

# MID6: Lipofection & Stats Practice

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9/30/14

*\* Next time lab Nbt.s are due :*

1. Pre-lab discussion
2. 1/2 lab in TC to set up HR experiment
3. 1/2 in lab to work on statistics practice — today is also a good notebook ‘catch up’ day. (or nap)
4. Switch!

**Thank you Isaak!!!**

# Review MID3 FNT: Figures & Data Interpretation

When you are writing your R&D, consider the following:

★ Title

a. What was the overall goal of these data/figure?

★ Intro statement

- prepare DNA for cloning/ligation

b. What was your expected result?

- bands @ 4237bp + 663bp

c. What was the result?

- bands @ ~4000bp + ~650bp

d. What evidence do you have that your result is correct or incorrect?

- single digest controls for RE function  
"suggest" our experiment is a success

e. In sum, what does this data suggest or indicate? What does this motivate you to do next?

→ ready for ligation

# Review MID3 FNT: Figures & Data Interpretation

- ★ Each R+D section needs a descriptive title
- ↪ Limit each figure (experiment) 1-2 pages max
- ★ Dont ~~to~~ include Methods

# Revisit Methods section: What experiments fit together?

*Due Monday 10/6 @ 5pm*

PCR  
PCR product purification

Transformation

XbaI/EcoRI Digest

Diagnostic Digest GE

XbaI/EcoRI Digest  
Purification (GE)

O/N e.coli cultures

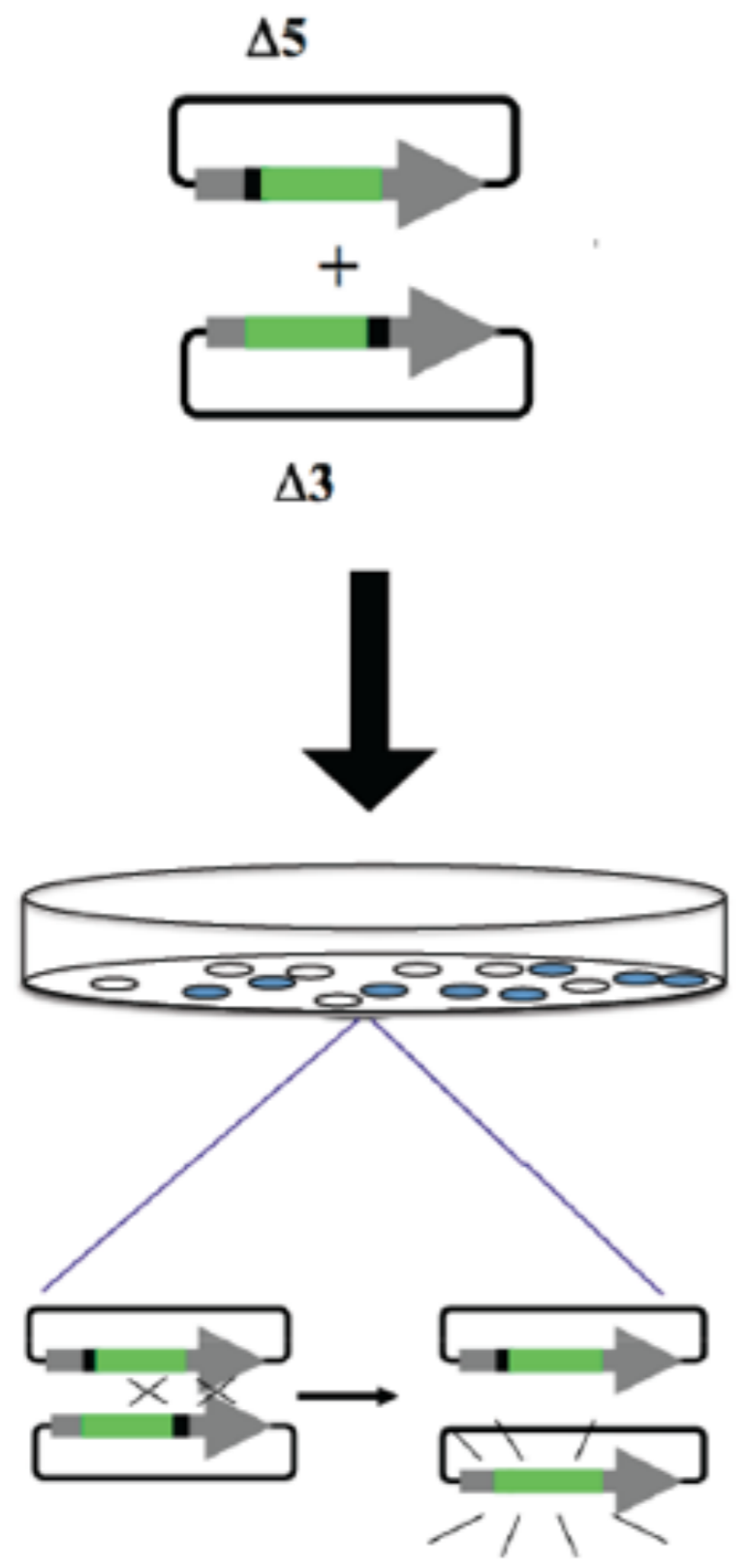
Plasmid purification

Ligation/Precipitation

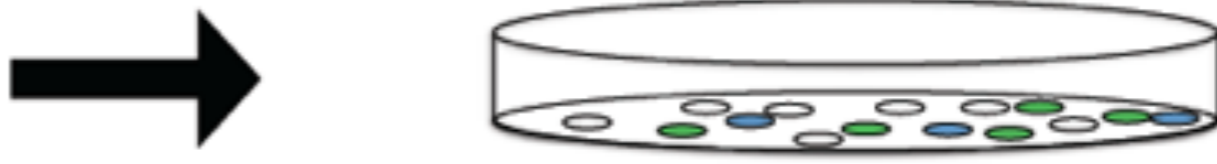
Diagnostic Digest



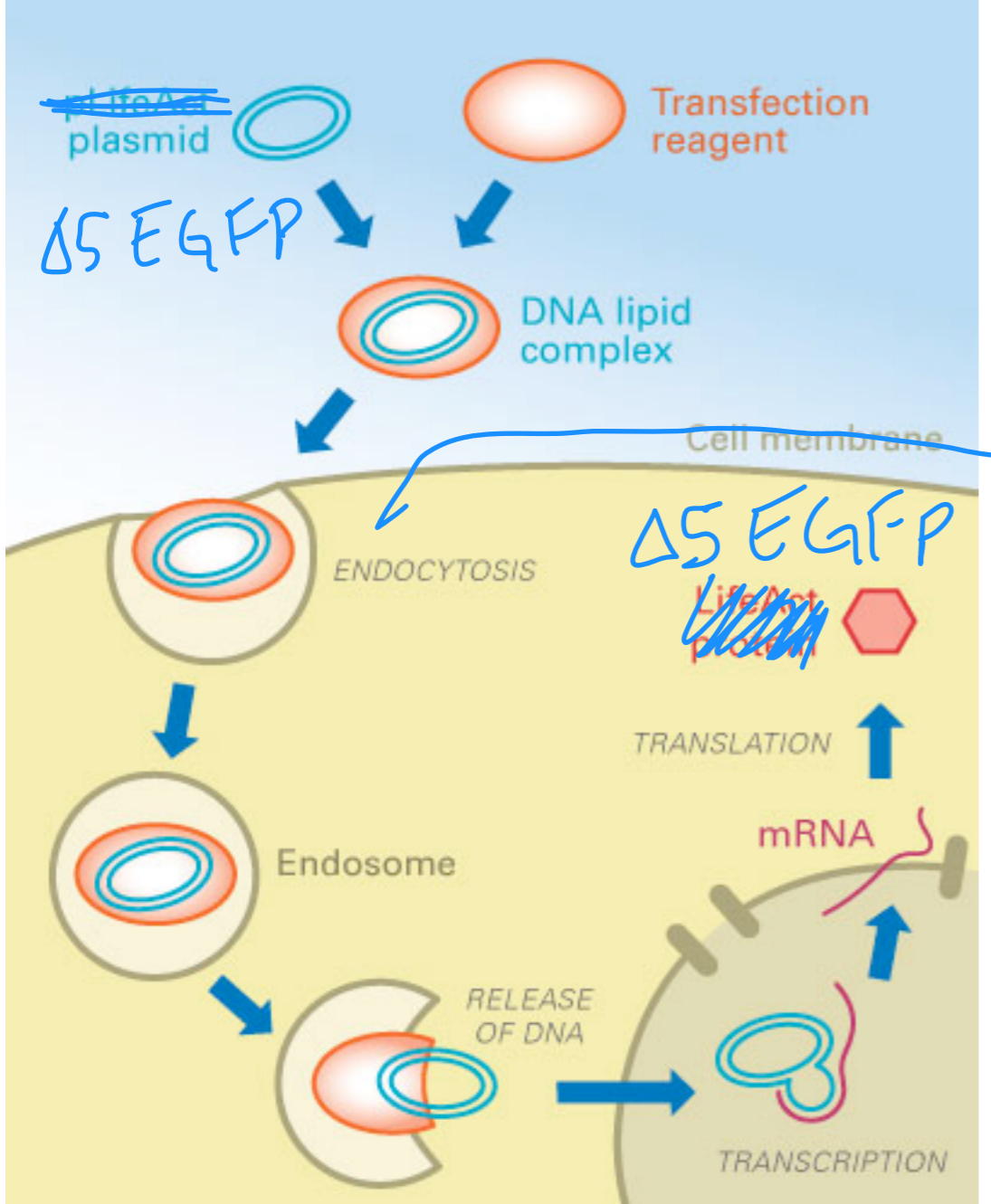
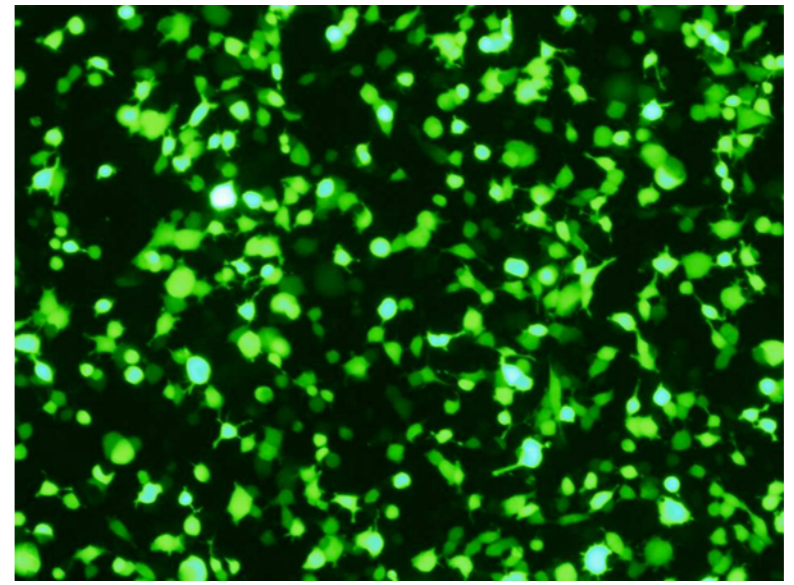
# Step 2: Test the system!



Julian Voss-Andreae  
Steel Jellyfish (Green Fluorescent Protein), 2006  
Stainless steel, 4' x 3' x 3' (1.20 x 0.90 x 0.90 m)  
Location: Friday Harbor Laboratories (San Juan Island, WA)



# Step 2: Test the system!



$\Delta 5$  EGFP

~~LifeAct~~ protein

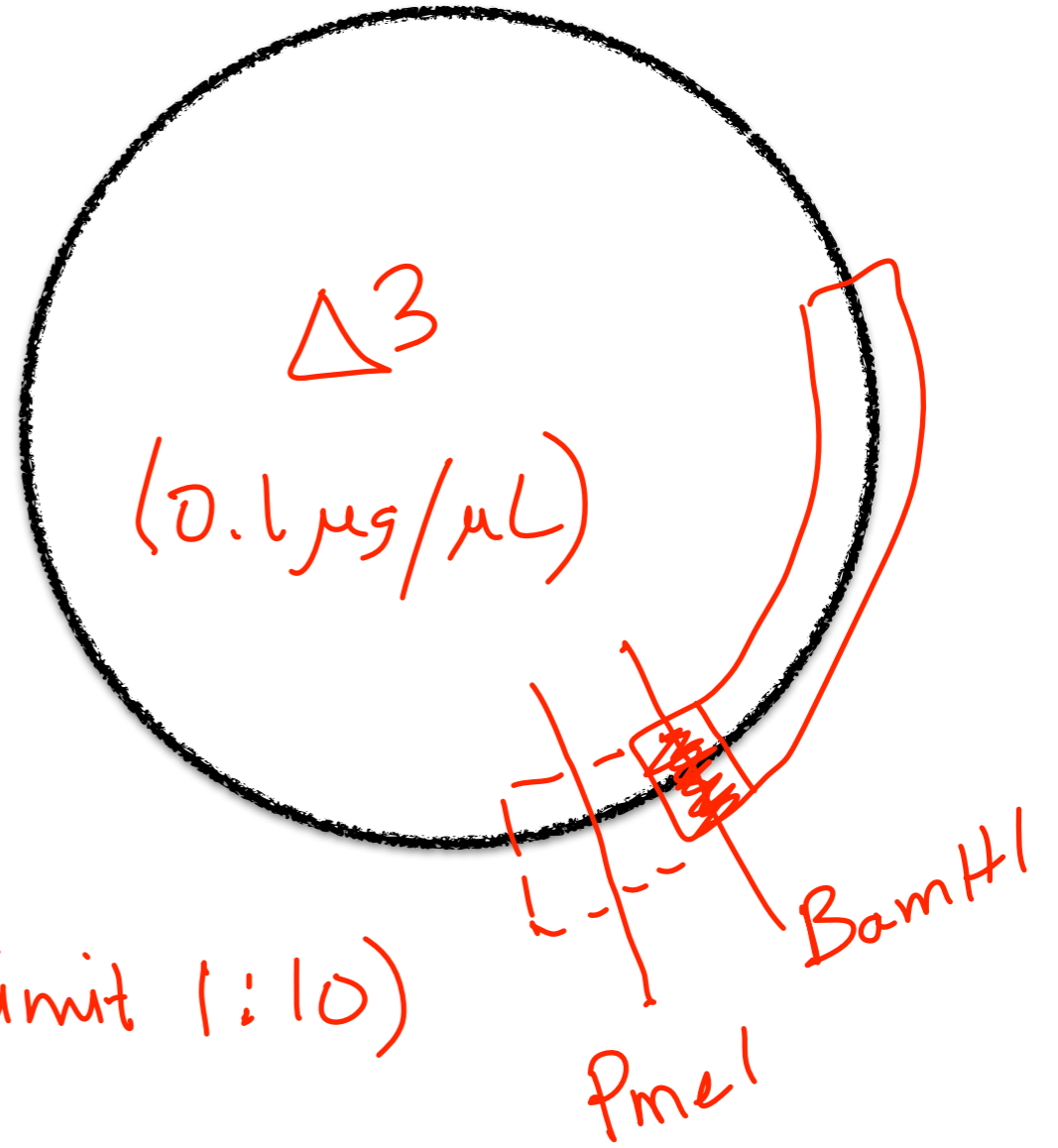
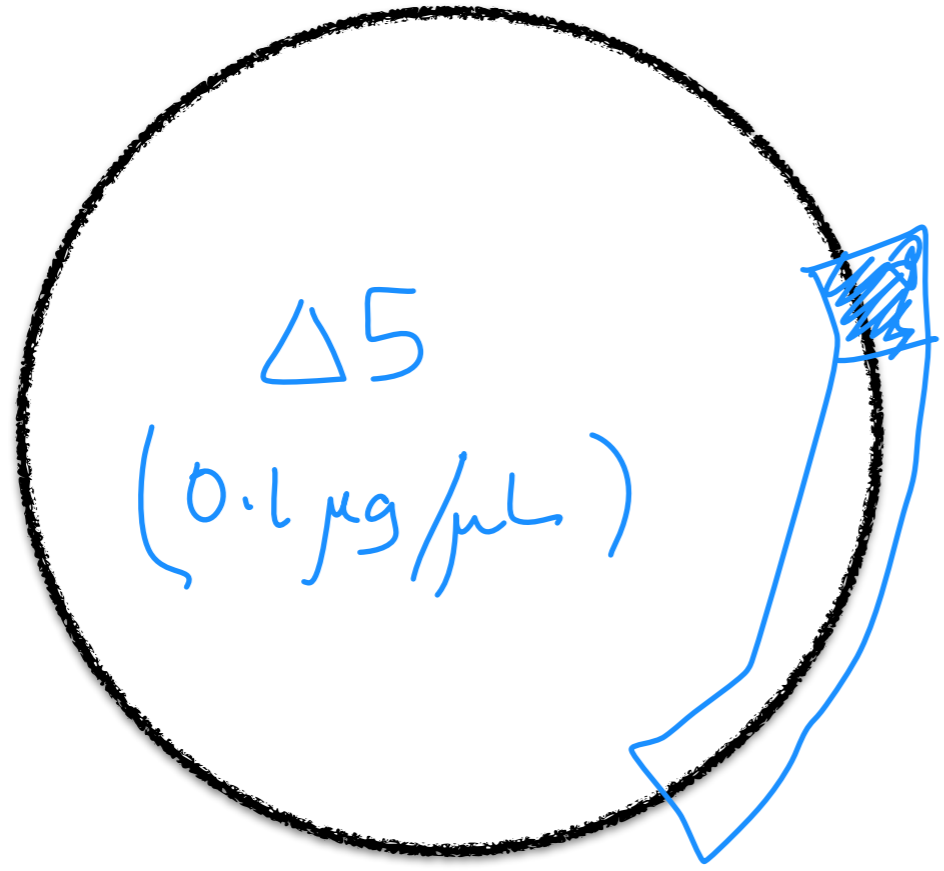
TRANSLATION

mRNA

RELEASE OF DNA

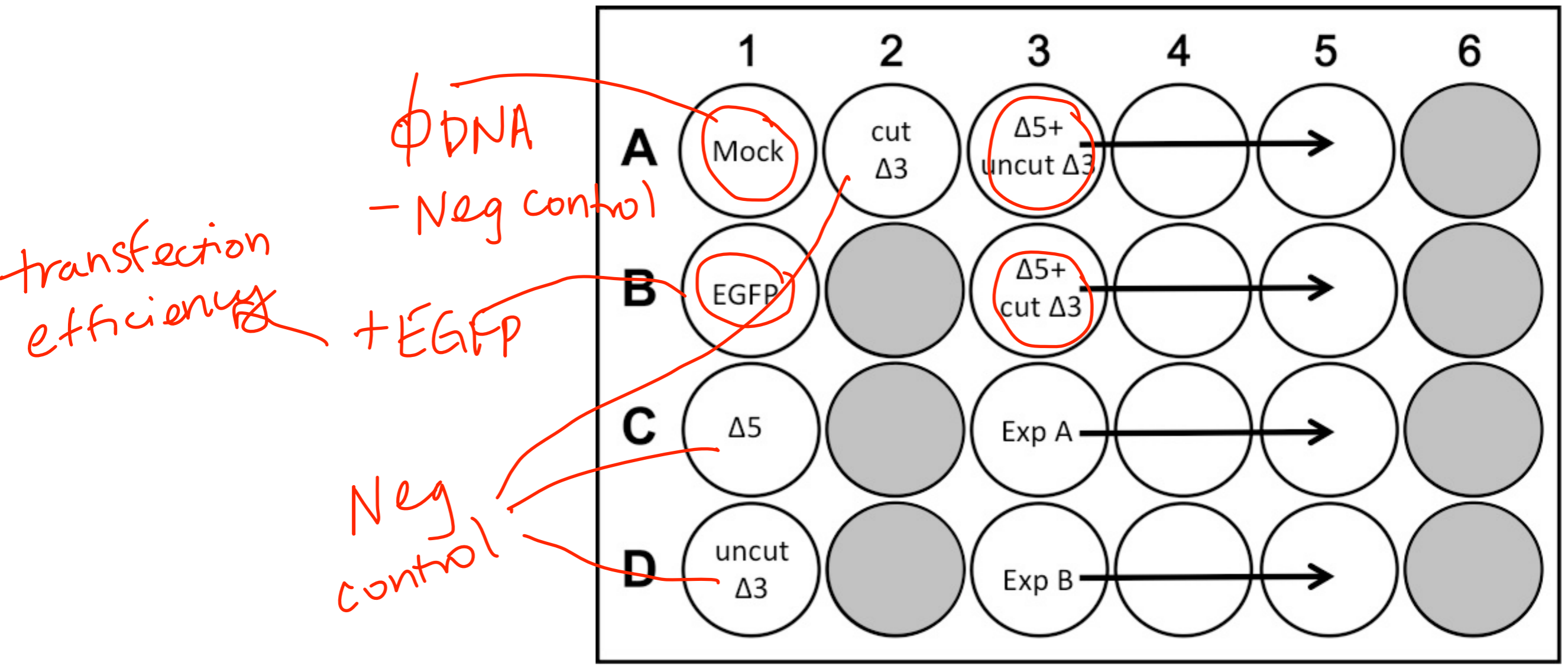
TRANSCRIPTION

# Today you have some options:



- 1) Ratio of  $\Delta 5 : \boxed{\Delta 3} \Rightarrow 1 : 1$  (limit 1 : 10)
- 2) Cut  $\Delta 3$  vs. intact  $\Delta 3 \rightarrow$  endogenous GFP levels (some HR)  
 $\hookrightarrow \uparrow$  HR
- 3) cut  $\Delta 3$  - PmeI vs. cut  $\Delta 3$  - BamHI

# Today you have some options:



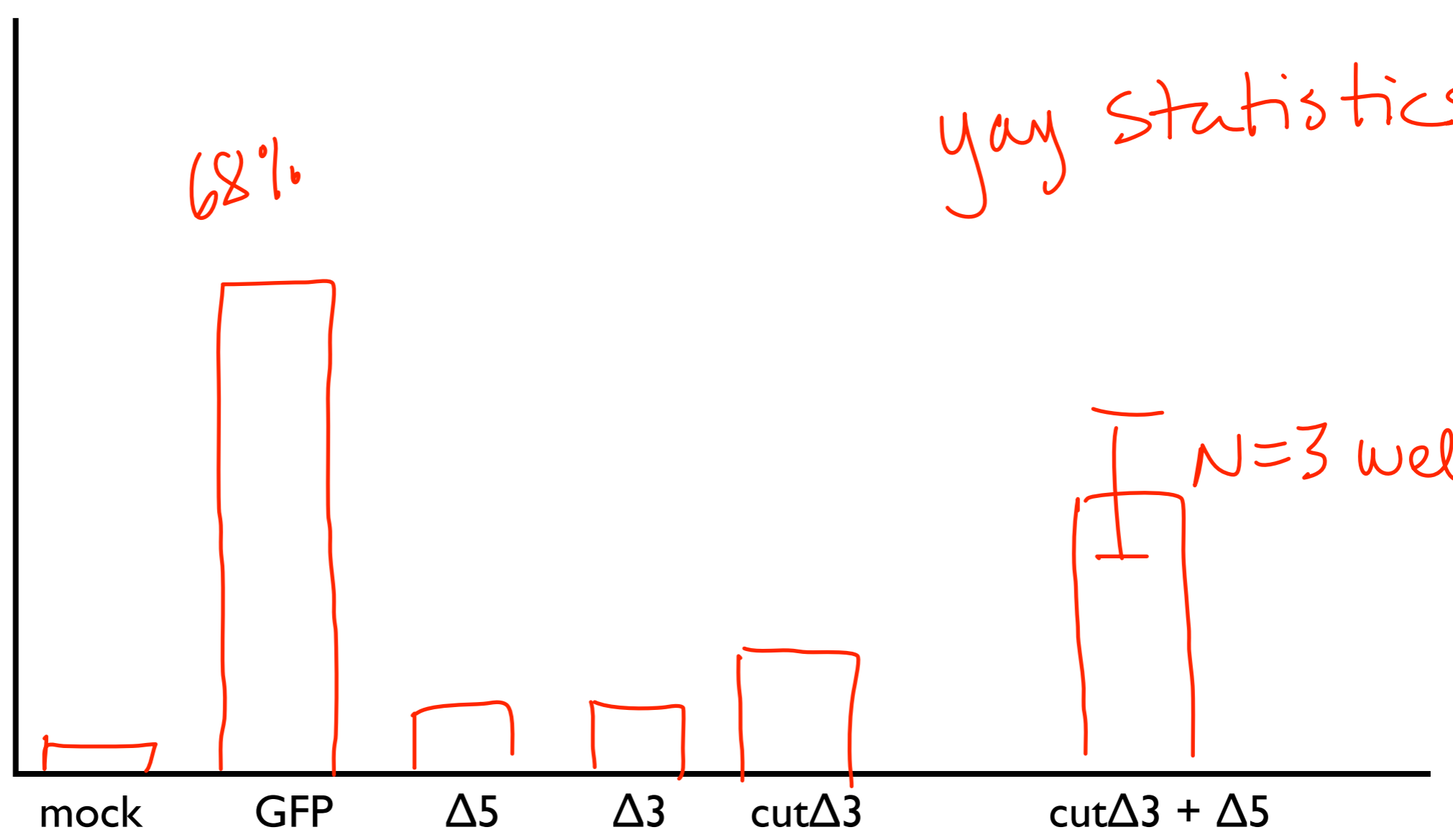
You get to pick

- 1) What is your cut Δ3
  - PmeI
  - BamHI
- 2) Ratio of Δ5 : cut Δ3 (or uncut Δ3)



(how many cells underwent HR)

% EGFP Expression



yay statistics!!

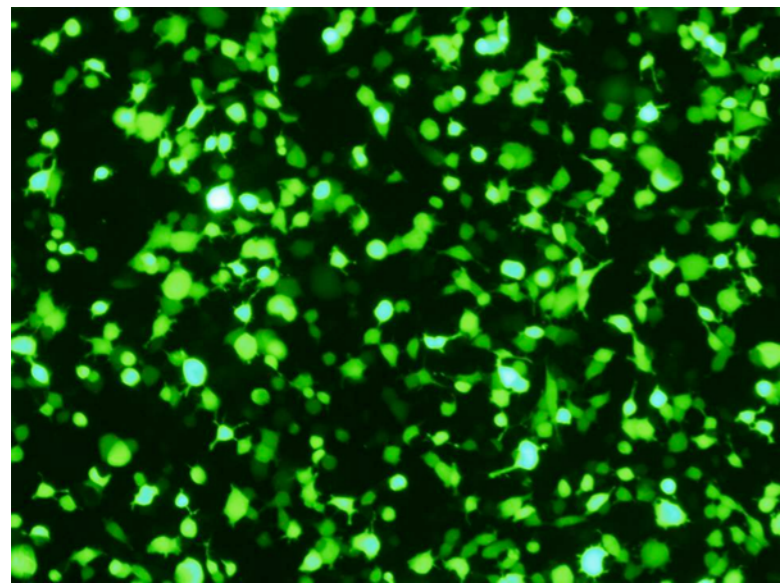
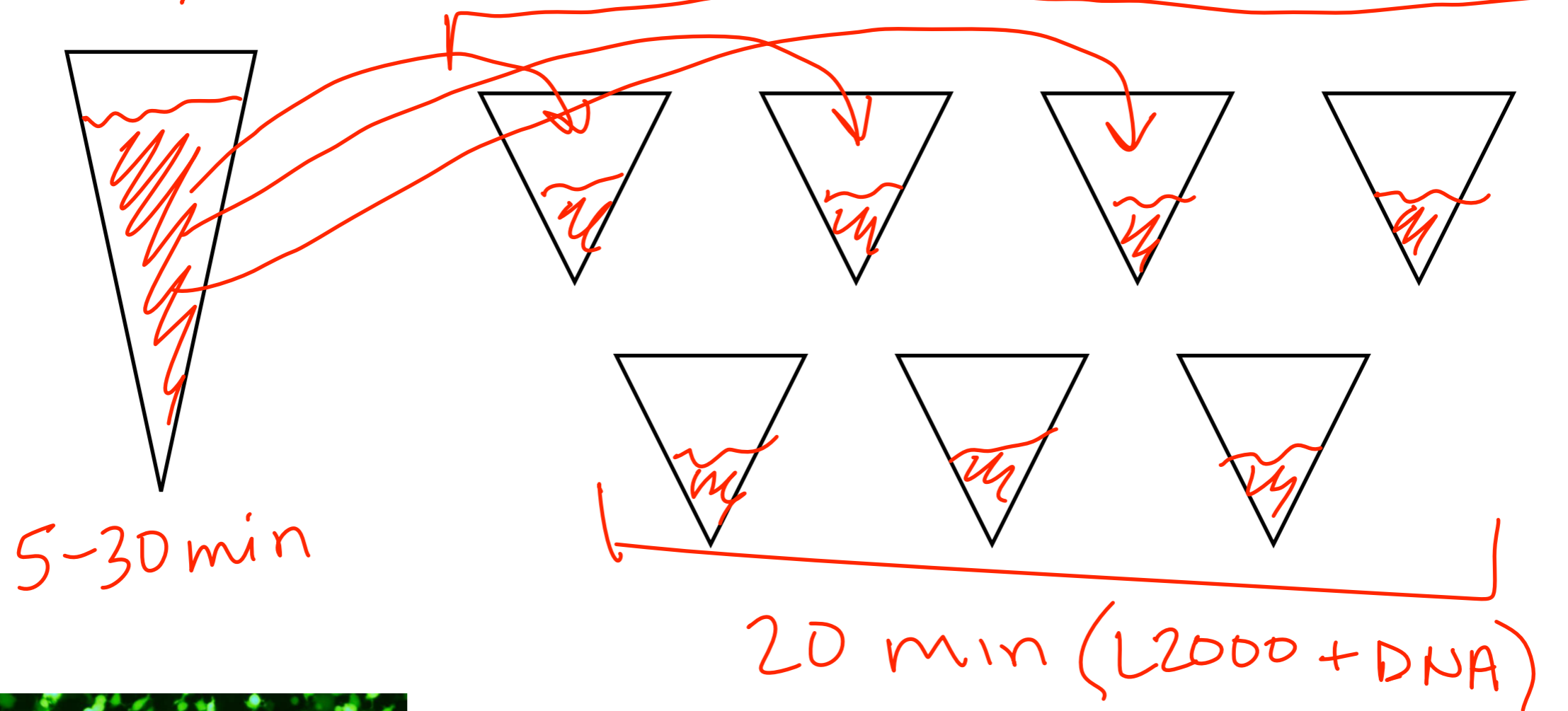
N=3 wells

Class-wide data

# Step 2: Test the system!

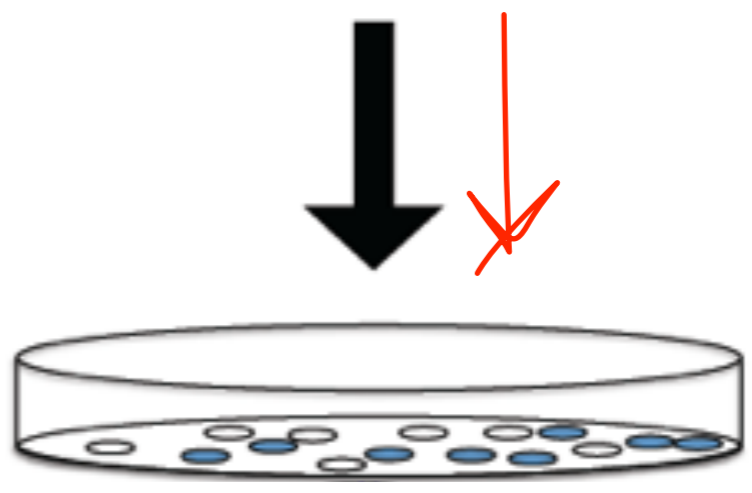
L2000 + OptiMEM

OPTI MEM + DNA



48 hrs

Flow cytometry



# Tissue culture tips

- Set up a few inches *behind* the barrier/grate
- Minimize opportunities to bump or expose sterile equipment or your samples
  - Uncap bottles *before* opening pipet
  - Keep tips and dishes *closed* when not in use
  - Avoid passing your hands/arms over open dishes
  - Don't try to hold > 2 things at once! 😊
- Take care not to clog the pipet-aids

## Today in the lab:

- Set-up lipofections -- pick your conditions and do your calculations BEFORE you start TC
- Look for feedback on Methods ASAP — for sure Thursday in lab, possibly tomorrow night.

## Next time in the lab:

- Sign-up on MID7 Talk Page
- FACS analysis to measure HR efficiency
- Post FACS data on MID7 Talk page promptly!!!