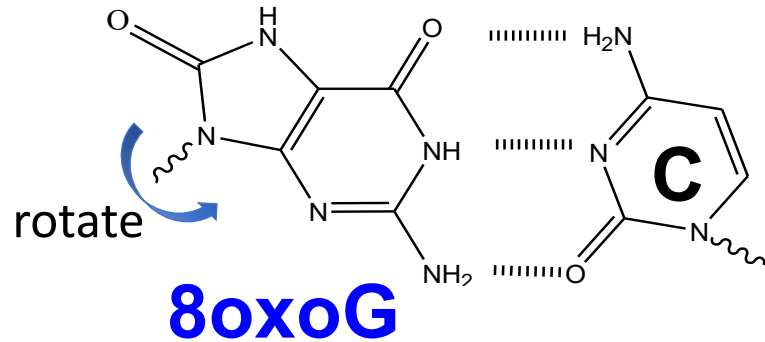
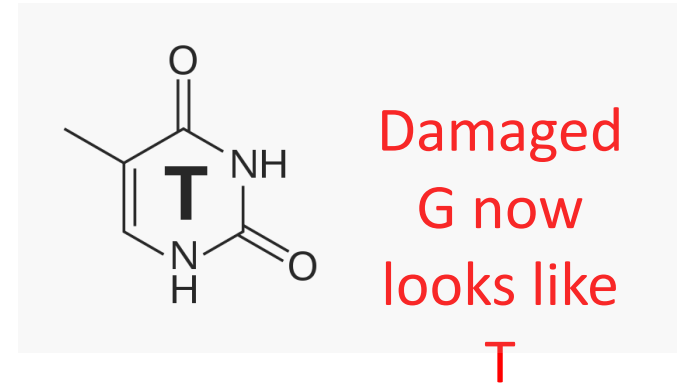
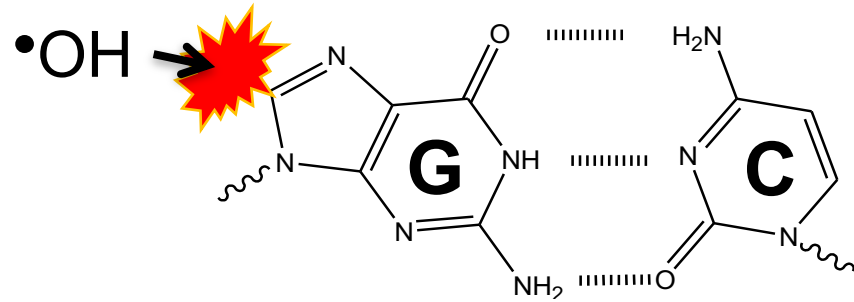
- Measure effects of  $\text{H}_2\text{O}_2$  +/- As on DNA damage by measuring DNA migration in agarose matrix

# How does $\text{H}_2\text{O}_2$ damage DNA?

ROS = \_\_\_\_\_



# How does $\text{H}_2\text{O}_2$ damage DNA?

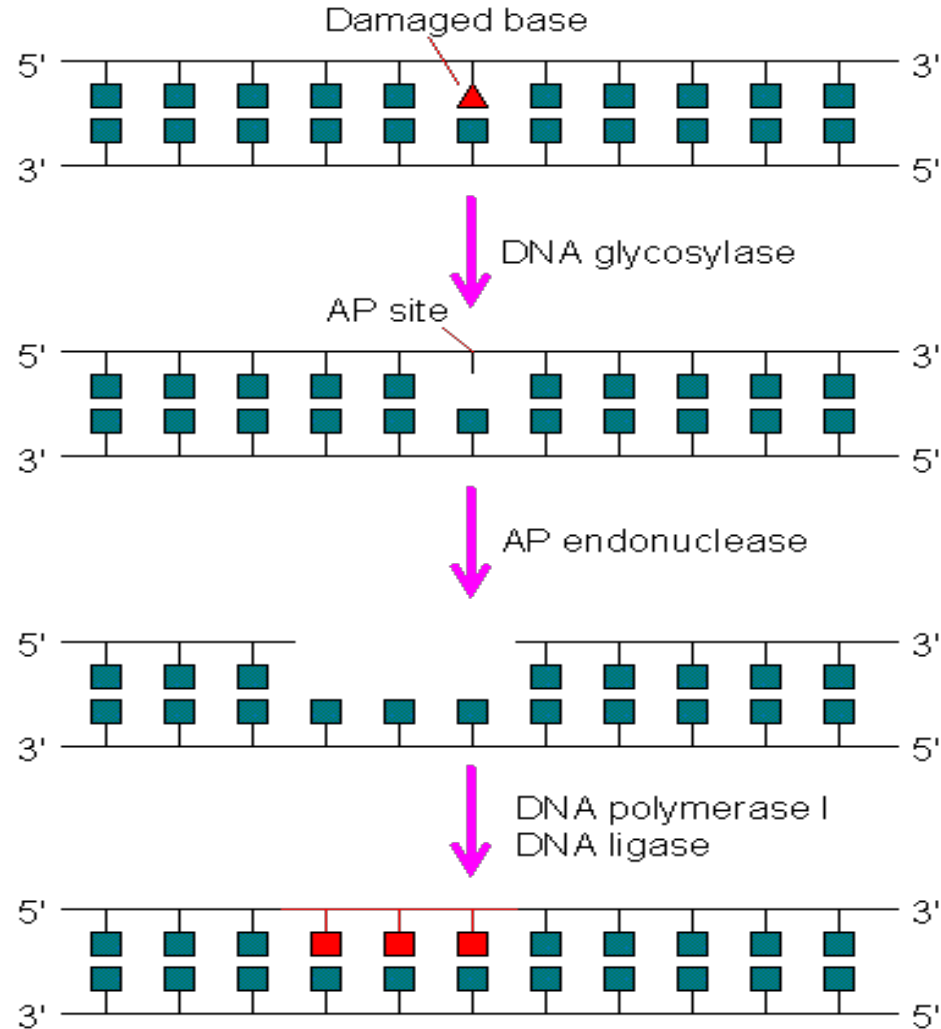


Mutation if replicated



# How do cells repair oxidative DNA Damage?

Typically, the BER pathway



# How do we look at DNA damage in intact cells?

## Look for $\gamma$ -H2AX foci

H<sub>2</sub>O<sub>2</sub> can cause damage to DNA, resulting in a damaged base



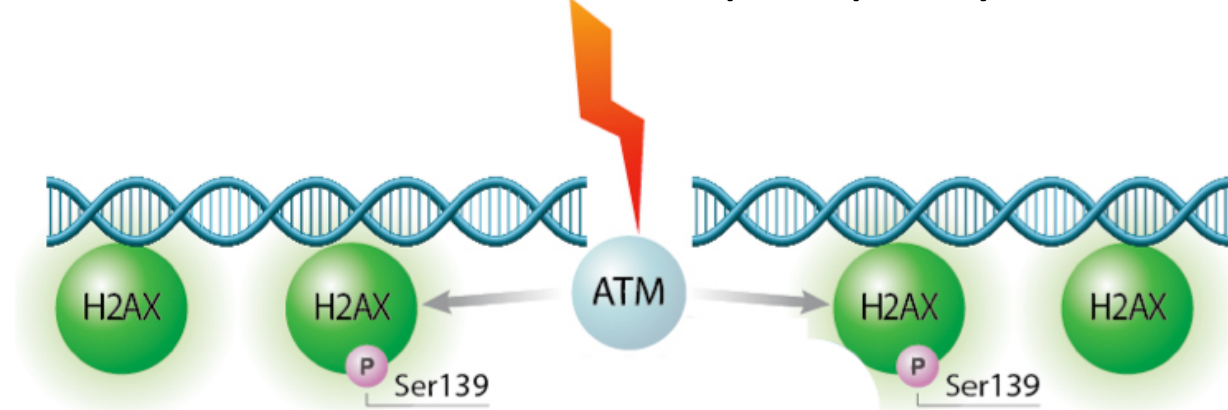
That damage causes a single strand break as cell tries to repair the DNA



Multiple single strand breaks cause double strand breaks



At double strand breaks, ATM phosphorylates the histone H2AX



We can identify the frequency of these DSBs as a measure of DNA damage

# Treatment conditions for this experiment

- Goal: identify any additive effect pretreatment with As has on H<sub>2</sub>O<sub>2</sub> induced DNA damage
  - Treat cells with As for 24 hours, then treat cells with H<sub>2</sub>O<sub>2</sub> for 30 minutes

Experimental Condition

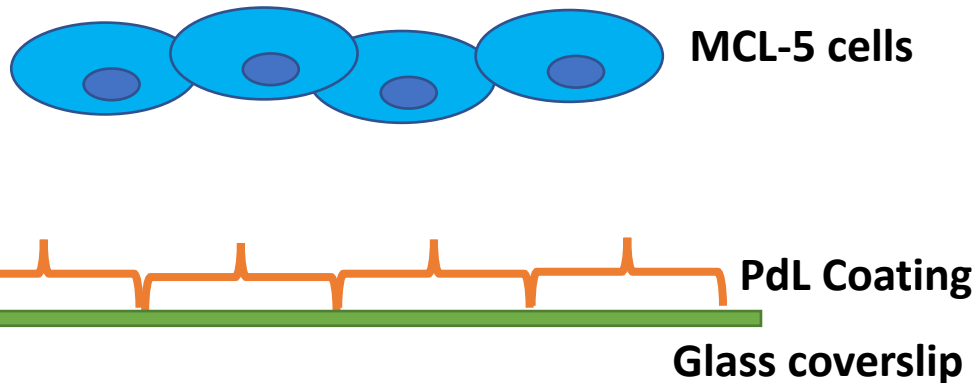
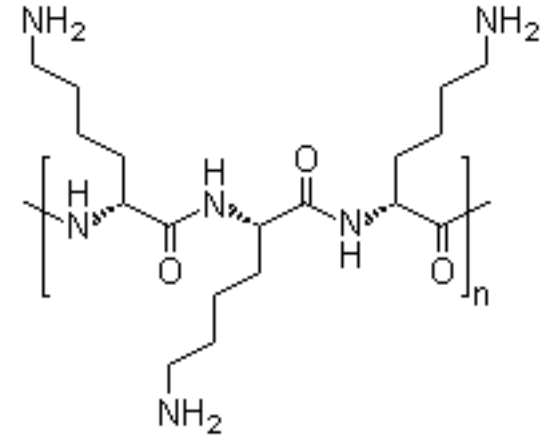
Control Conditions

Our imaging protocol requires cells to adhere to glass coverslips in monolayer

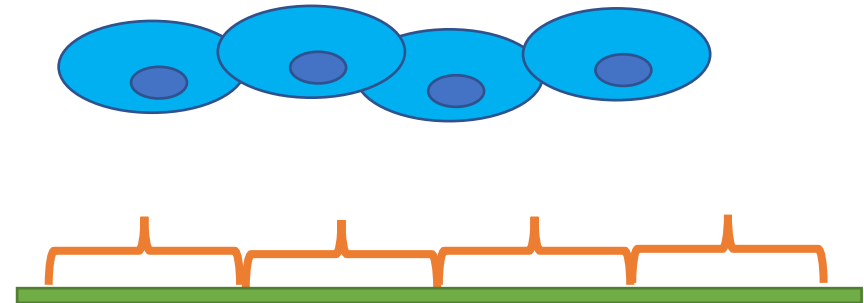
- We have non-adherent cells!
- Must adhere them to coverslips prior to imaging

# Poly-d-Lysine

- Cannot image  $\gamma$ -H2AX foci with cells in suspension
  - Want clear images of nuclei
  - Immobilize cells in a monolayer on glass coverslips
- Many ways to get cells to adhere to glass or plastic
  - ECM molecules (like laminin)
  - Charged polymer molecules (like PdL)
  - Biological substrates (like Polyphenolic Proteins secreted by marine mussels)

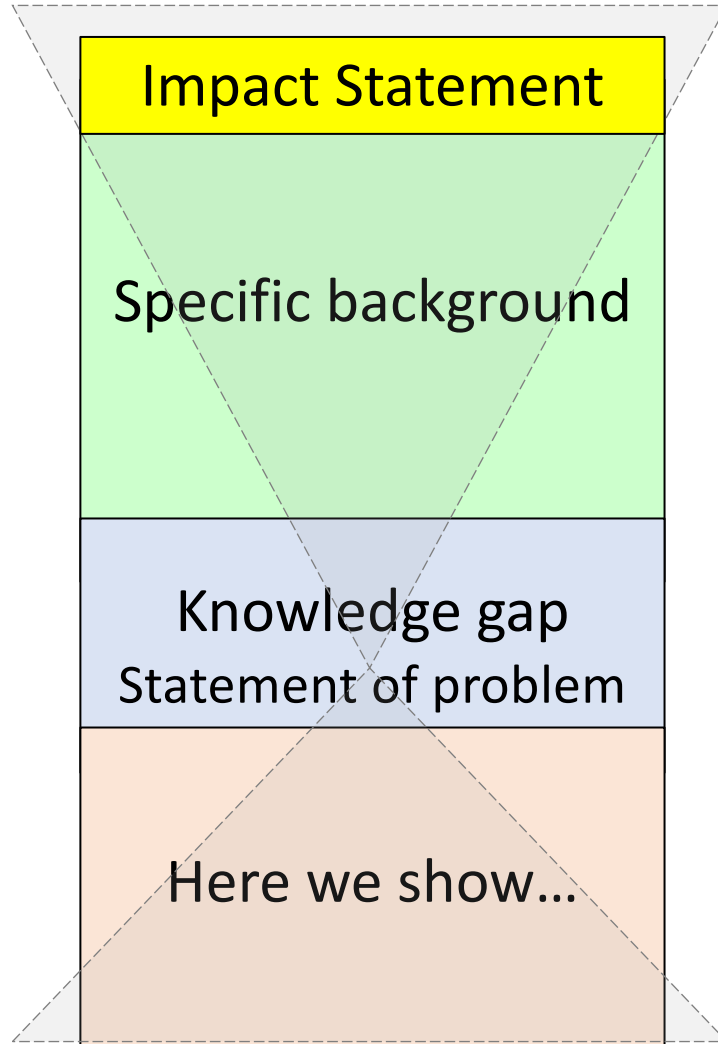


15 min  
Incubation





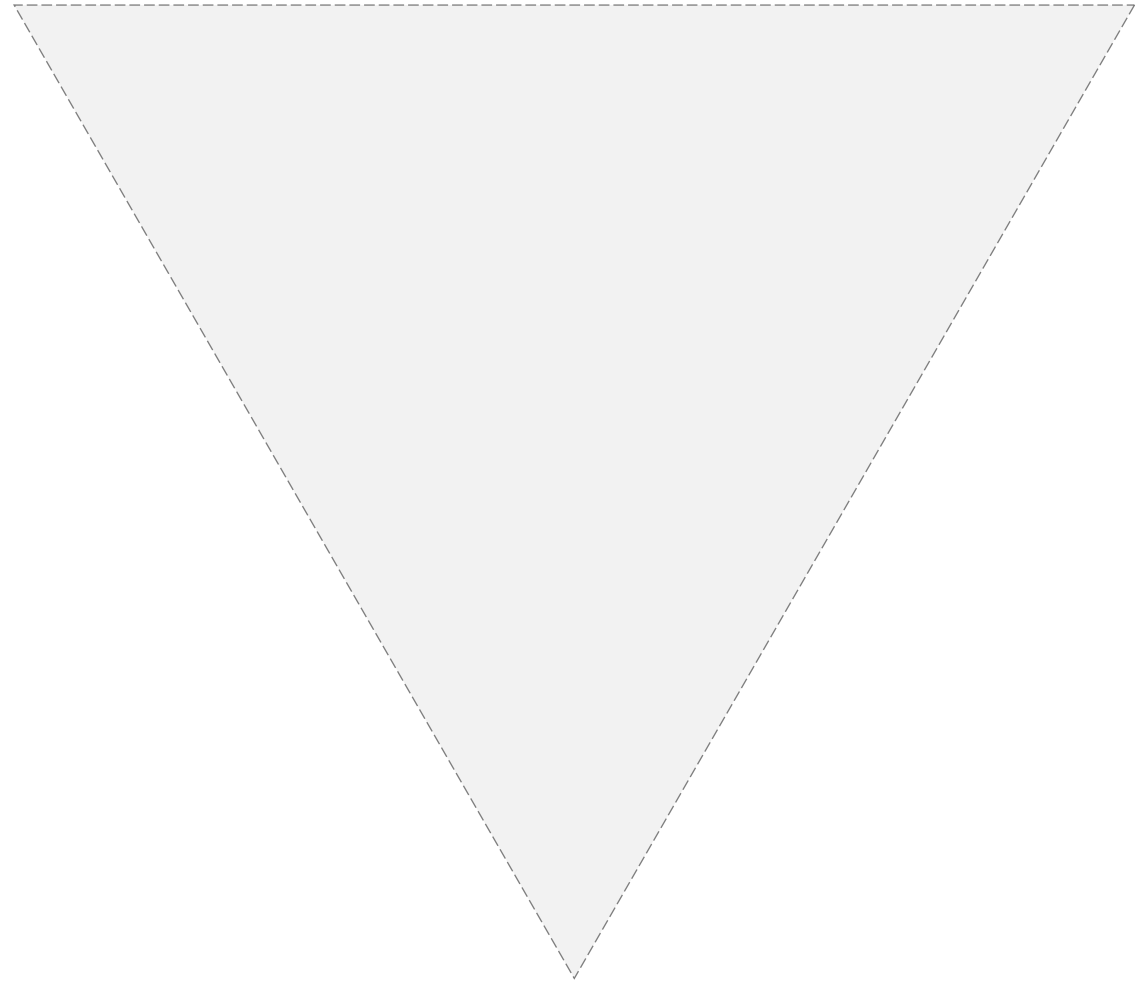
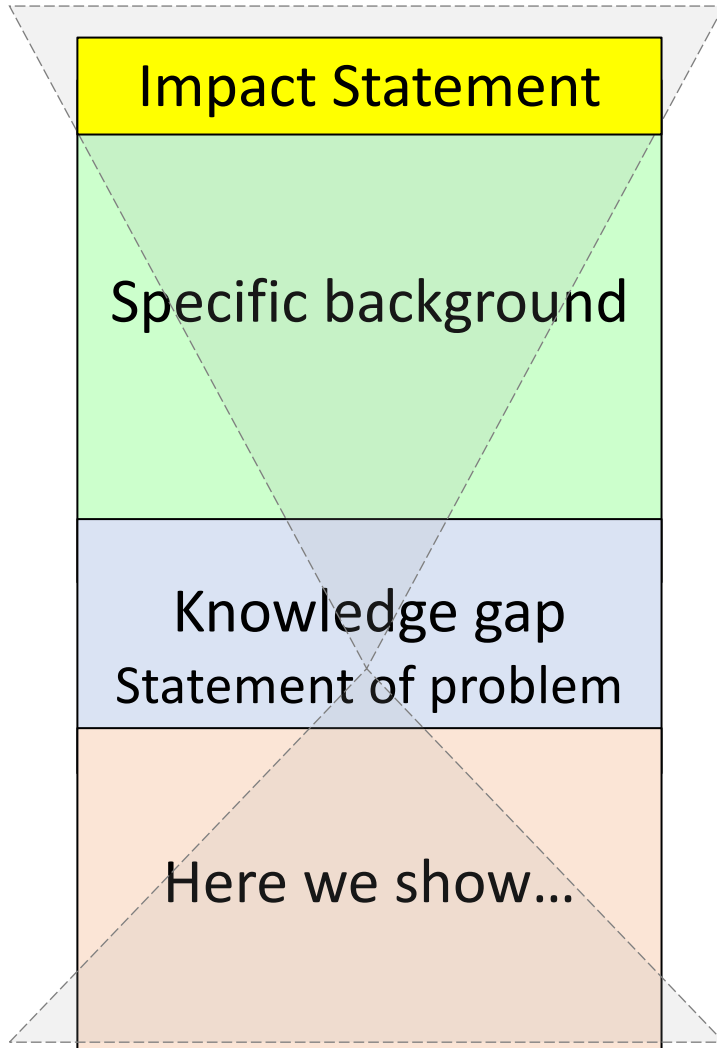
# What goes into a background/motivation section?



- Your research is anchored in a general topic that your audience cares about or could be interested in.
  - Focus on describing previous work in the field
- Specific background connects your project with the general background.
  - Minimum essential information
  - References current work in the field
  - Introduce specific technologies necessary for understanding the project
- The question you address is clearly articulated, connected to the background, and has appropriate scope for the project
  - Give evidence of incompleteness of current understanding, therefore motivating the investigation
  - Include a space holder for your hypothesis (or come up with one)
- A preview of your findings and their implications
  - Light on Methods

# What goes into your introduction?

*Choose one narrative*



# In lab today

1. Work through wiki to treat and fix MCL-5 cells in preparation for  $\gamma$ -H2AX immunofluorescent staining

## M1D3HW

1. Write topic sentences (1<sup>st</sup> sentence) to outline your Data Summary's Background and Motivation section (3-5 sentences total)
  - Remember to include references with summary & why you chose it
2. Read paper on wiki to prepare for in-class discussion
3. Schedule appointment with BE Comm Fellow before M1D5