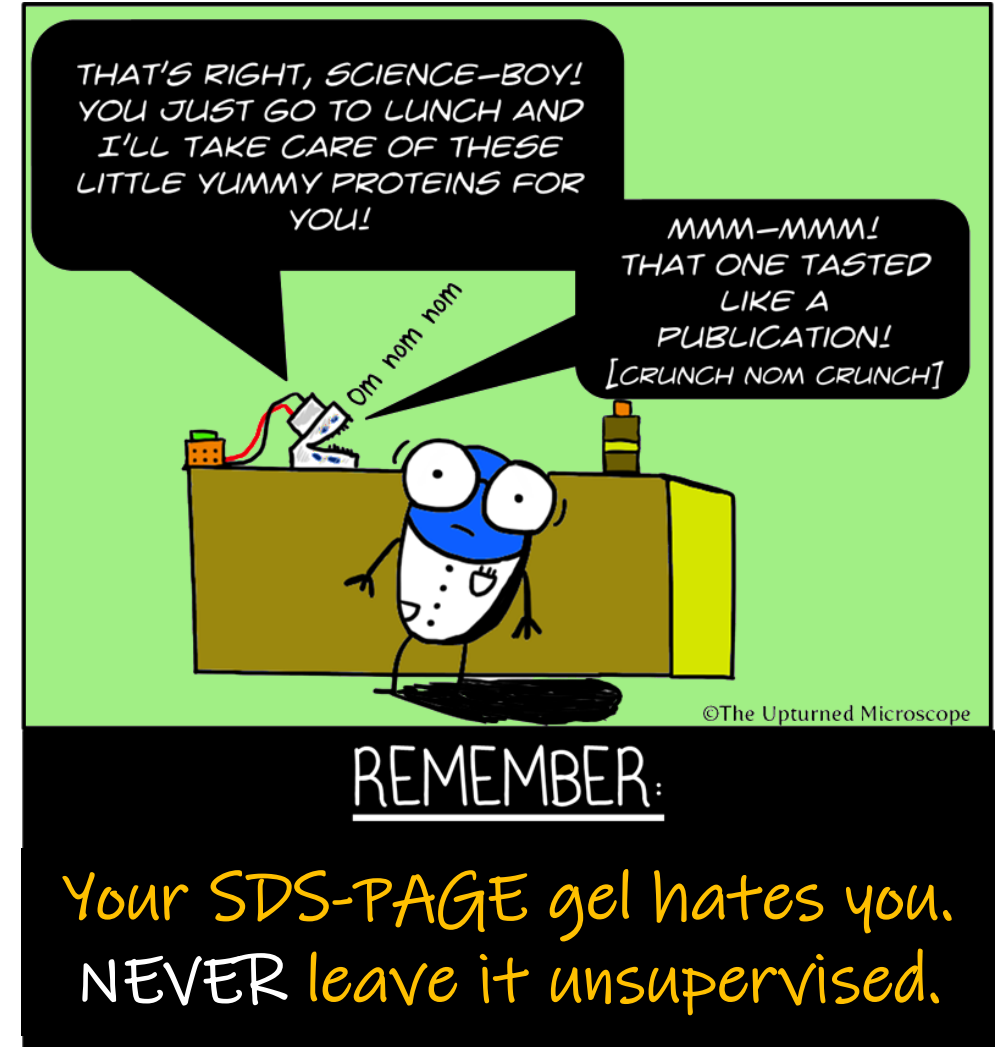


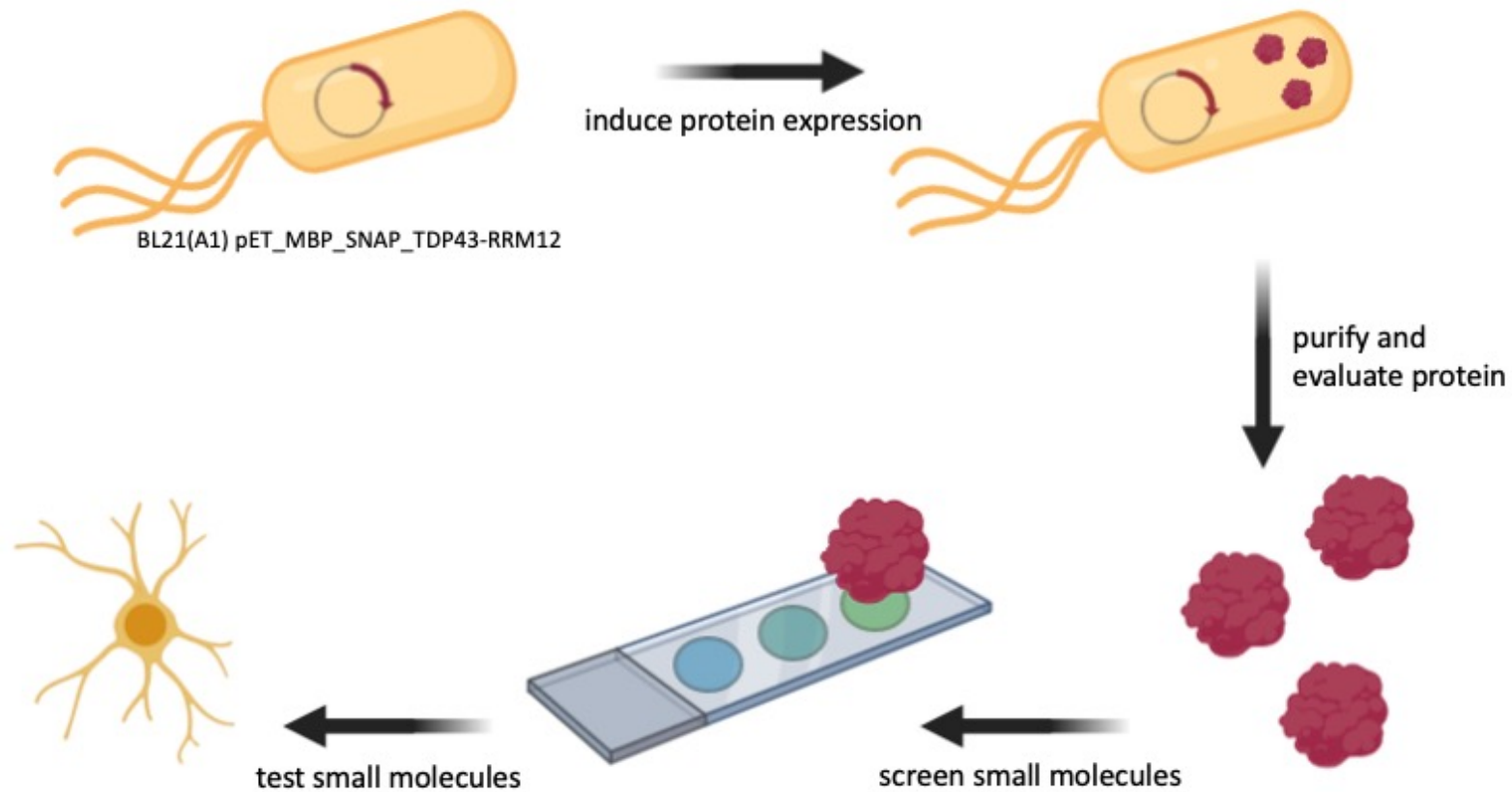
# M2D2: Assess purity and concentration of purified protein

1. Prelab discussion
2. Visualize protein purity using SDS-PAGE
3. Measure protein concentration using BCA assay



# Overview of M2

**Research goal:** Identify and characterize small molecule binders to a protein drug target.



# Protein induction review

① aggregation potential - specific for TDP43

② stressor for bacteria

- What were the two chemicals used to induce TDP43\_RRM12 expression?

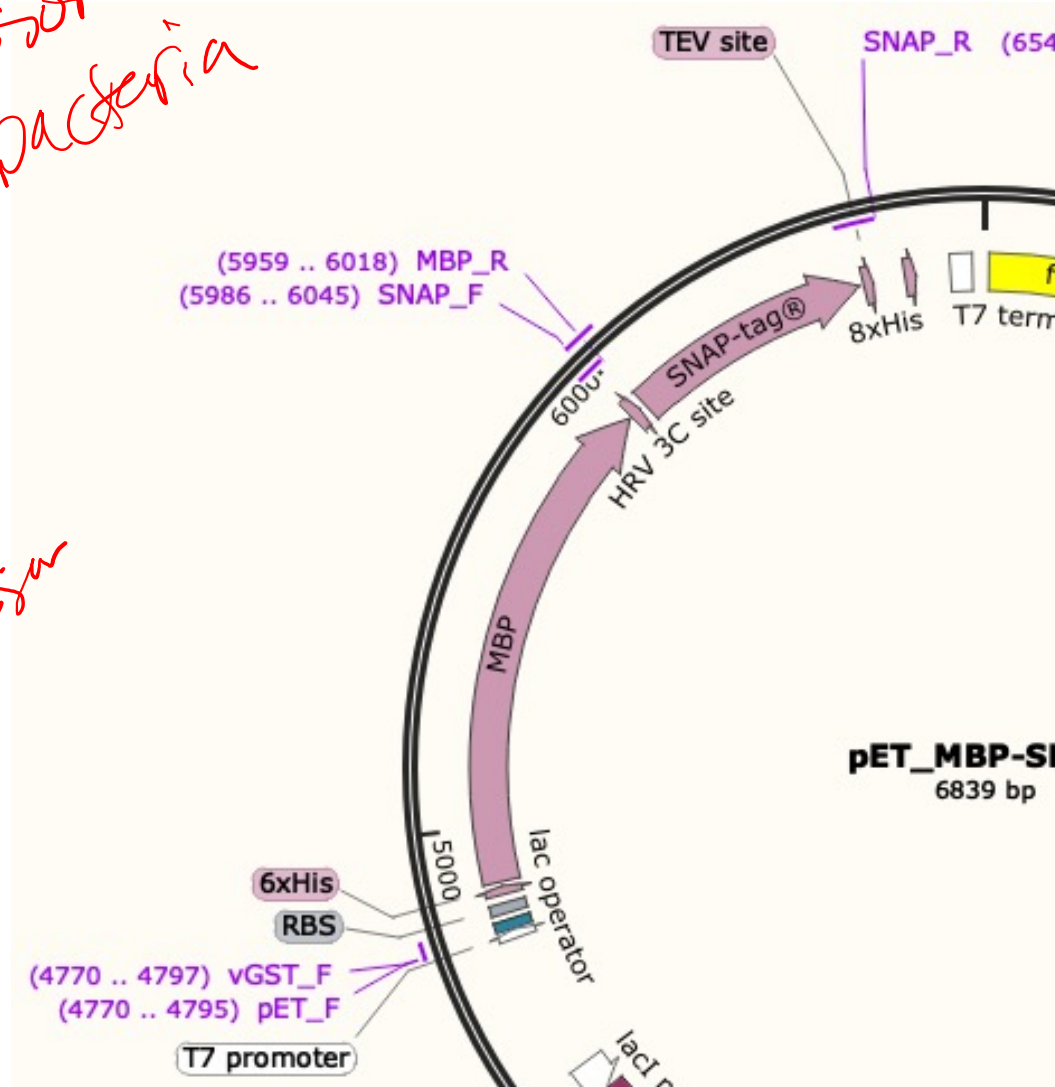
Arabinose

IPTG

- What do they allow to be expressed/how?

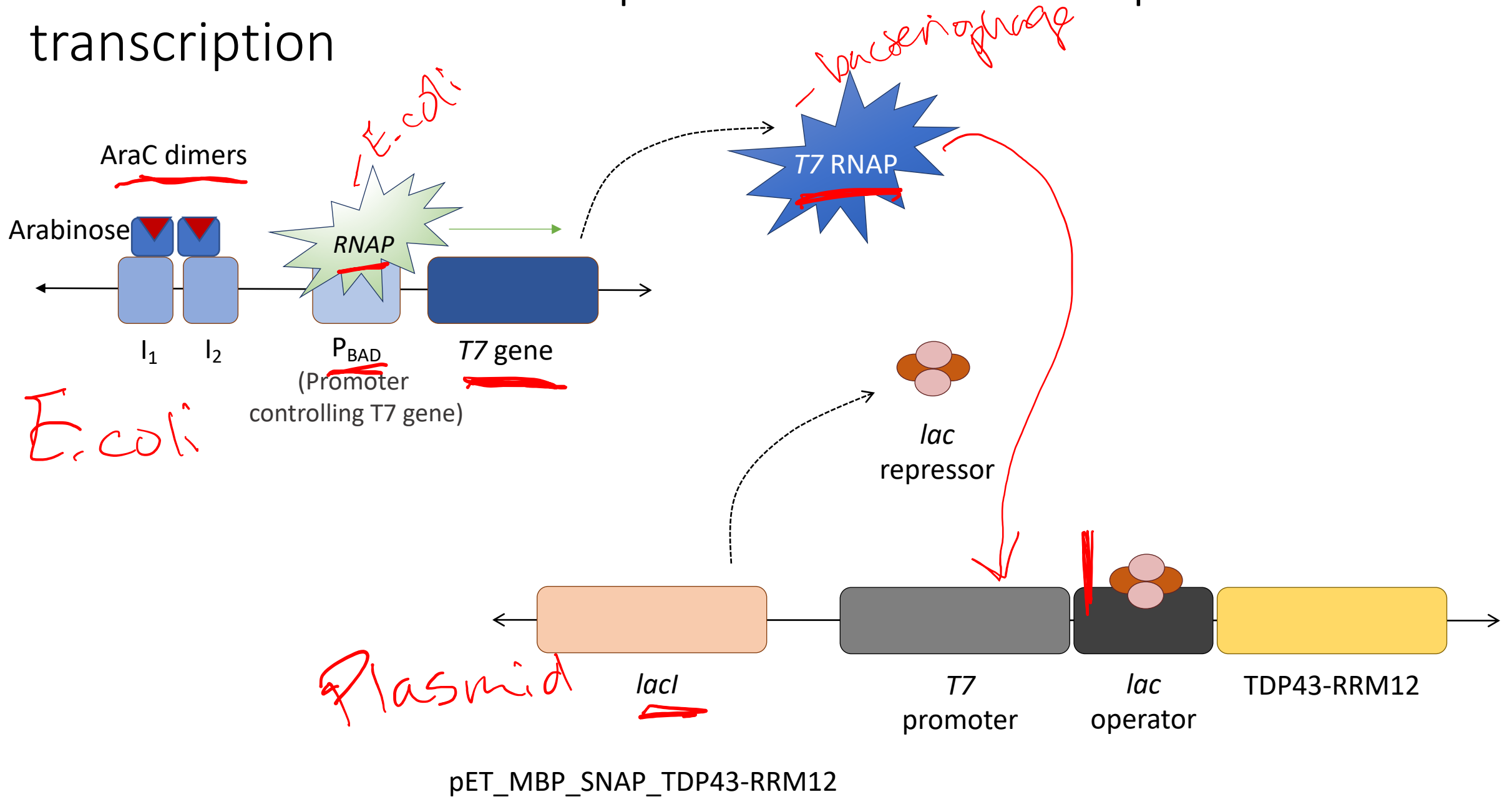
IPTG = lactose analog  
- bind lac repressor

Arabinose = T7 RNAP

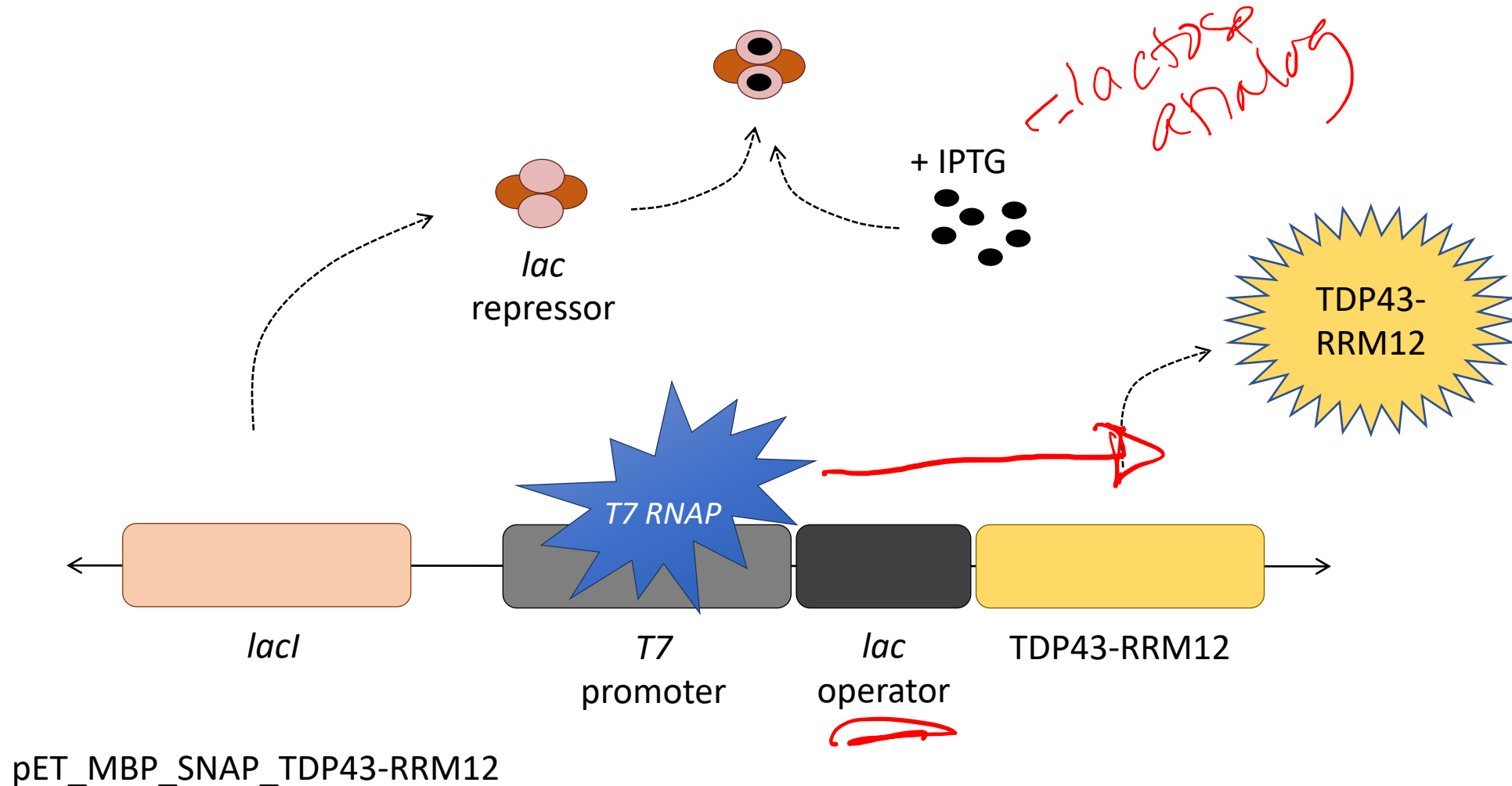


pET\_MBP-SNAP 6839 bp

# Arabinose controls T7 expression while LacI repressor blocks transcription

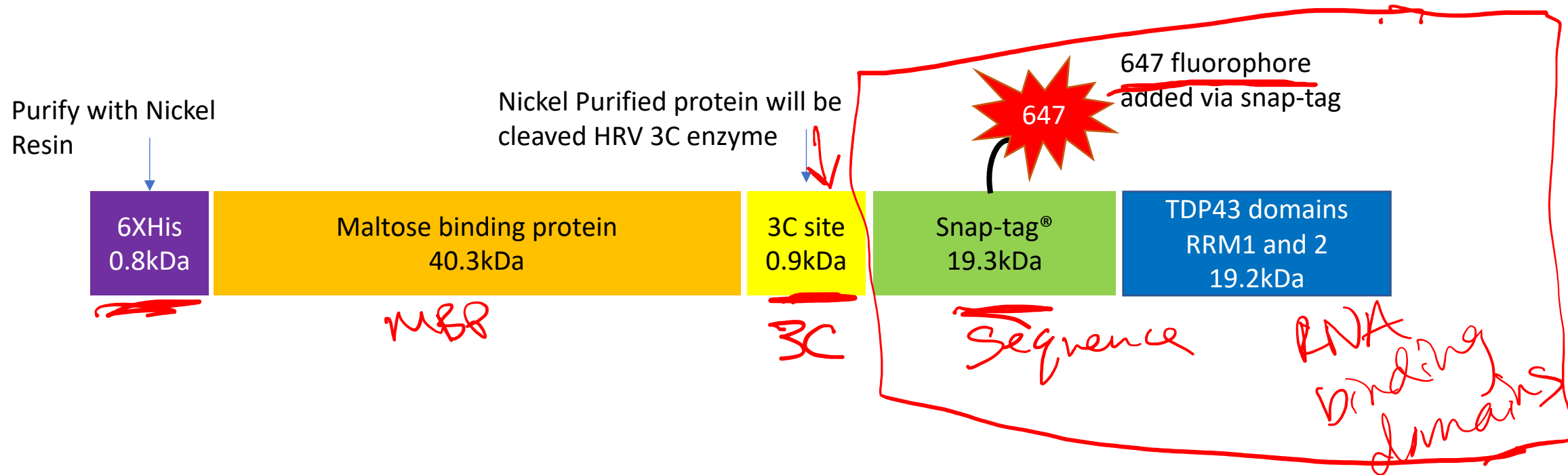


# IPTG 'induces' protein expression by preventing LacI repression



# What is protein expressed in our system

## Our protein for this module:



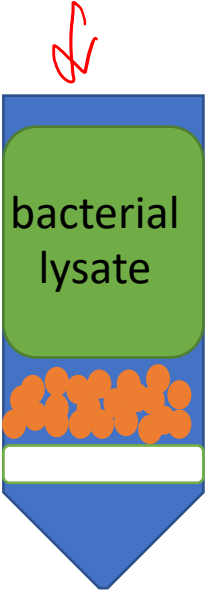
# Protein purification review

• Why this step?

Remove unbound proteins

Remove NS proteins  
Deads / resin / agarose

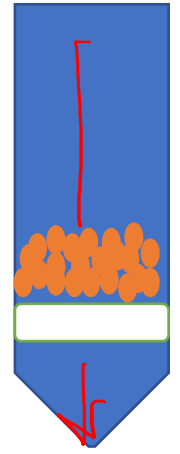
get TDP43  
H2V3C enzyme  
agarose



Ni<sup>2+</sup> resin

**Flowthrough**

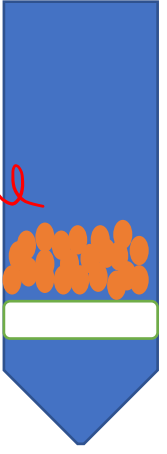
On beads:  
- TDP43  
- His<sup>6</sup> rich proteins



- bacterial proteins, etc...

**Wash**

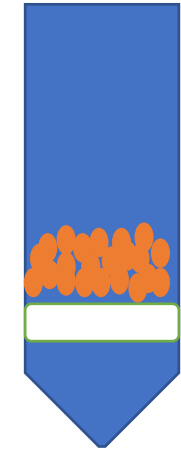
On beads:  
- TDP43  
- imidazole



- imidazole  
- NS proteins

**Elution**

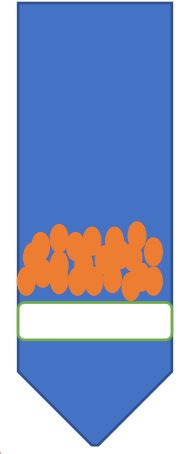
On beads:  
- some imidazole  
- His  
- MBP



- snap  
- TDP43  
- RRM12

- any proteins still bound

**Slurry**



How effective was cleavage?

• What's on the Ni<sup>2+</sup> beads?  
• What's in the expelled liquid?

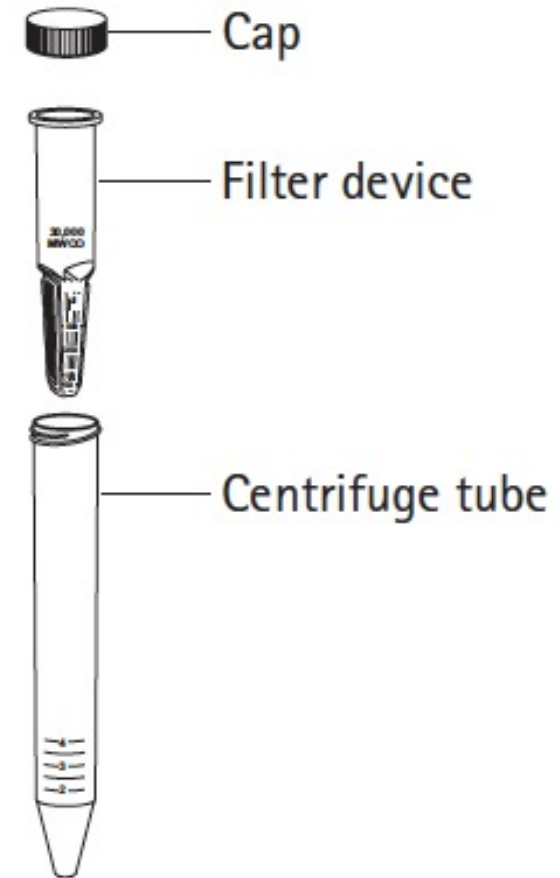
# Protein is concentrated after purification

*1 ml elution*

- Filter device sits within centrifuge tube
  - **Protein added to filter device** before centrifugation
- Filter has MW cutoff of 3 kDa
  - **protein retained in the filter device** during centrifugation

- 
- TDP43-RRM12 + Snap-tag = ? *~ 40 kDa*
  - 6x His tag = 2.5 kDa

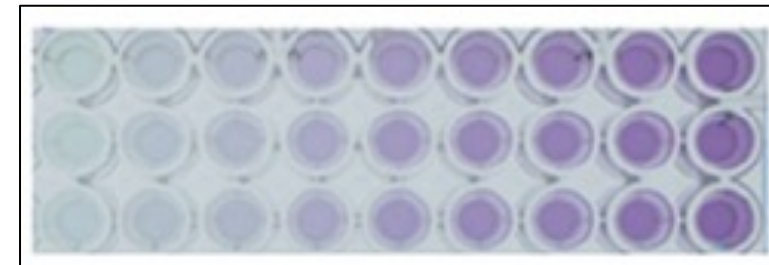
- How does this concentrate the protein?





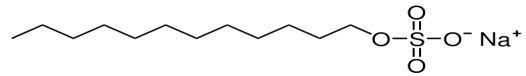
# How will you assess purity and concentration?

- Check purity using SDS-PAGE
  - Identifies presence of protein during purification procedure
  - Visual detection of other proteins in sample
- Measure concentration using BCA assay
  - Colorimetric assay
  - Calculate concentration from standard curve



Bradford  
Lowry

Purity: Sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE)

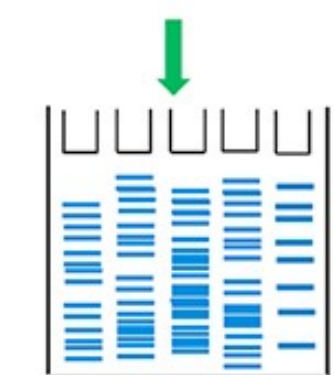
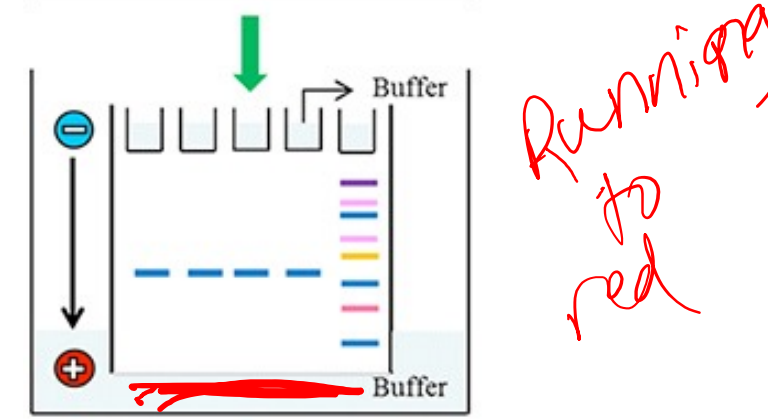
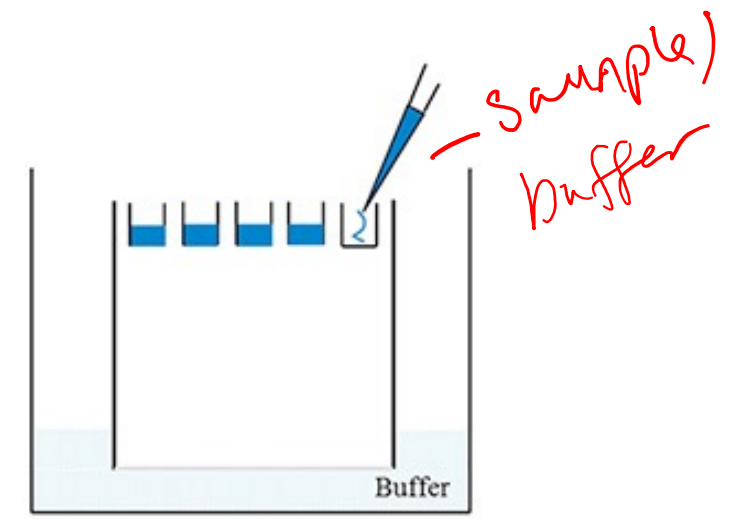


# How are proteins separated?

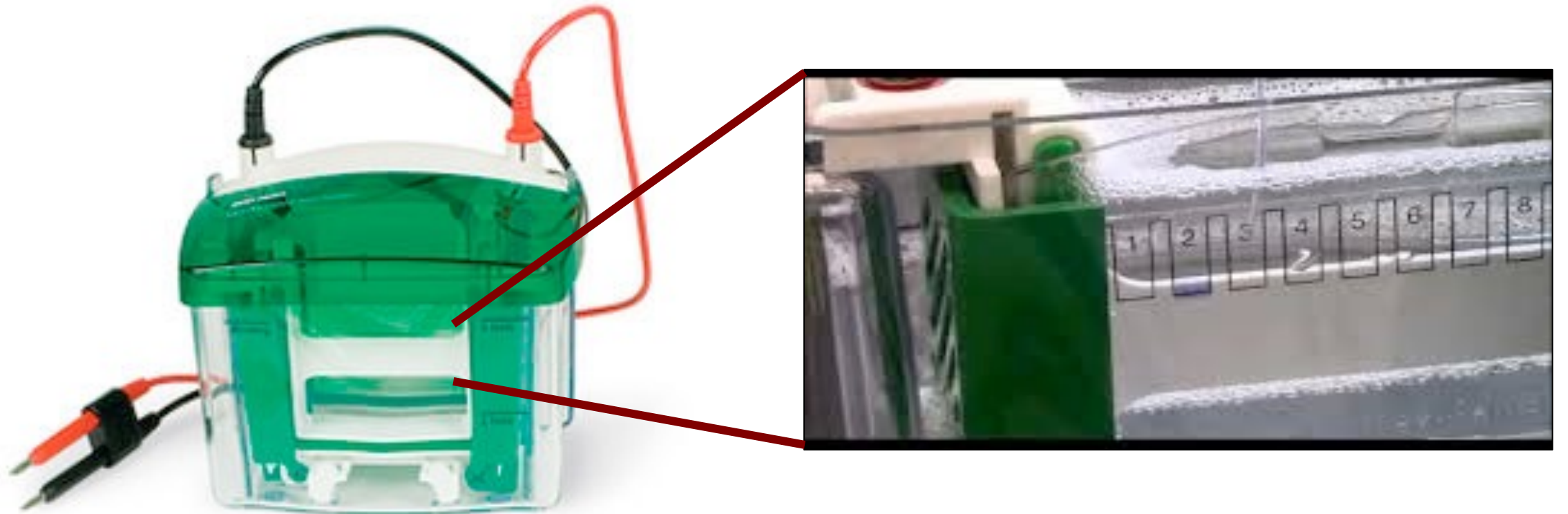
- Laemmli buffer and boiling results in denatured and (-) charged proteins
- SDS-PAGE separates proteins by size
- Electrophoresis completed in TGS buffer
  - Tris-HCl
  - SDS
  - Glycine

*- regulate salts*  
*- regulate pH*

*- Running buffer*



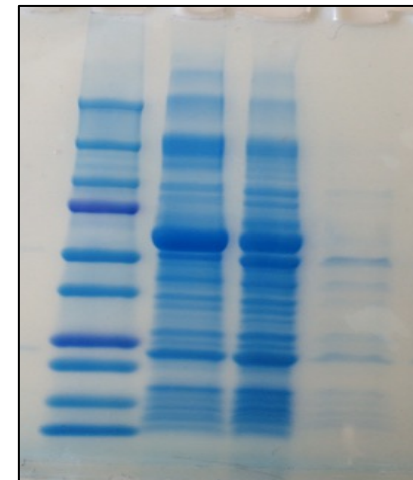
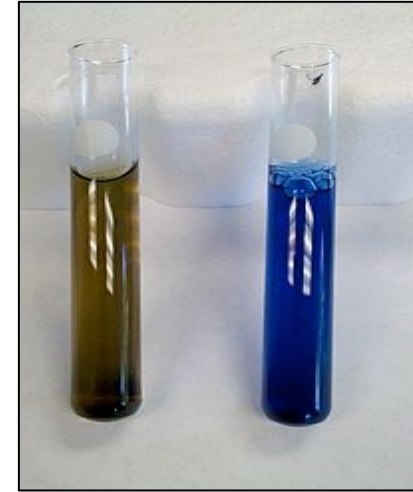
# Demonstration of SDS-PAGE



# How are proteins visualized?

Coomassie brilliant blue G-250 dye used to stain gel after electrophoresis

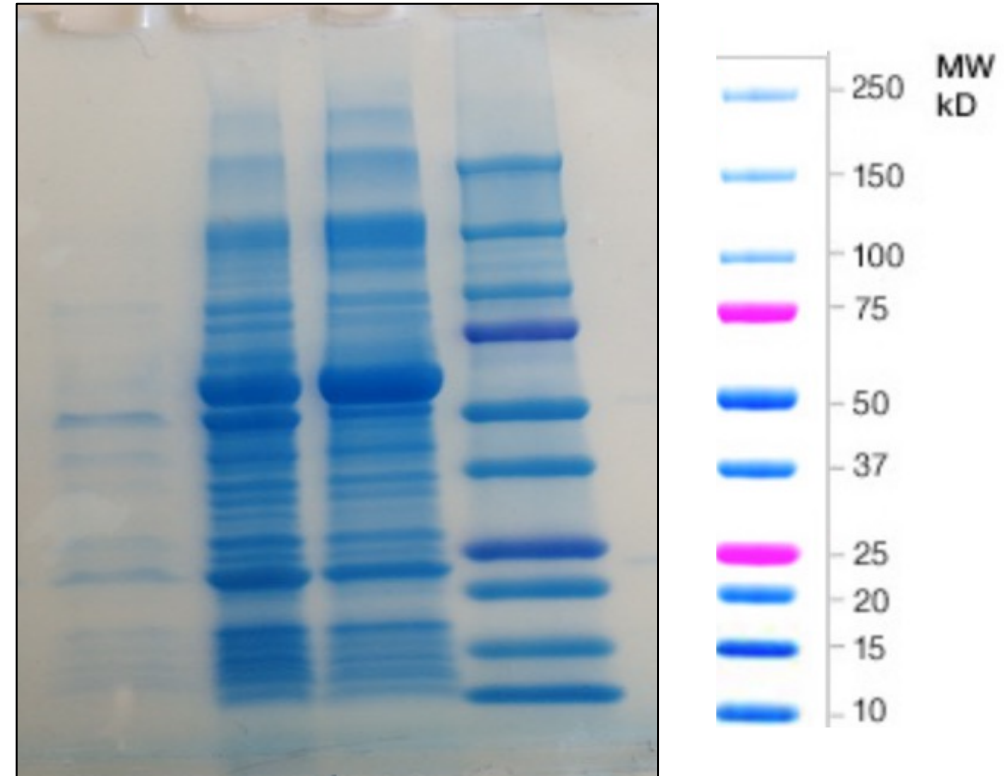
- Red if unbound (cationic form)
- Blue if bound to protein (anionic form)
- Hydrophobic and electrostatic interactions with basic residues
- Arg (also His, Lys, Phe, Trp)



# What are the expected results of SDS-PAGE?

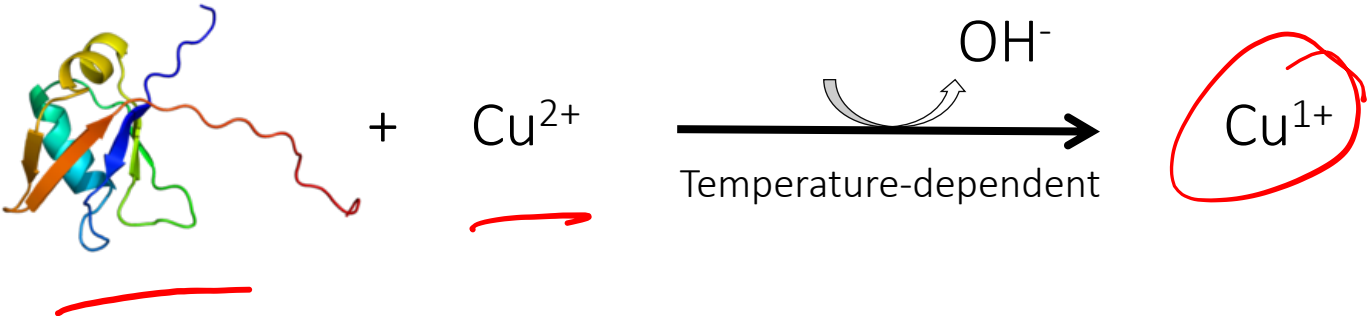
Each lane of the gel should be explained in the results

- What bands are expected? Do you see the bands you expected?
- Do you see any unexpected bands?
- What do the bands tell you about the purity of your protein?
- What does might this tell you about the protein concentration calculated in the next step?



# Concentration: Bicinchoninic acid (BCA) protein assay

Step 1: Biuret reaction; chelation of copper with protein, reduction of copper



*- peptide bonds reduce copper ions proportional to [protein]*

Step 2: BCA complexes with cuprous cation

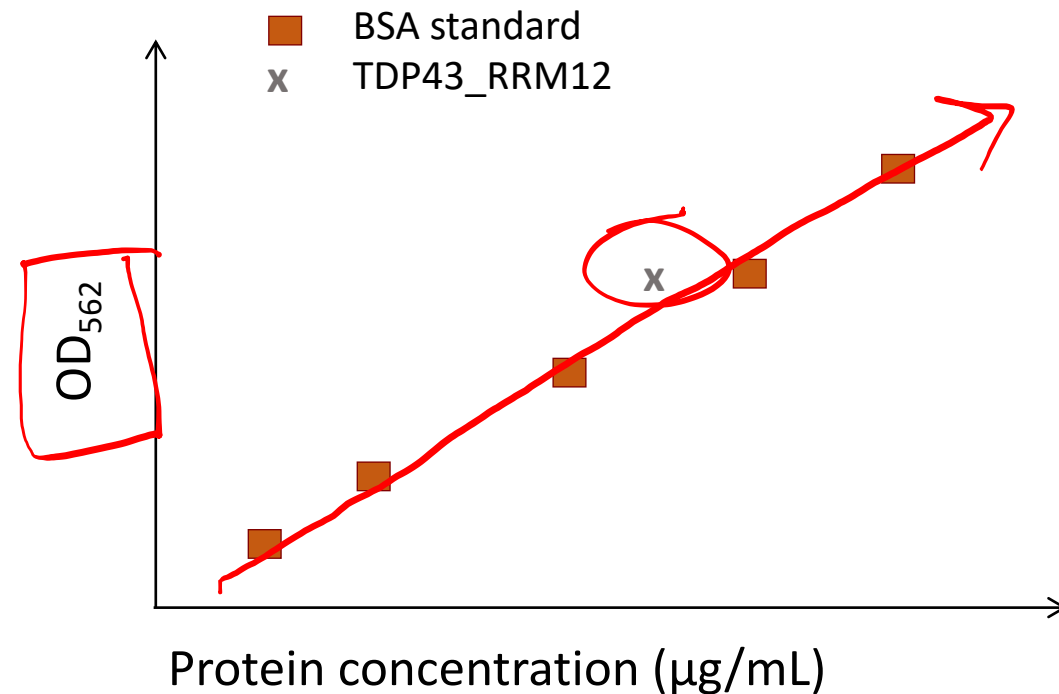


*- BCA chelate Cu+ to complex*

# BCA/Cu<sup>1+</sup> absorbance proportional to protein concentration

Standard curve generated using serial dilutions of bovine serum albumin (BSA)

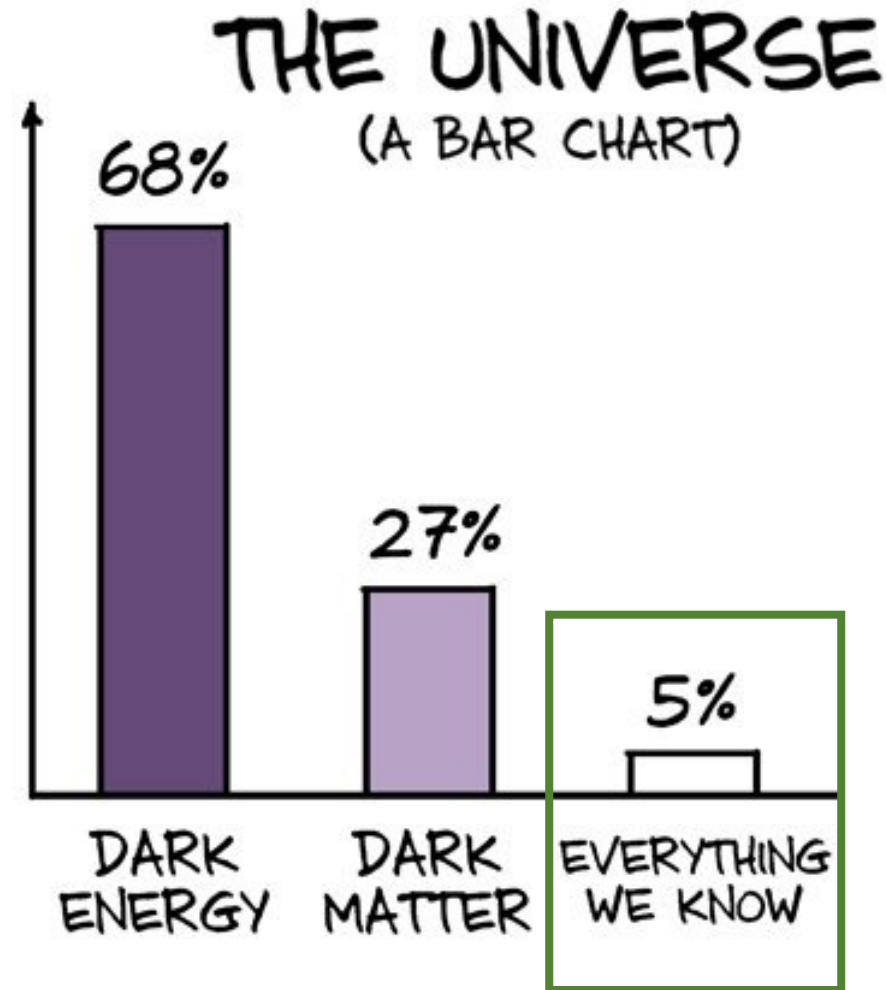
- Equation of the line used to calculate protein concentration
- What does the R<sup>2</sup> value tell you about the standard curve?  
What does this tell you about the calculated concentration?





We know a small fraction of what the universe has to offer (a take-home message)

- Minimal text included to understand the figure
- Oooh, 5%. That's better than expected.
- But, what does that even mean?



## For today...

- Work through M2D2 laboratory exercises with partner
- Work on Mini-presentation!

## For M2D3...

- Outline the Introduction section for Research article
- Review paper for in-class discussion with partner,
  - Draft slide, script for presenting Figure 1 from that paper