

# MID5: DNA Sequencing

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2/24/15

Office Hours this week:

Thursday

Leslie, 1-2pm in 16-429C

Noreen, 2-4 pm in 16-429C

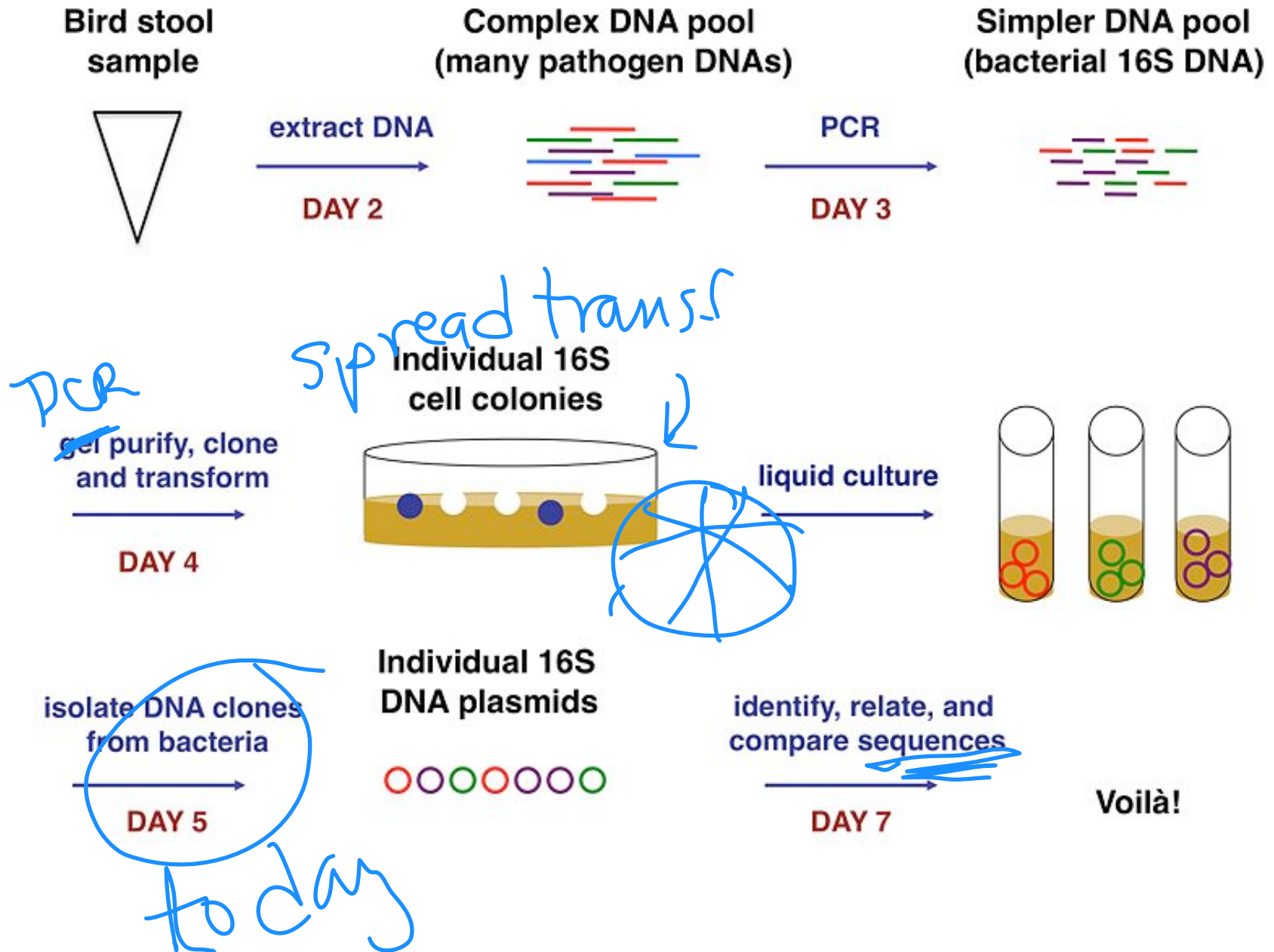
# Announcements

- Lab treat today!



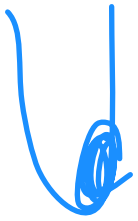
- Journal club next time: Meet in **16-336**
  - Presentation order will be determined by upload order on Stellar
  - MID6 presenters at 1:15pm to setup
  - Presentations start at 1:30pm SHARP
- MID3 Homework — Noreen will return asap.

# Bird Microbial Communities -- Experimental Overview



# Overview: Plasmid Purification -- Miniprep

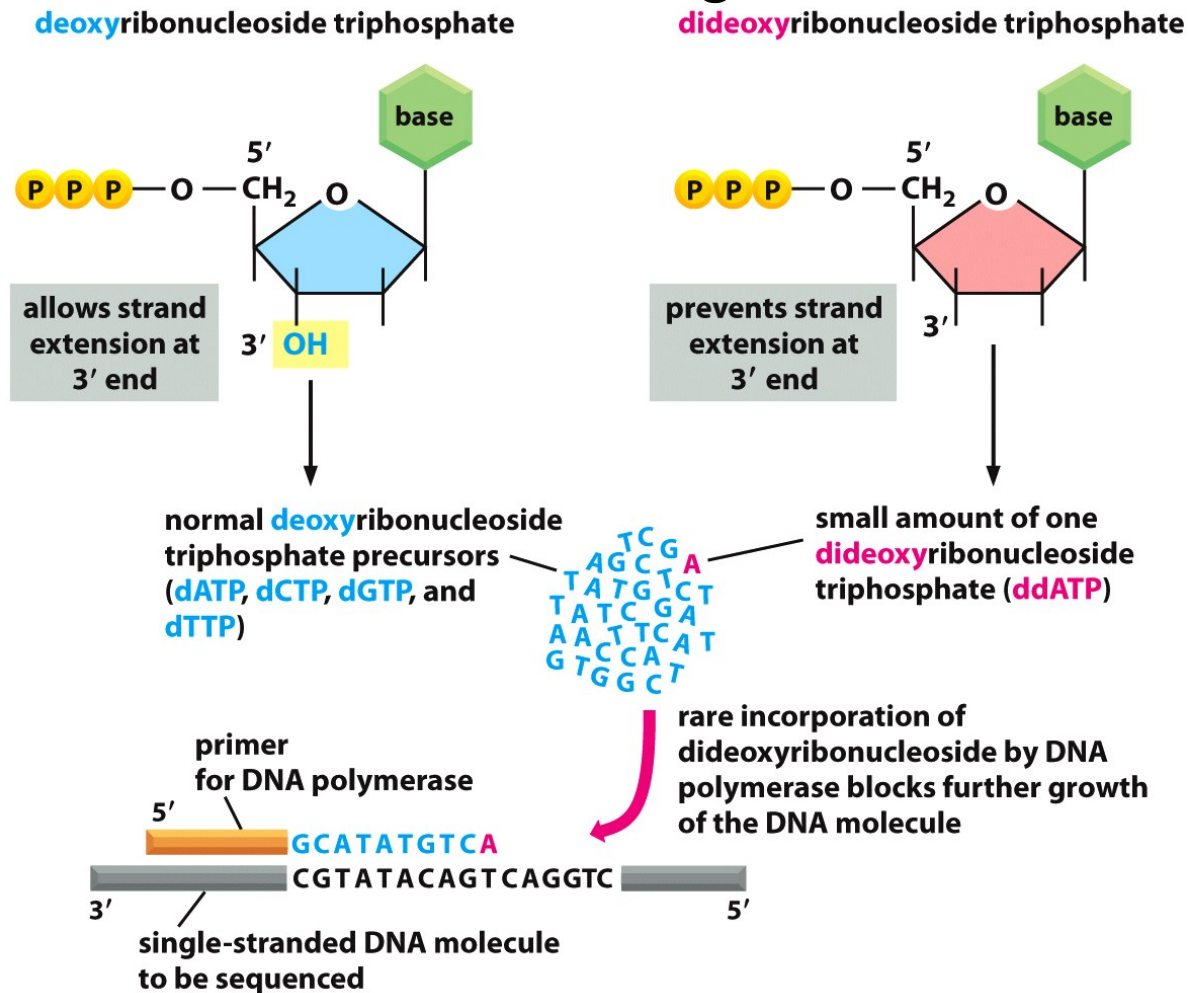
Clean it up!



Step	Contents	Purpose
Prepare	Tris & EDTA Buffer	1) resuspend/buff 2) cations neutral
Lyse	SDS NaOH ↑ pH	precip denatures DNA
Neutralize	Acetic Acid/KAc	↓ pH re fold DNA
Concentrate	Spin all	plasmid super
Wash	EtOH, dry	plasmid DNA ≠ column

# Overview: Sanger Sequencing

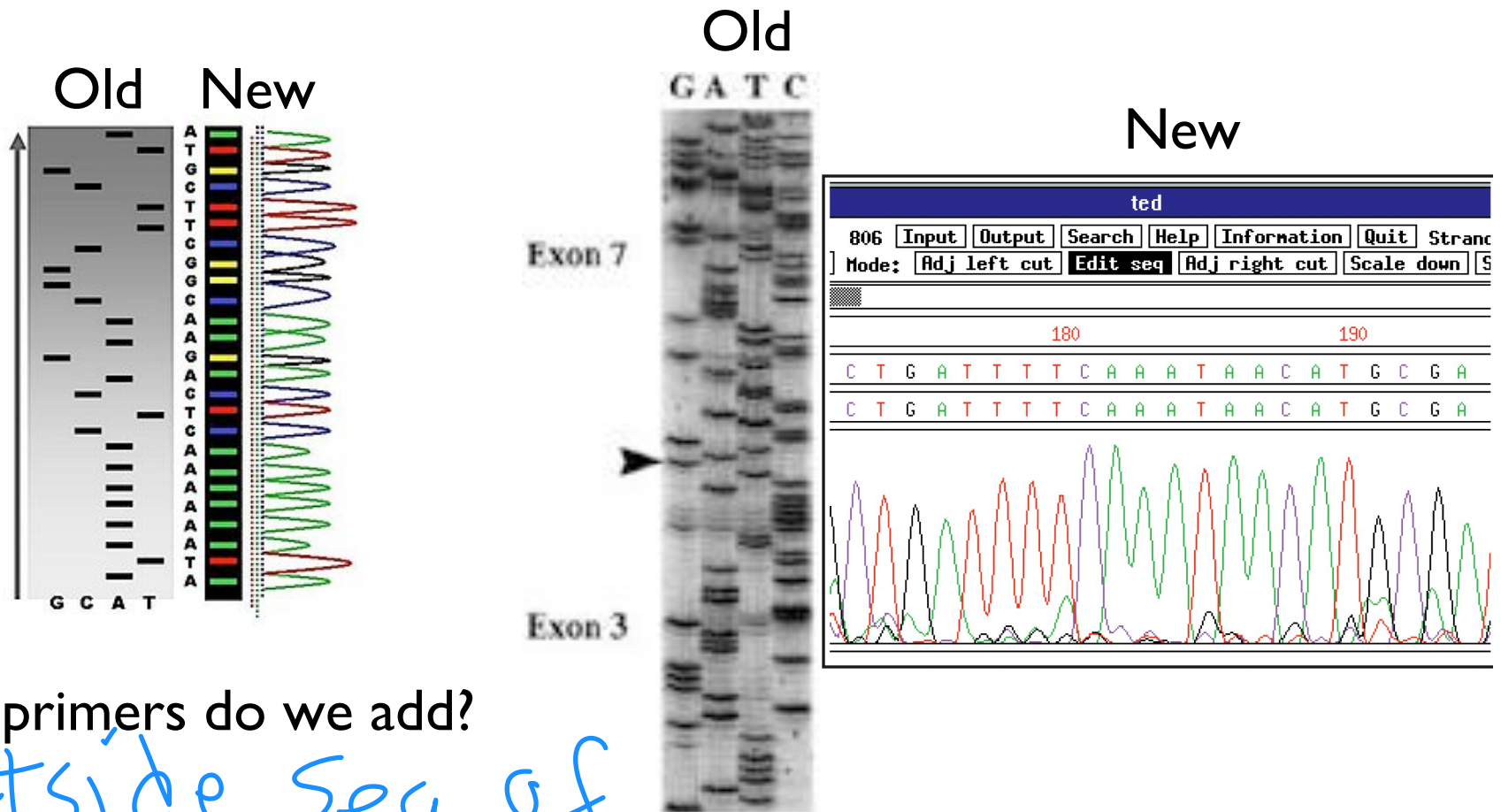
Four dye labeled dideoxynucleotides added to each reaction  
'Chain terminating reaction'



<https://www.youtube.com/watch?v=nudG0r9zL2M>

# Overview: Sanger Sequencing

Four dye labeled dideoxynucleotides added to each reaction

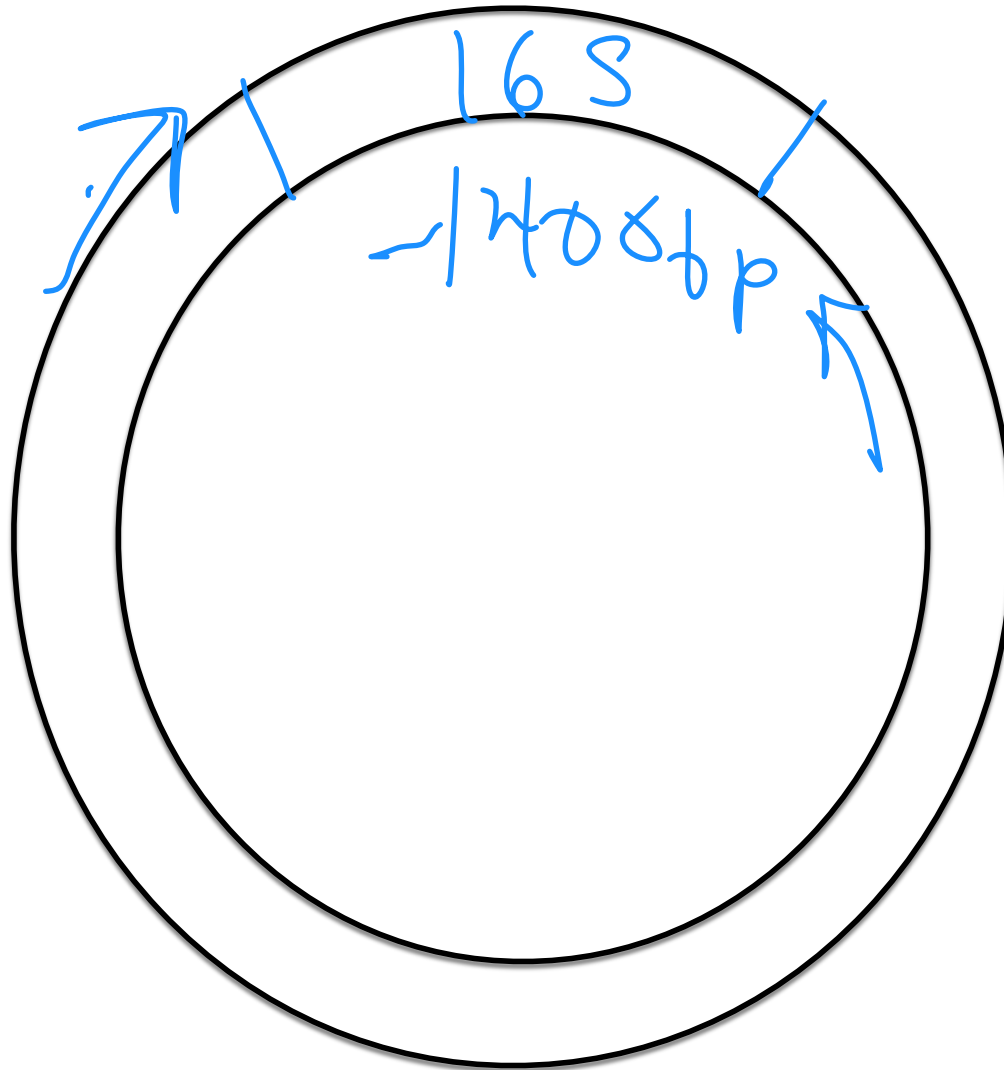


What primers do we add?

outside seq of interest

good rxn 1000 bp

# Overview: 16S rRNA sequencing



## Today in lab:

- Extract DNA from 8 (!) clones **\*\*\*LABEL TUBES\*\*\***
  - may choose to do this in shifts
- Measure DNA concentration
  - 260 nm all nucleic acids for concentration
  - 280 nm, proteins for purity
- Set up duplicate sequencing reactions for each clone (why?)
- Set up qPCR reactions using your AIV sequencing primers.
- Count colonies!
- Have a most wonderful Wednesday evening! It's above 0°F!!