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# **Module 2 overview**

#### lecture

- 1. Introduction to the module
- 2. Rational protein design
- 3. Fluorescence and sensors

#### lab

- 1. Start-up protein eng.
- 2. Site-directed mutagenesis
- 3. DNA amplification

### **SPRING BREAK**

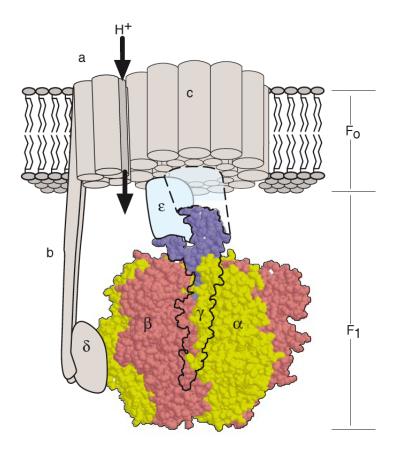
- 4. Protein expression
- 5. Purification and protein analysis
- 6. Binding & affinity measurements
- 7. High throughput engineering

- 4. Prepare expression system
- 5. Induce protein
- 6. Characterize expression
- 7. Assess protein function

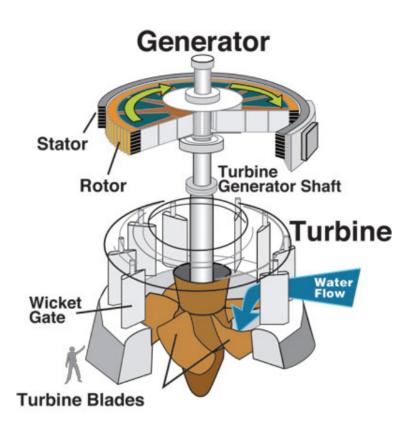
#### Lecture 1: Introduction to the module

- I. Engineering proteins
- II. Pericam: an engineered protein sensor
  - A. Imaging calcium signaling
  - B. Calmodulin and GFP
  - C. Pericam variants
- III. Reengineering Pericam: experimental overview
  - A. Structure-based design
  - B. Protein expression and purification
  - C. Measurements and analysis

#### **Proteins are machines!**



Wang & Oster (1998) *Nature 396:* 279-82

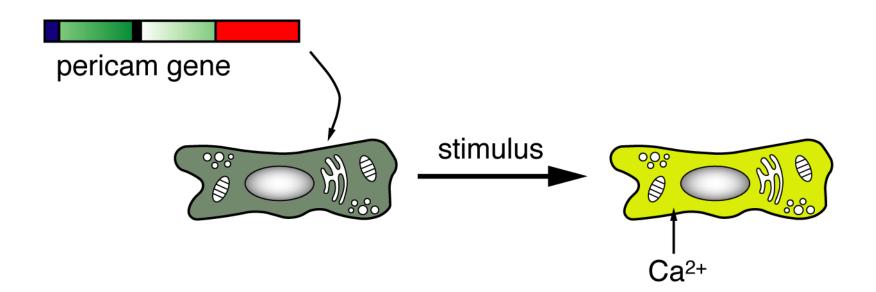


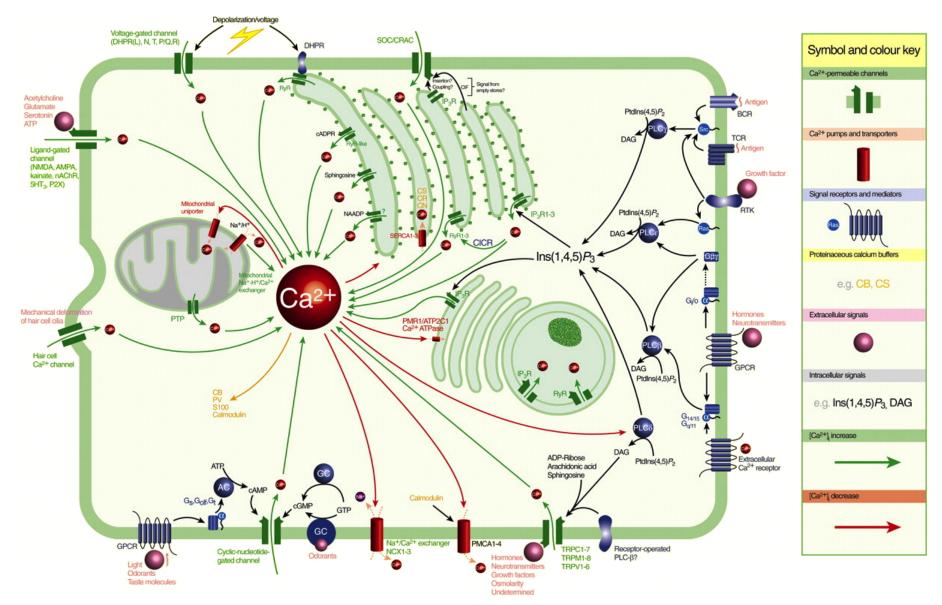
www.symscape.com/node/420



http://www.youtube.com/watch?v=PjdPTY1wHdQ

Pericam: a protein-based machine for measuring [Ca<sup>2+</sup>]





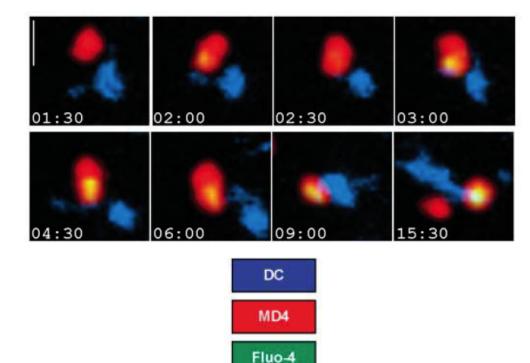
Nowycky et al. (2002) J. Cell. Sci. 115: 3715-3716

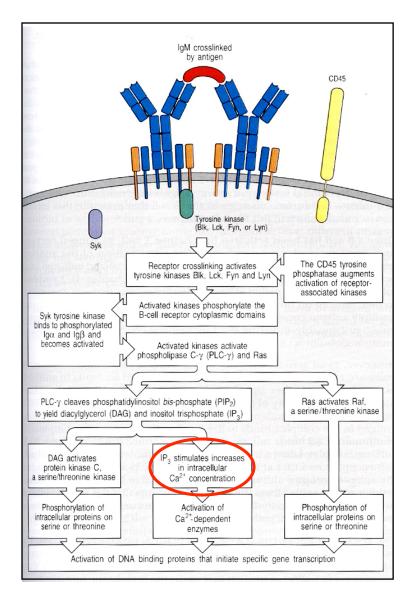
Calcium indicators can be used to detect signaling in individual cells and multicellular ensembles. Two purposes:

- learn what stimuli trigger calcium fluctuations and how calcium behaves in context of an organism or system
- use calcium as a "handle" on cell-cell interaction (*e.g.* neural activity)

J. W. Wang et al. (2003) Cell 112: 271-82.

Calcium is important to cellular signaling in the immune system. Activation of Bcells can be detected by calcium imaging in lymph nodes (Qi *et al.*, 2006, *Science*).





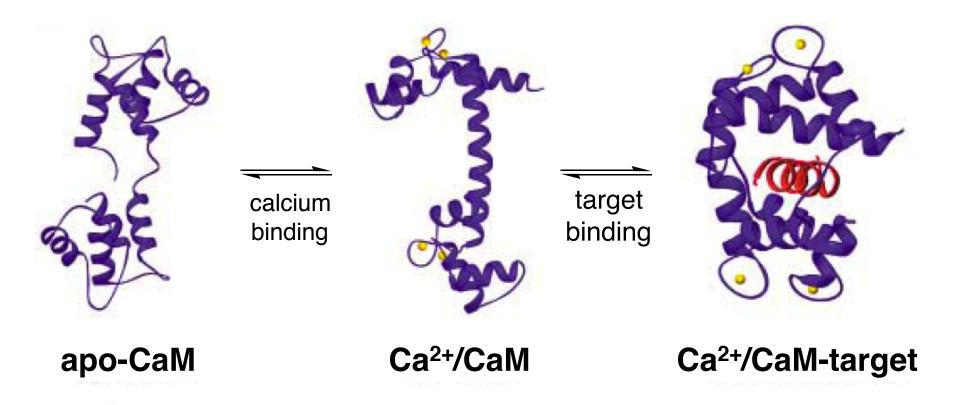


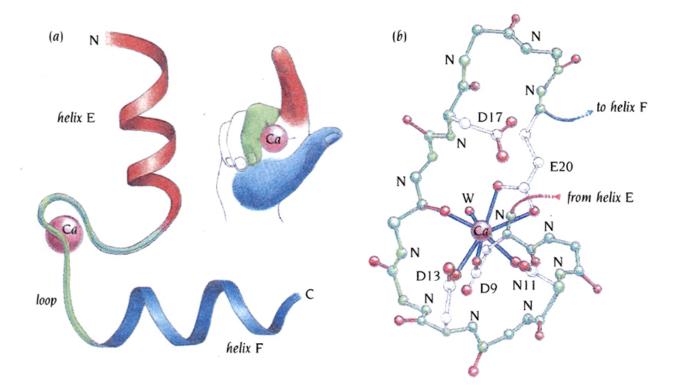
pericam gene

Pericam is a second generation calcium sensor, based on design strategies originally developed by Roger Tsien and colleagues. Tsien won a 2008 Nobel Prize for engineering novel forms of GFP.

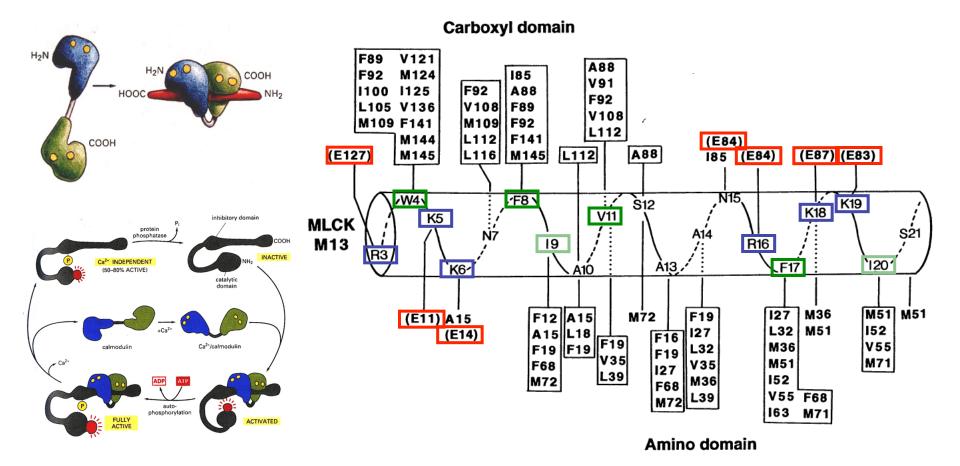
### Calmodulin (CaM) facts and figures

- 16-18 kD (depending on species), ~20 x 40 Å protein
- highly conserved among eukaryotes (vertebrate and yeast calmodulin are functionally interchangeable)
- binds four Ca<sup>2+</sup> ions using EF hand amino acid sequence motifs
- Ca<sup>2+</sup>-CaM binds short segments of target proteins, modulates activity





EF hand binding motif named for E & F helices of the calcium-binding protein parvalbumin; example of helix-loop-helix structure, with calcium bound in the loop Ca<sup>2+</sup>-saturated CaM binds to peptides by "grasping" target sequences, in helical conformation, between N- and C-terminal domains. In many cases, this activates an enzyme by sequestering an inhibitory domain (*e.g.* **M13** from MLCK). Interactions between CaM and targets involve hydrophobic contact area and charge-charge interactions.

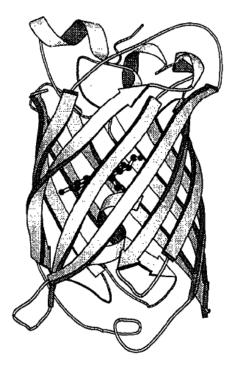


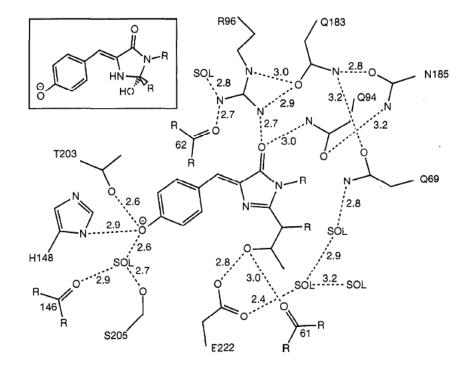
Ikura et al. (1992) Science 256: 632-8.



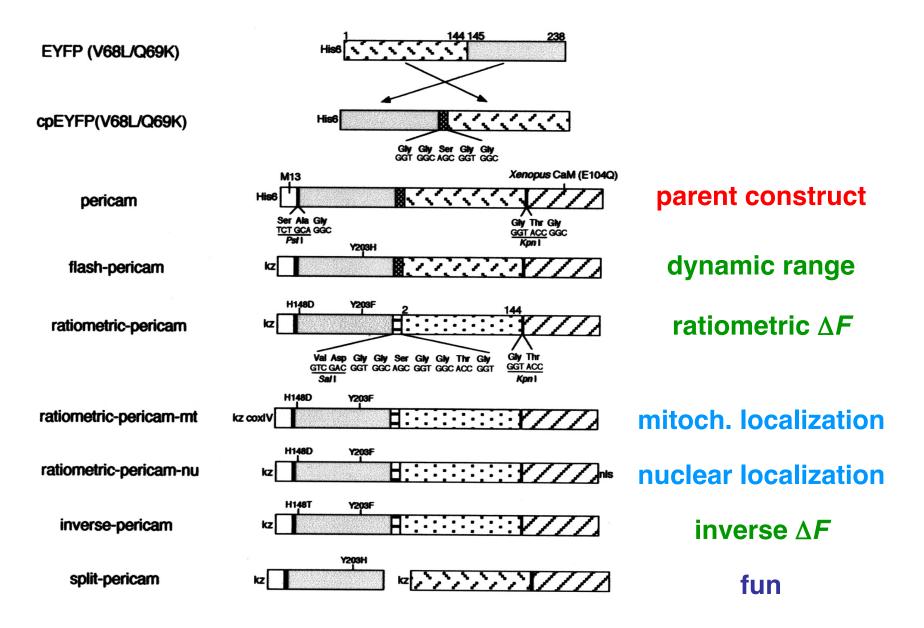
### **Green Fluorescent Protein (GFP)**

from the jellyfish *Aequoria victoria* is a protein fluorophore and component of genetically-encoded calcium indicators. The molecular structure (1996) shows a chromophore formed by spontaneous cyclization and oxidation of three amino acids (Ser/Thr65, Tyr66, and Gly67).

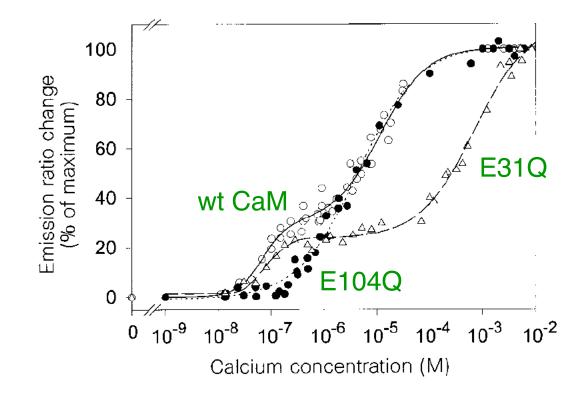




Ormo et al. (1996) Science 273: 1392-5.



Mutations can also affect calcium sensitivity; both  $K_d$  (affinity) and cooperativity (slope/shape of transition) can be affected. Miyawaki *et al.* engineered calcium sensitivity of CaMeleons, a related type of engineered protein calcium sensor:

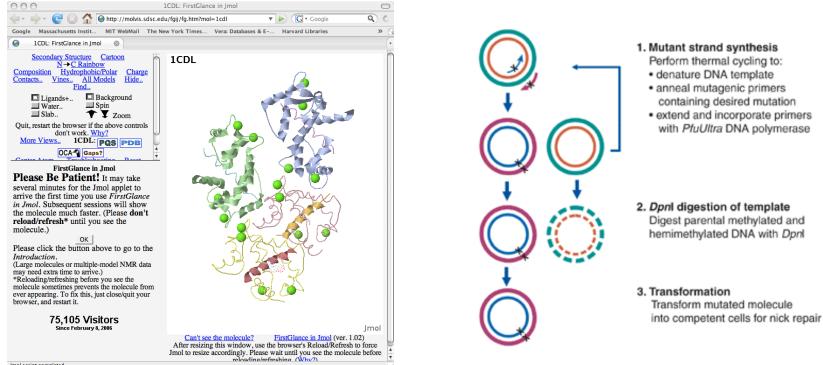


In this module, our goal will be to influence the calcium sensitivity of "inverse pericam."

**Step 1:** Design and implement mutations to affect inverse pericam's calcium sensitivity.

### Skills:

- Use computational tool to look closely at protein structures
- Design primers to make site mutations in the pericam gene
- Perform mutagenesis using PCR

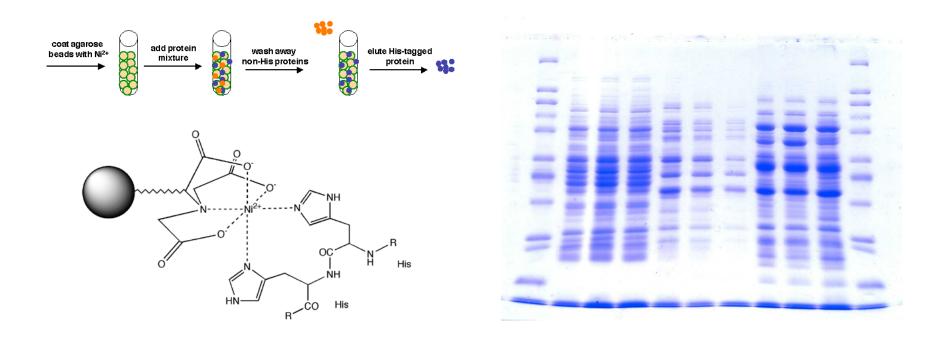


Imol script completed

Step 2: Express and purify mutant inverse pericams for analysis.

### Skills:

- Transform plasmid DNA into E. coli
- Induce protein expression using IPTG
- Purify mutant pericams using affinity-based separation
- Assay protein expression and purity using SDS-PAGE



Step 3: Analyze calcium titration behavior of mutant pericams.

## Skills:

- Perform fluorescence assays to measure calcium binding
- Use software to extract binding parameters from the data
- Pool data from across the class to observe patterns

