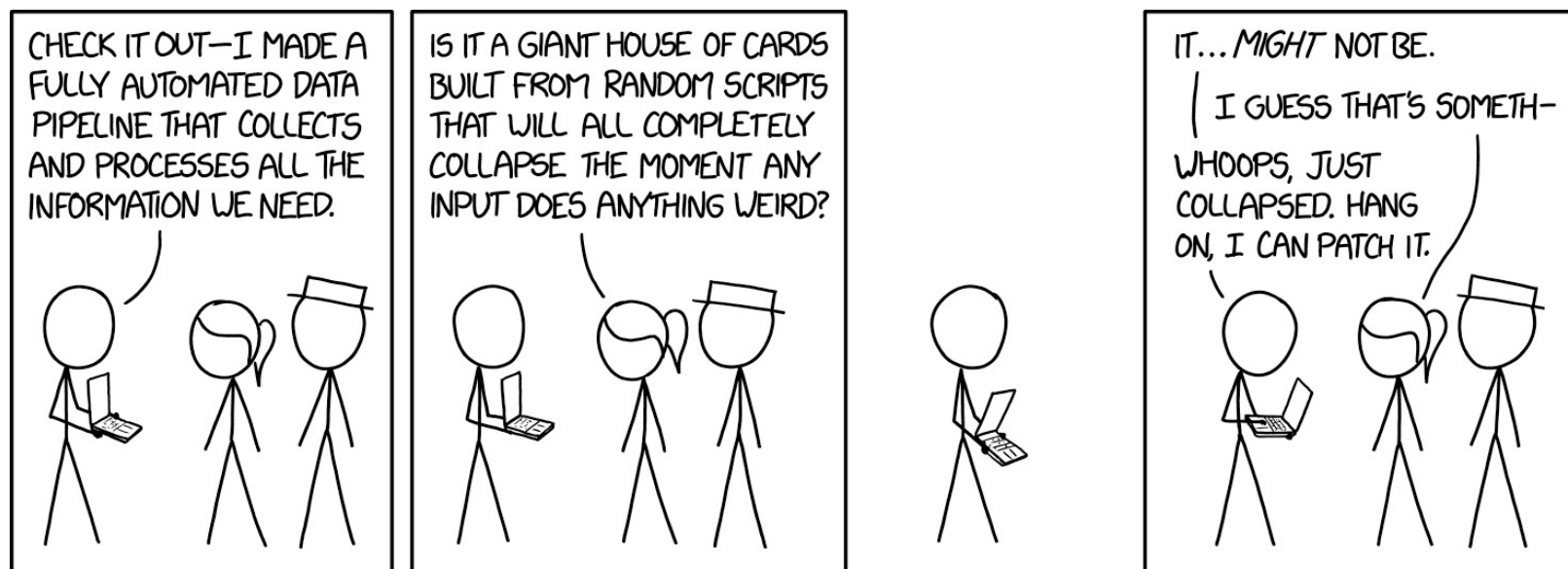


M2D1: Review small molecule microarray (SMM) experiment and results

1. Prelab
2. Walk through SMM
3. Examine chemical structure of hits



Due dates are approaching!

Mod2

- **Journal Club presentation** (15%)
 - Individual
 - Presentations on 10/26 & 10/28
- **Research article** (20%)
 - Individual
 - due 11/22
- Laboratory quizzes (collectively 5%)
 - M2D4 and M2D7
- Notebook (collectively 5%)
 - one entry will be graded by Ben 24 hr after M2D7
- Blog (part of 5% Participation)
 - due 10/30 & 11/23 via Slack channel

Wrap-up Mod 1

- Data summary due Wed. 10/13 @10pm
 - via Stellar
- Research talk due Sat. 10/16 @10pm
 - via GMAIL
- Data summary revisions due Sat. 10/14 @10pm
 - via Stellar
- Blog due 10/18 @ 10pm
 - via Slack

Module 2 Roadmap

Determine putative PF3D7_20109-F21 binders via high throughput screening (SMM)



Create plasmid of PF3D7_20109-F21 to use in validation assays



Express PF3D7_20109-F21 (from plasmid) in bacteria and purify protein



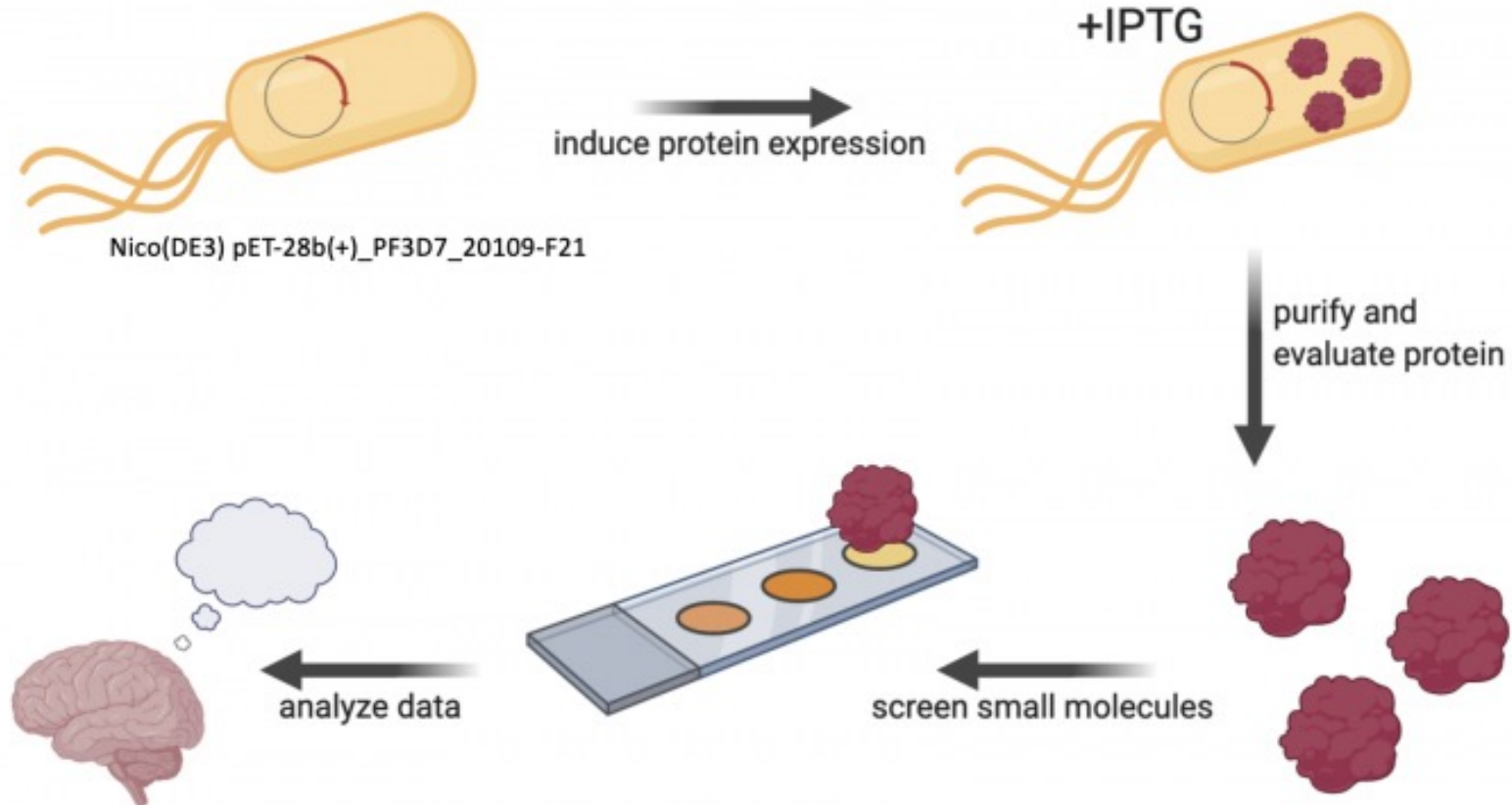
Assess purity and concentration of purified protein



Use purified protein to validate binding of small molecules identified in SMM

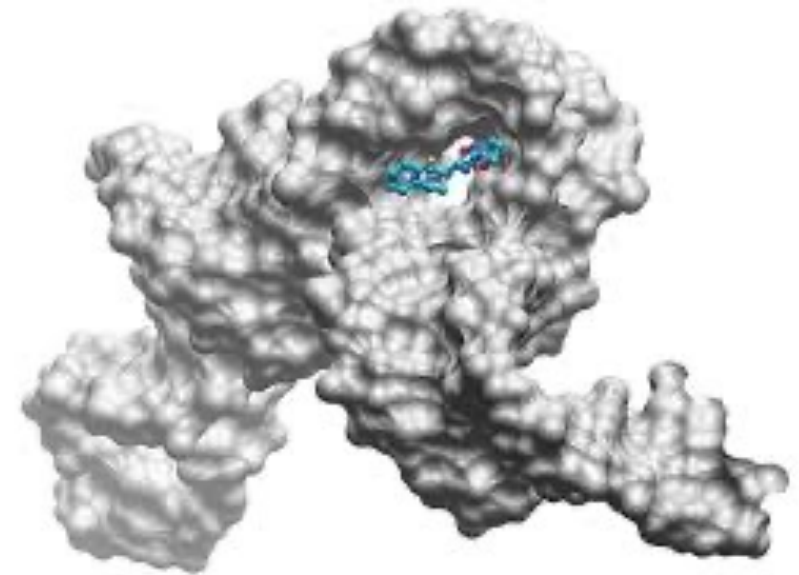
SMM Overview

Research goal: Identify small molecules that bind to the PF3D7_20109-F21 protein in *Plasmodium falciparum* using small-molecule microarray

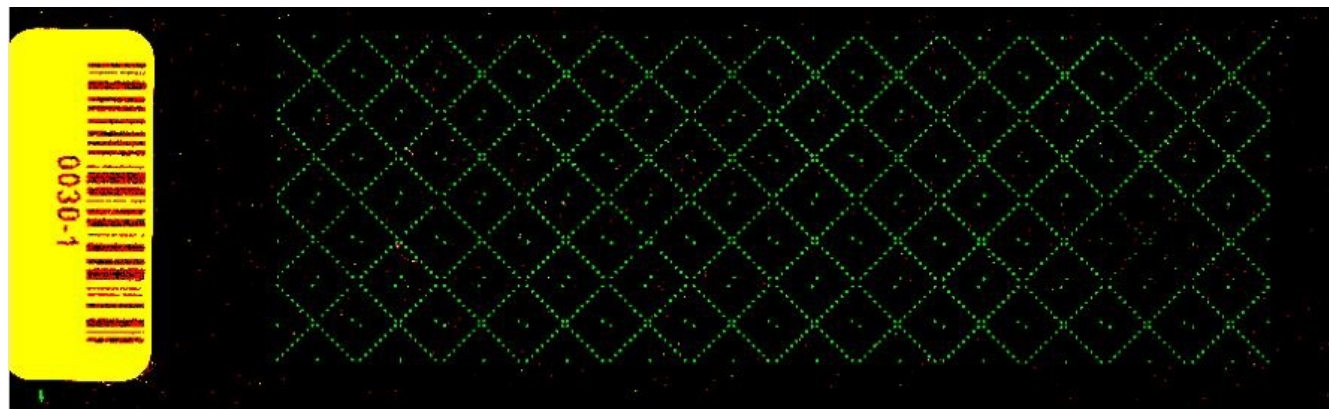
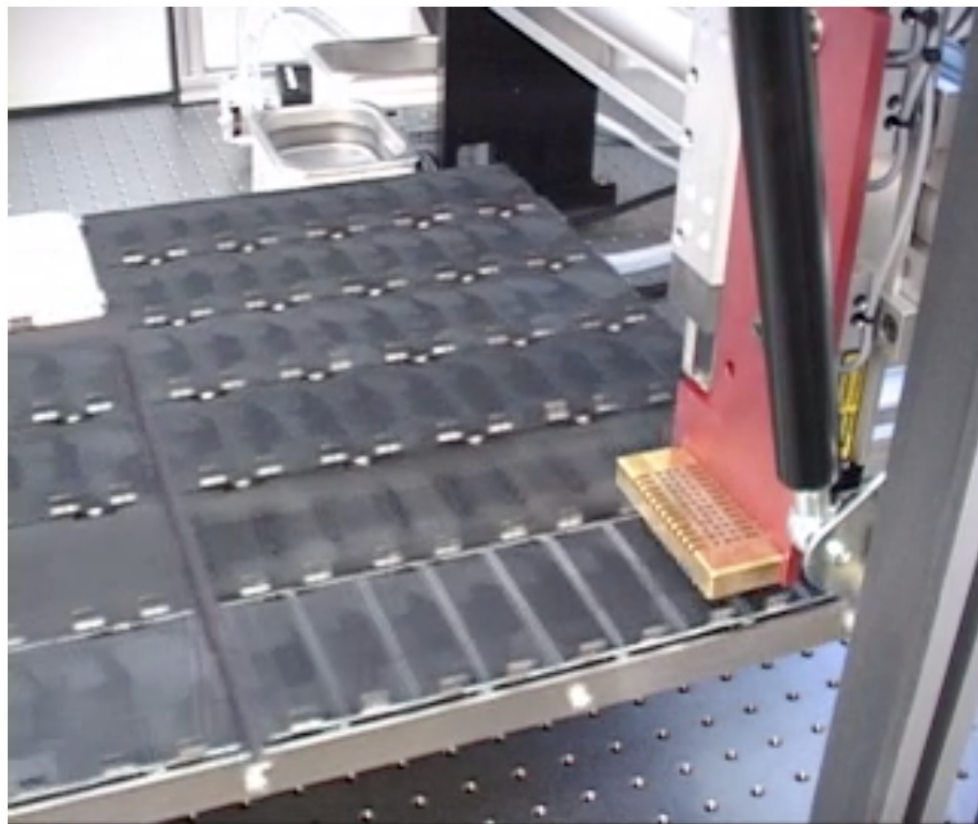


Why are we taking this approach?

- High throughput assays are useful in screening potential therapeutic targets
 - Allows unbiased exploration of potential therapeutics
 - Allows examination of targets with limited information
- Small molecules
 - $M_w < 500$ Da
 - Natural or synthetic
 - Frequently comprised of Carbon/Nitrogen/Oxygen



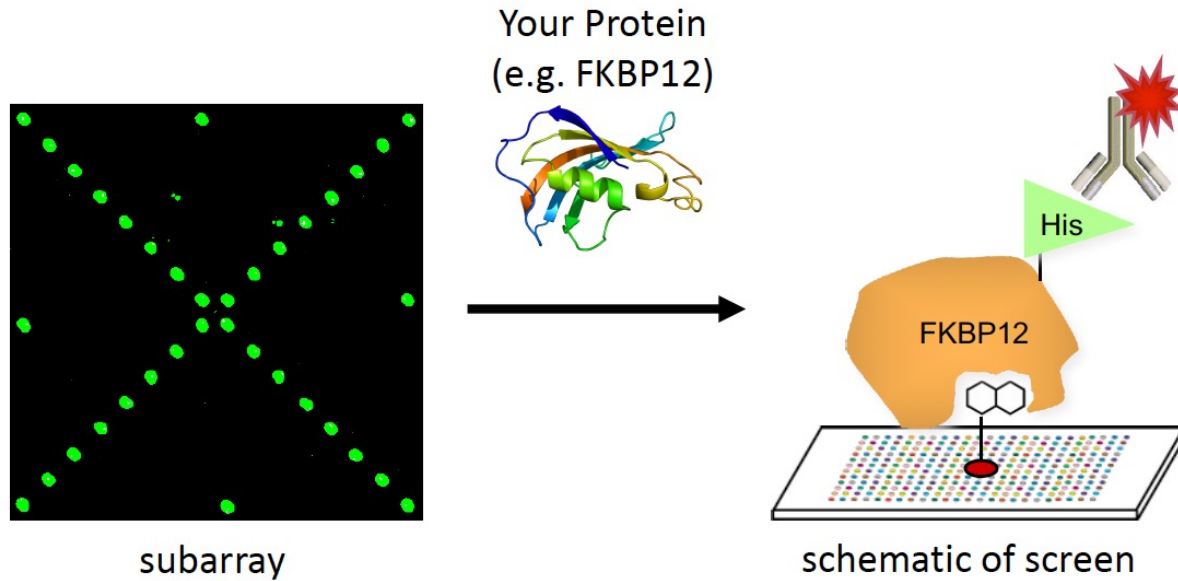
Small Molecule Microarray (SMM)



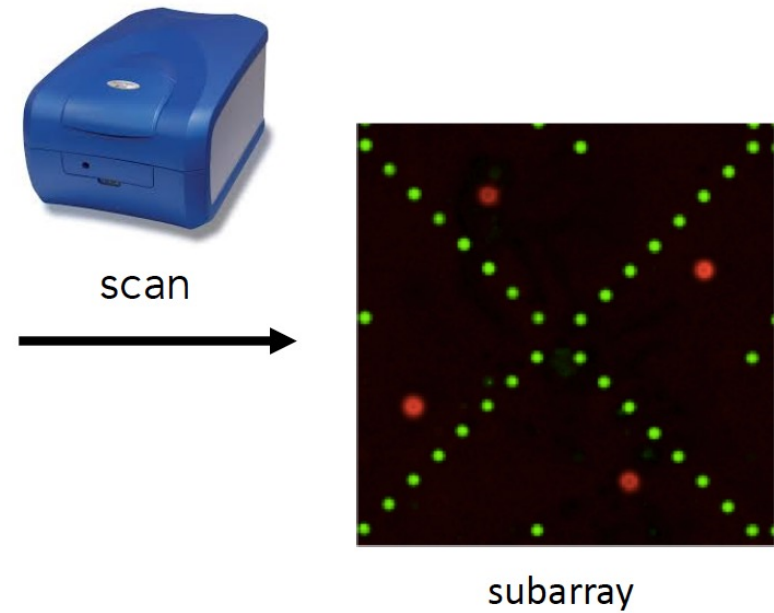
- Each slide contains ~12,000 spots
 - ~4,200 small molecules / ligands (in duplicate = ~8,400)
 - Fluorescein sentinel spots
 - DMSO negative control spots

SMM workflow

SMM Screen

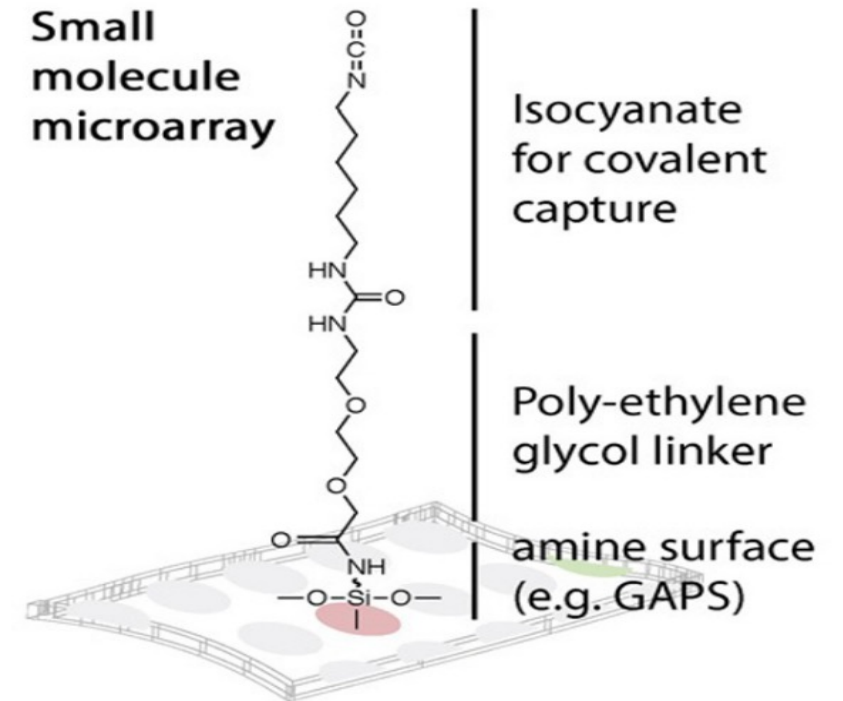


Data Acquisition



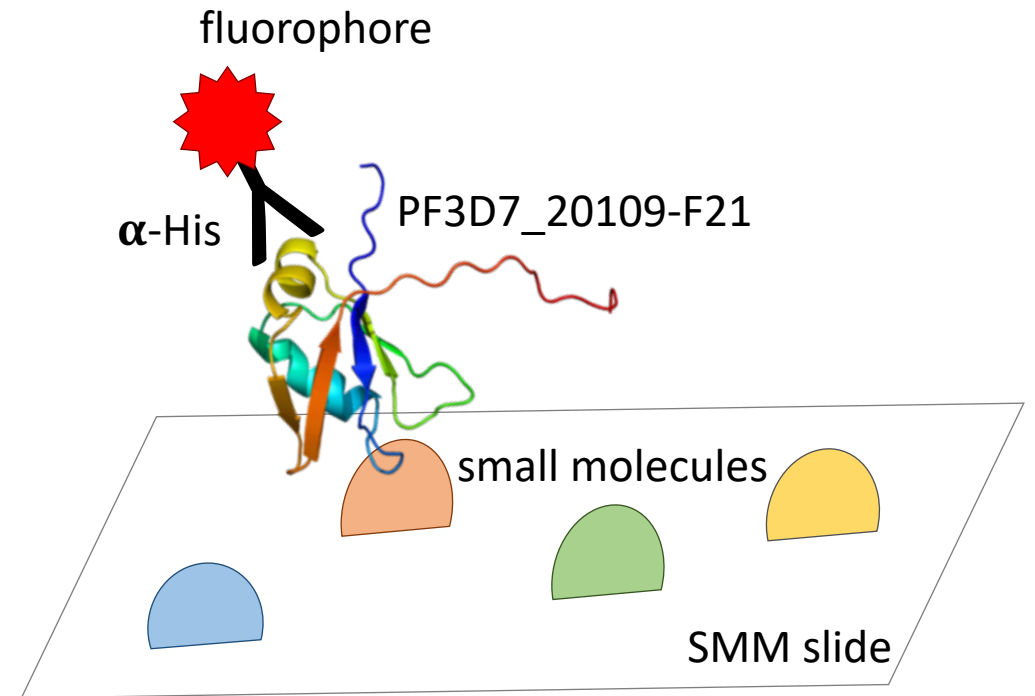
SMM slide preparation

- Gamma-aminopropylsilane (GAPS) slide coated with polyethylene glycol (PEG) spacer
- PEG coupled to 1,6-diisocyanatohexane to generate isocyanate-functionalized slide
- Isocyanate able to react with nucleophilic functional groups



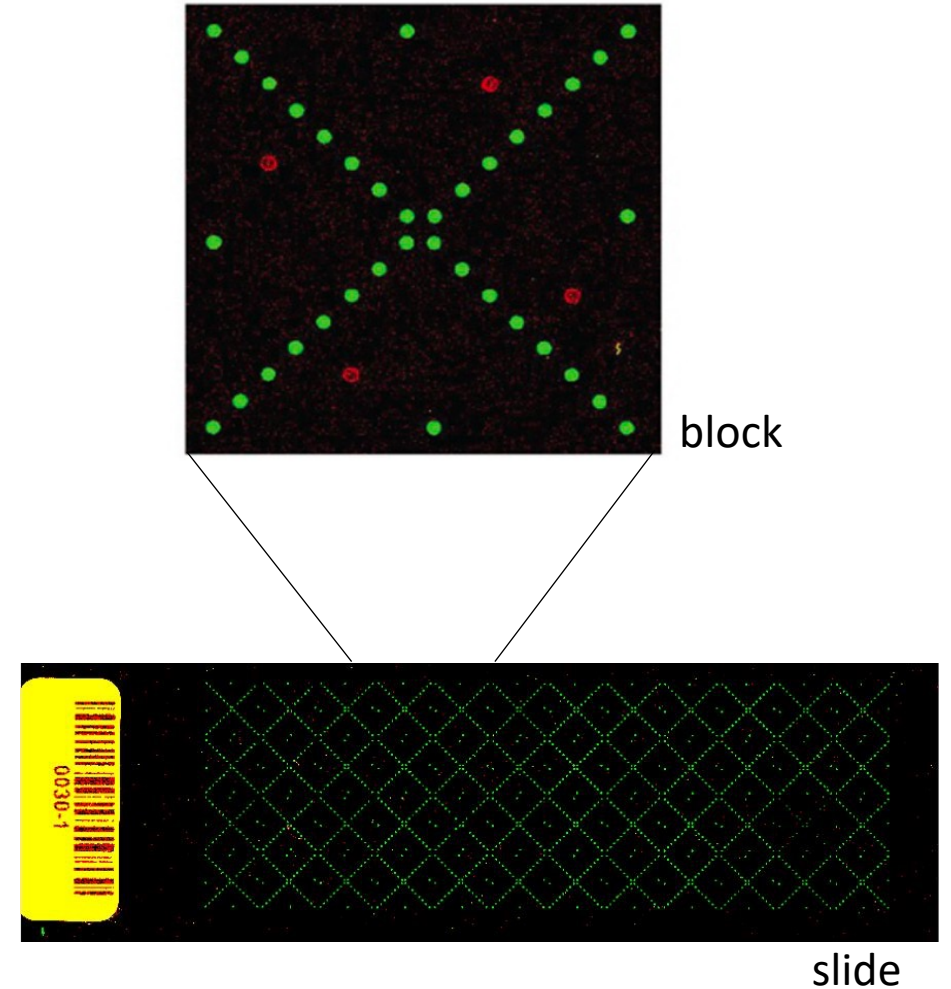
How do we screen for ligands that bind PF3D7_20109-F21?

- Incubate the SMM slide with 3ml of our purified PF3D7_1351100
- Wash away unbound protein
- Incubate SMM slide with AlexaFlour 647 anti-His antibody
- Wash away excess antibody
- Store for scanning



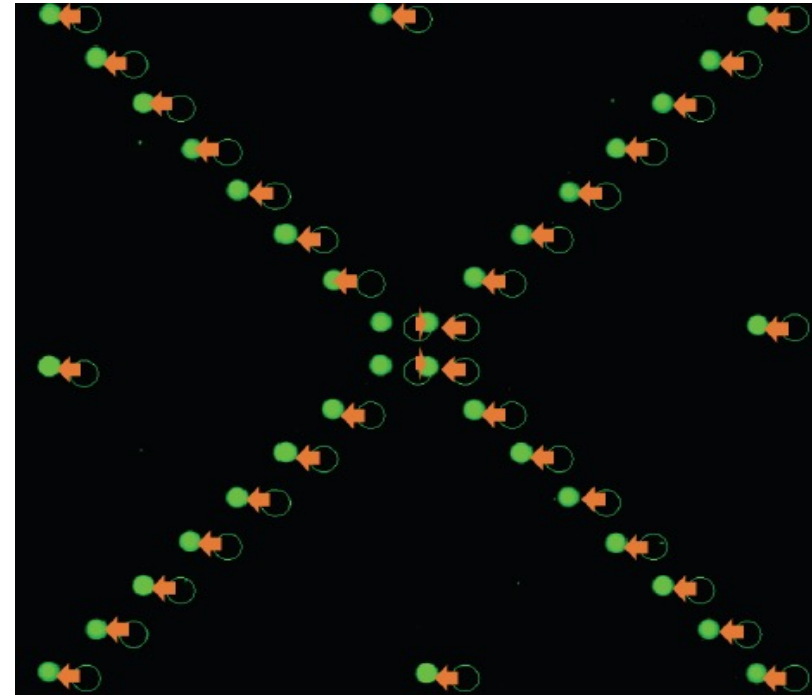
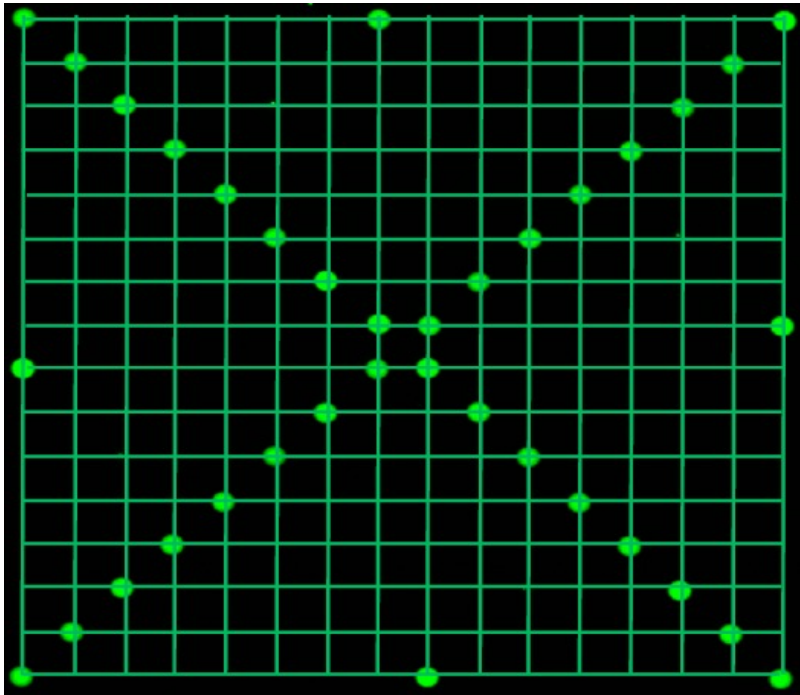
Workflow for SMM data analysis

1. Align spots using fluorescence on 532 nm channel (sentinel spots)
↓
2. Quantify fluorescence on 635 nm channel
↓
3. Identify 'hits' with improbably high fluorescence
↓
4. Complete 'by eye' analysis of putative hits to manually remove false positives



Align SMM using sentinel spots

- Slides are printed in block patterns (16 rows x 16 columns)
- Each ligand spot is identifiable via intersecting lines between sentinels



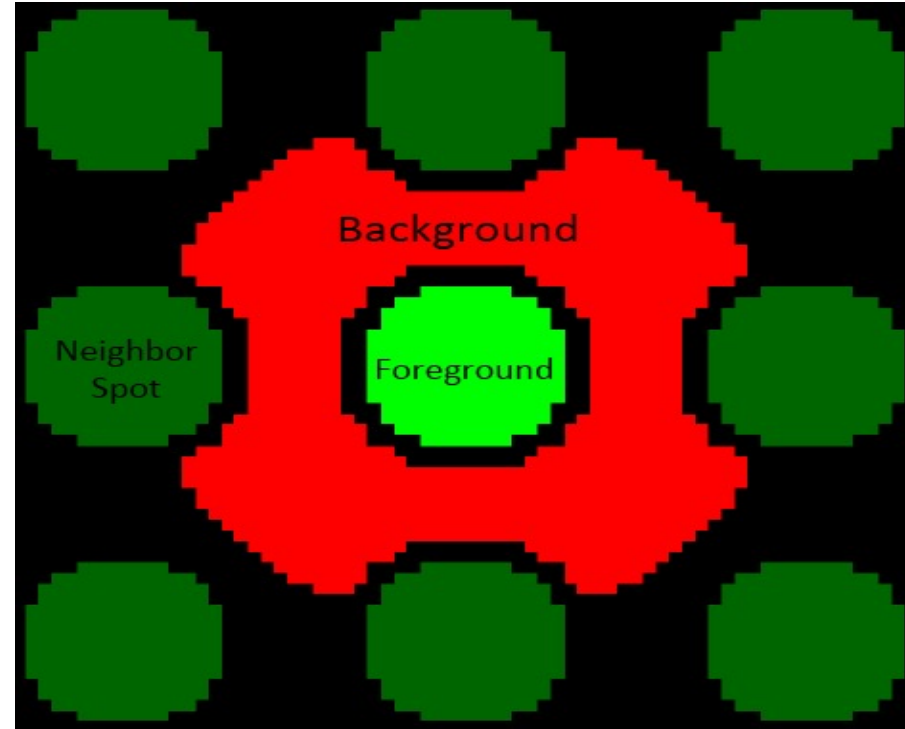
Spots are represented by an array of numerical values

- Each pixel is represented by a number that indicates intensity of the signal
- Computational analysis used to define 'hits'

| | | | | | | | | | | | | | | | | | | |
|---|---|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|---|---|
| 4 | 3 | 4 | 4 | 3 | 2 | 3 | 4 | 3 | 5 | 4 | 6 | 3 | 3 | 3 | 2 | 3 | 2 | 2 |
| 3 | 5 | 4 | 3 | 3 | 3 | 5 | 6 | 7 | 8 | 5 | 6 | 4 | 4 | 4 | 3 | 3 | 3 | 3 |
| 3 | 3 | 3 | 3 | 4 | 8 | 12 | 92 | 275 | 311 | 256 | 61 | 11 | 6 | 3 | 3 | 3 | 3 | 4 |
| 4 | 3 | 3 | 4 | 8 | 173 | 625 | 818 | 823 | 856 | 815 | 831 | 568 | 136 | 9 | 5 | 4 | 4 | 3 |
| 5 | 3 | 4 | 8 | 273 | 830 | 814 | 835 | 873 | 890 | 836 | 857 | 818 | 771 | 201 | 9 | 6 | 2 | 2 |
| 3 | 4 | 7 | 175 | 780 | 805 | 877 | 941 | 936 | 920 | 973 | 921 | 842 | 819 | 714 | 125 | 6 | 3 | 2 |
| 4 | 4 | 29 | 568 | 868 | 867 | 905 | 909 | 936 | 994 | 954 | 931 | 963 | 875 | 813 | 490 | 15 | 5 | 4 |
| 4 | 5 | 131 | 754 | 852 | 906 | 958 | 920 | 963 | 923 | 917 | 904 | 951 | 930 | 851 | 716 | 95 | 6 | 3 |
| 4 | 5 | 229 | 796 | 879 | 924 | 934 | 923 | 962 | 961 | 993 | 993 | 945 | 989 | 867 | 780 | 162 | 6 | 4 |
| 3 | 7 | 254 | 827 | 879 | 965 | 949 | 960 | 982 | 926 | 918 | 955 | 927 | 984 | 872 | 765 | 204 | 7 | 3 |
| 4 | 5 | 175 | 808 | 883 | 996 | 951 | 998 | 935 | 976 | 971 | 940 | 922 | 961 | 872 | 804 | 132 | 4 | 4 |
| 4 | 4 | 57 | 666 | 859 | 968 | 999 | 947 | 977 | 985 | 916 | 928 | 960 | 974 | 841 | 678 | 62 | 4 | 4 |
| 4 | 3 | 11 | 406 | 839 | 897 | 915 | 930 | 946 | 993 | 914 | 911 | 977 | 900 | 830 | 359 | 10 | 3 | 4 |
| 3 | 2 | 5 | 60 | 624 | 830 | 890 | 973 | 903 | 921 | 912 | 930 | 881 | 850 | 613 | 54 | 6 | 3 | 3 |
| 3 | 4 | 4 | 7 | 92 | 602 | 873 | 856 | 882 | 913 | 887 | 885 | 842 | 589 | 82 | 7 | 4 | 3 | 3 |
| 3 | 4 | 3 | 4 | 5 | 23 | 266 | 697 | 838 | 828 | 837 | 667 | 261 | 21 | 5 | 4 | 4 | 5 | 4 |
| 3 | 3 | 4 | 4 | 4 | 6 | 9 | 12 | 27 | 49 | 28 | 11 | 9 | 7 | 5 | 3 | 3 | 4 | 3 |
| 3 | 5 | 3 | 5 | 4 | 4 | 7 | 4 | 4 | 6 | 6 | 3 | 5 | 3 | 3 | 3 | 3 | 4 | 4 |

Fluorescence is quantified to identify hits

- Foreground:
- Background:

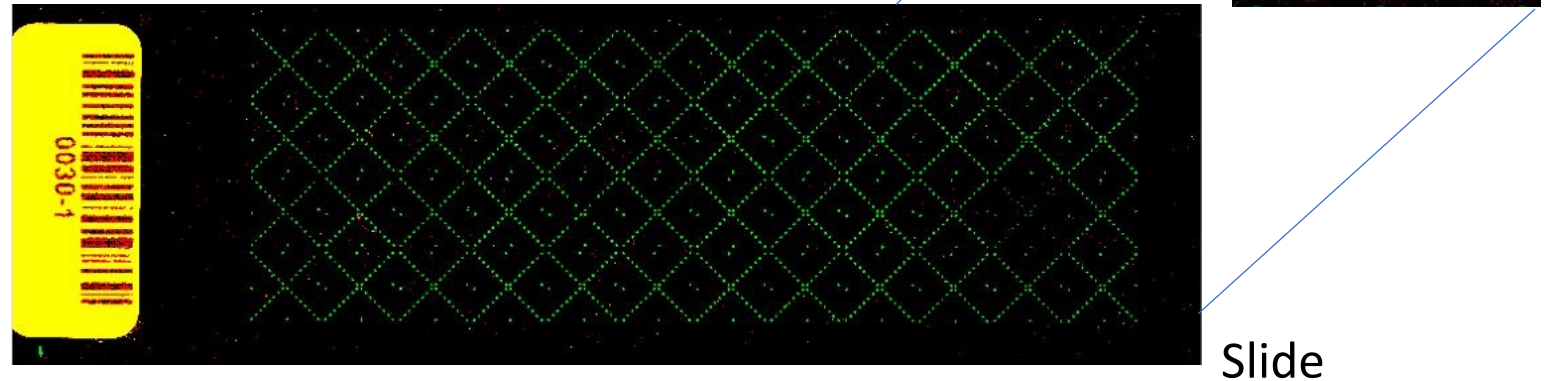


$$\text{Signal-to-noise ratio (SNR)} = \frac{\mu_{\text{foreground}} - \mu_{\text{background}}}{\sigma_{\text{background}}}$$

How do you identify hits from the SMM data?

First, consider bias that exists in the data set

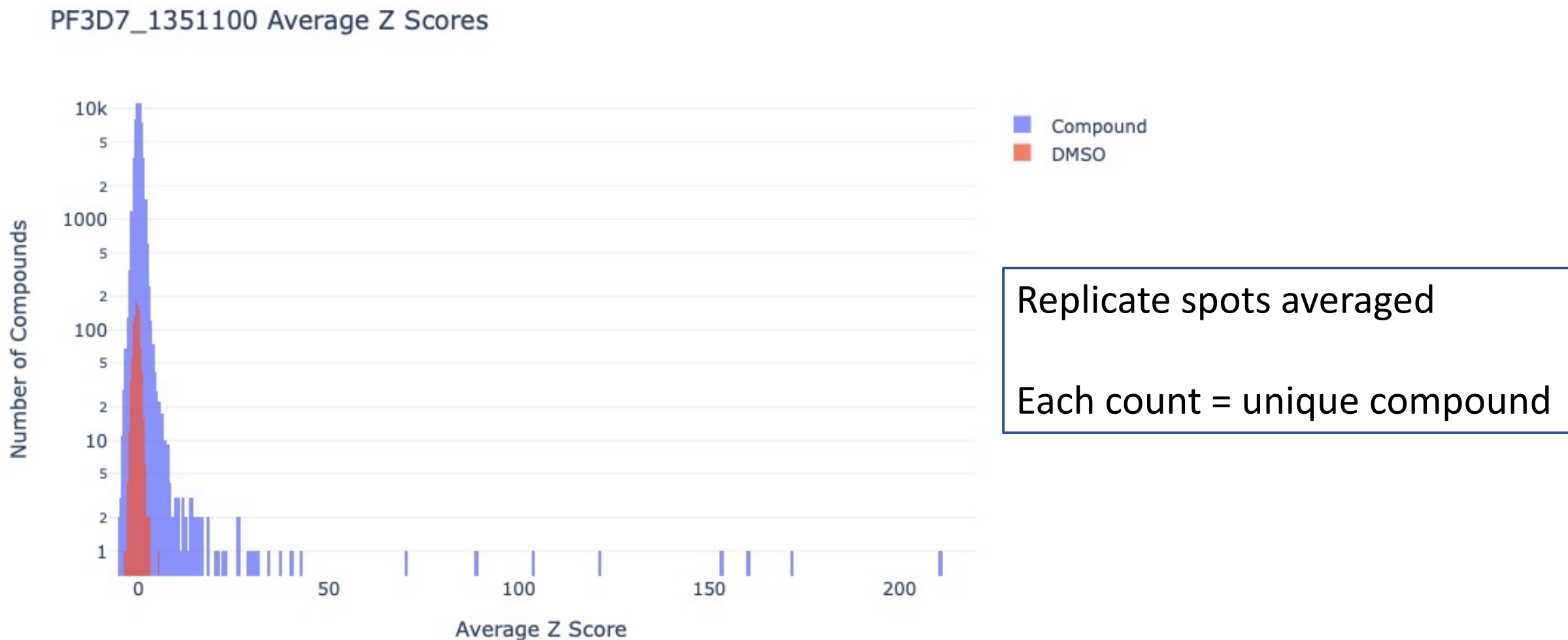
- Across all slides
- Within each block
- Within each slide



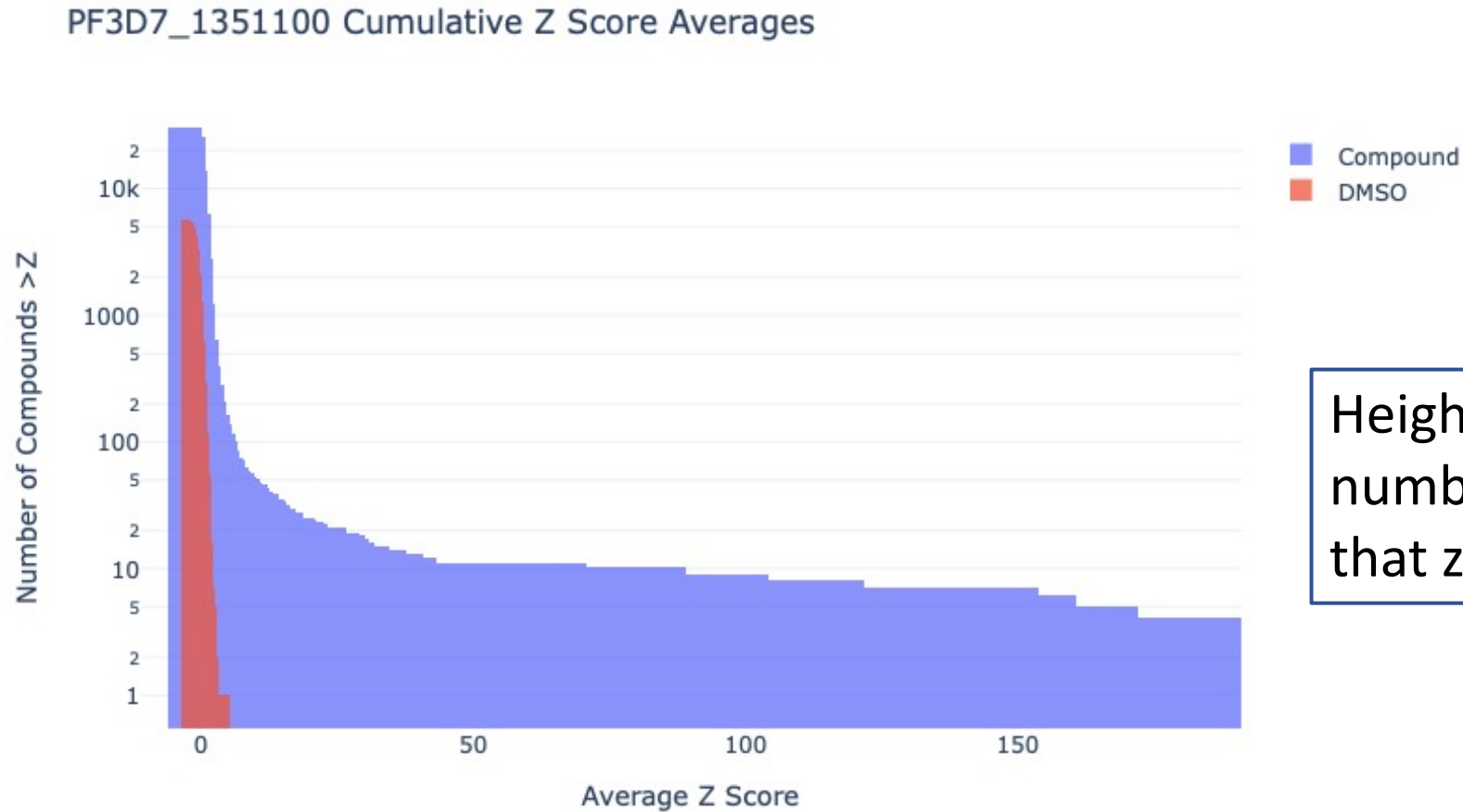
Then, identify hits with significantly higher fluorescence over background

Lastly, manually confirm hits to eliminate false positives

Average Z-score calculated for all compounds



How do you determine a threshold Z-score?



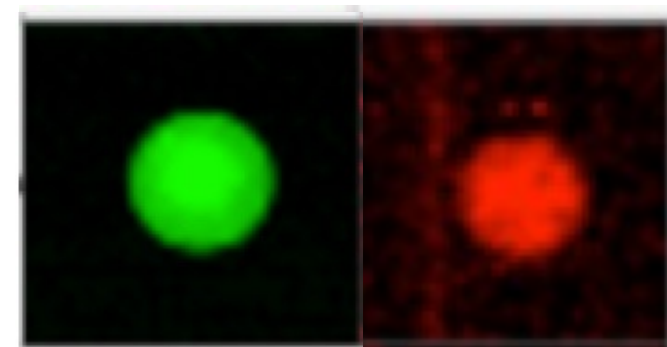
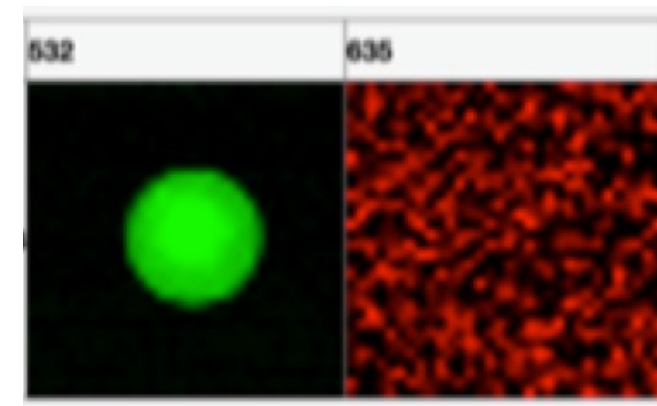
Height at x (average z score) =
number of compounds (y) with
that z-score or higher

How do you validate hits manually?

| ID | Robust Z | SMILES | Validated | |
|-------|--------------|----------|-------------|---------|
| 49592 | 13:KI0001... | 51.03151 | C[C@H](C... | -1 |
| 42089 | 11:KI0001... | 45.09263 | CC1=C(C(... | example |
| 6782 | 02:KI0001... | 39.91118 | CCNC(=O... | -1 |
| 29108 | 08:KI0001... | 39.59436 | C1C(C2=... | -1 |
| 44736 | 12:KI0001... | 33.03555 | C1CN(C2... | -1 |
| 29660 | 08:KI0001... | 31.94118 | CC1=NC2... | -1 |
| 11360 | 03:KI0001... | 26.13059 | C1CN(CC... | -1 |

| | 532 | | 635 | |
|---------|-----|--|-----|--|
| 0011-08 | | | | |
| 0012-08 | | | | |
| 0014-08 | | | | |

Cc1c(C)nn(Cc2c(C)nn2)cc1C(=O)NCCCNc3ccncc3



For Today

- Work through SMM procedure
- Evaluate chemical structures of identified hits
- Discuss close reading of scientific papers with Noreen

For M2D2

- Choose a journal article and sign up on the wiki
 - An article can be presented by only 1 person in a section (first come first served)
- Write and submit a short summary based on wiki guidelines