# Metal contamination and bioremediation of agricultural soils for food safety and sustainability

Deyi Hou[1], David O'Connor[1], Avanthi D. Igalavithana[2], Daniel S. Alessi[3], Jie Luo[4], Daniel C. W. Tsang[5], Donald L. Sparks[6], Yusuke Yamauchi[7,8], Jörg Rinklebe[9] and Yong Sik Ok[2] ✉

Abstract | Agricultural soil is a non-renewable natural resource that requires careful stewardship in order to achieve the United Nations' Sustainable Development Goals. However, industrial and agricultural activity is often detrimental to soil health and can distribute heavy metal(loid)s into the soil environment, with harmful effects on human and ecosystem health. In this Review, we examine processes that can lead to the contamination of agricultural land with heavy metal(loid)s, which range from mine tailings runoff entering local irrigation channels to the atmospheric deposition of incinerator and coal-fired power-plant emissions. We discuss the relationship between heavy metal(loid) biogeochemical transformations in the soil and their bioavailability. We then review two biological solutions for remediation of contaminated agricultural land, plant-based remediation and microbial bioremediation, which offer cost-effective and sustainable alternatives to traditional physical or chemical remediation technologies. Finally, we discuss how integrating these innovative technologies with profitable and sustainable land use could lead to green and sustainable remediation strategies, and conclude by identifying research challenges and future directions for the biological remediation of agricultural soils.

Bioavailability
The proportion of a soil constituent that interacts with living organisms.

Soil is a non-renewable resource, generated at a rate of a few centimetres per thousand years[1]. It plays a critical role in supporting ecosystems and human society through providing a habitat for the majority of Earth's species and serving as a medium for crop production[2,3]. However, anthropogenic activities are causing widespread soil contamination and degradation[4,5]. A 2018 study predicted the presence of 2.8 million sites in the EU potentially contaminated with soil pollution[6], and, in China, 19% of agricultural soils contain harmful pollutants at levels exceeding environmental quality standards[7].

The United Nations has set 17 Sustainable Development Goals to be reached by 2030, and eight of these goals rely on a healthy soil environment (FIG. 1). Soil and permafrost together hold the largest terrestrial pool of carbon, storing an estimated 4.1 trillion tonnes — nearly five times the estimated mass of atmospheric carbon[8]. Contaminated soil might not be able to fulfil its role in the carbon cycle, thus, aggravating climate change. The degradation of agricultural soil and the resultant loss of crop yield are particularly alarming, as they put the most vulnerable people on the planet at greater risk of poverty and malnutrition[9]. Moreover, soil pollutants can cause seriously impaired neurological development and life-threatening cancers[10,11] after entering the human body through ingestion of contaminated crops, inhalation of contaminated soil dust or inadvertent ingestion of contaminated soil.

Heavy metals and metalloids, henceforth, referred to as heavy metal(loid)s, are important agricultural soil pollutants due to their toxicity, ubiquity, non-biodegradability and bioavailability for crop uptake, and are a major threat to global food safety and food security[12]. Global agricultural production must double by 2050 to meet the projected demand of a growing population with improved living standards[13]; however, the industrialization of developing countries is causing widespread heavy metal(loid) pollution of agricultural land. The most commonly encountered heavy metal(loid)s in soil include cadmium, arsenic, copper, mercury, lead and chromium; a 2014 national soil survey in China showed that these respectively accounted for 43%, 17%, 13%, 10%, 9% and 7% of all soil quality exceedances[14]. Cadmium is the most widespread and bioavailable heavy metal(loid) in rice paddy soils, leading to concerns that the rice produced is cadmium contaminated[15]. Millions of hectares of agricultural land

✉e-mail: yongsikok@korea.ac.kr
https://doi.org/10.1038/s43017-020-0061-y

**Key points**

- Agricultural soil is a non-renewable natural resource that requires careful stewardship in order to achieve the United Nations' Sustainable Development Goals.
- Global agricultural soil pollution by heavy metal(loid)s represents one of the biggest challenges to sustainable development, particularly in developing countries.
- Bioremediation, including phytoremediation and microbially mediated bioremediation, is a promising nature-based solution for treating heavy metal(loid) contamination.
- It is imperative that the international community realizes the seriousness of the heavy metal(loid)s contamination in soils, takes actions to prevent further pollution and instigates the remediation of contaminated sites with environmentally friendly techniques.
- Policymakers should foster a bioremediation-enabling environment through policy instruments and increased field-based research funding.

are now being taken out of production due to cadmium pollution[15,16], and, despite the increasing stringency of environmental protection regulations, cadmium pollution continues to accumulate[16].

The growing issue of soil pollution has caught the attention of national and international bodies, both governmental and non-governmental[17]. In 2013, the 68th session of the United Nations General Assembly declared December 5th to be 'World Soil Day' and 2015 as the 'International Year of Soils'. In 2017, the United Nations Environment Assembly (UNEA) adopted a resolution that requested a number of bodies to report on global soil pollution[18]. These bodies, including the World Health Organization (WHO) and the Food and Agriculture Organization (FAO), are required to assess the extent of the problem, monitor future trends and identify associated risks and impacts by 2021 (REFS[18,19]). Individual countries are also taking action; China, for example, revealed an ambitious action plan in 2016 to clean up approximately 700,000 hectares of seriously contaminated agricultural land by 2020 and make 95% of the nation's contaminated land safe for use by 2030 (REF.[15]).

In this Review, we discuss the latest findings on the sources of agricultural soil pollution and the natural and anthropogenic processes that influence the distribution of soil heavy metal(loid)s, ranging from local surface runoff to atmospheric transport and deposition. We illustrate how soil heavy metal(loid)s undergo biogeochemical transformation and bioaccumulation in the food chain. Lastly, we review the mechanisms and applicability of plant-based and microorganism-based remediation strategies (phytoremediation and microbial bioremediation) in treating contaminated agricultural soils, and conclude by identifying the challenges and outlook of implementing bioremediation strategies on a large scale.

**Author addresses**

[1]School of Environment, Tsinghua University, Beijing, China.
[2]Korea Biochar Research Center, APRU Sustainable Waste Management Program and Division of Environmental Science and Ecological Engineering, Korea University, Seoul, Republic of Korea.
[3]Department of Earth and Atmospheric Sciences, University of Alberta, Edmonton, Alberta, Canada.
[4]College of Resources and Environment, Yangtze University, Wuhan, China.
[5]Department of Civil and Environmental Engineering, The Hong Kong Polytechnic University, Kowloon, Hong Kong, China.
[6]Department of Plant and Soil Sciences and Delaware Environmental Institute, University of Delaware, Newark, DE, USA.
[7]Australian Institute for Bioengineering and Nanotechnology (AIBN), The University of Queensland, Brisbane, Queensland, Australia.
[8]International Center for Materials Nanoarchitectonics, National Institute for Materials Science (NIMS), Tsukuba, Japan.
[9]School of Architecture and Civil Engineering, Institute of Foundation Engineering, University of Wuppertal, Wuppertal, Germany.

## Occurrence of soil heavy metal(loid)s

Heavy metal(loid)s have been extracted from minerals and used by humans for thousands of years[20] and, currently, are used in a wide variety of industrial, domestic and agricultural applications; chromium and cadmium are frequently used in metal plating, for example. Despite increasing awareness of the harm caused by heavy metal(loid)s in soils, their essential role in modern industry means that their production and use continue to increase. Over the past 50 years, global production of chromium and lead has increased by 514% to 37.5 Mt per year and by 232% to 11.3 Mt per year, respectively[21]. Heavy metal(loid)s are even required for renewable technologies in some cases[22]; cadmium and lead, for example, are used in lead–acid and nickel–cadmium battery cells[20,23,24], lead is used in perovskite solar cells and nickel is used in electric-car batteries[25,26]. Because of their intensive manufacturing, widespread usage and tendency to accumulate via adsorption, absorption and precipitation, heavy metal(loid)s have become the most widely distributed type of contaminants in agricultural soils[27]. Their occurrence in agricultural soils is associated with a wide variety of sources, which are discussed below.

### *Sources of soil pollution*

Anthropogenic sources of heavy metal(loid)s pollution are associated with agriculture, industry and mining. These sources include surface runoff from mine tailings[28,29], soil treatment with impure mineral phosphate fertilizer[30,31] or sewage sludge[32] and irrigation of farmland with polluted water[33,34]. Heavy metal(loid)s present in dusts and aerosols released during mining and smelting activities[35], fossil-fuel burning[36], vehicle use[37], cement manufacture[38] and electronic-waste processing[39] can also enter the soil through atmospheric deposition (FIG. 2).

Different types of contamination are often associated with different sources. Elevated levels of lead, mercury, copper and zinc are often associated with anthropogenic sources[40], and lead is especially associated with transportation activity because of the historical usage of lead in gasoline and the abrasive wear of lead-containing vehicle components[41]. Land treatment with impure mineral fertilizers and manures is associated with cadmium[42,43], copper[42] and zinc[42] contamination in agricultural soil. Irrigation with contaminated wastewater also causes heavy metal(loid)s accumulation[44]. For instance, wastewater land irrigation in a region of Beijing, China approximately tripled soil chromium concentrations over the past 30 years, and increased lead and cadmium levels by factors of 18 and 84, respectively[45].

Atmospheric deposition plays a major role in heavy metal(loid)s accumulation in agricultural soil[42,46]. In Europe, it was found that atmospheric deposition contributes more lead to soils than fertilizer application[43].

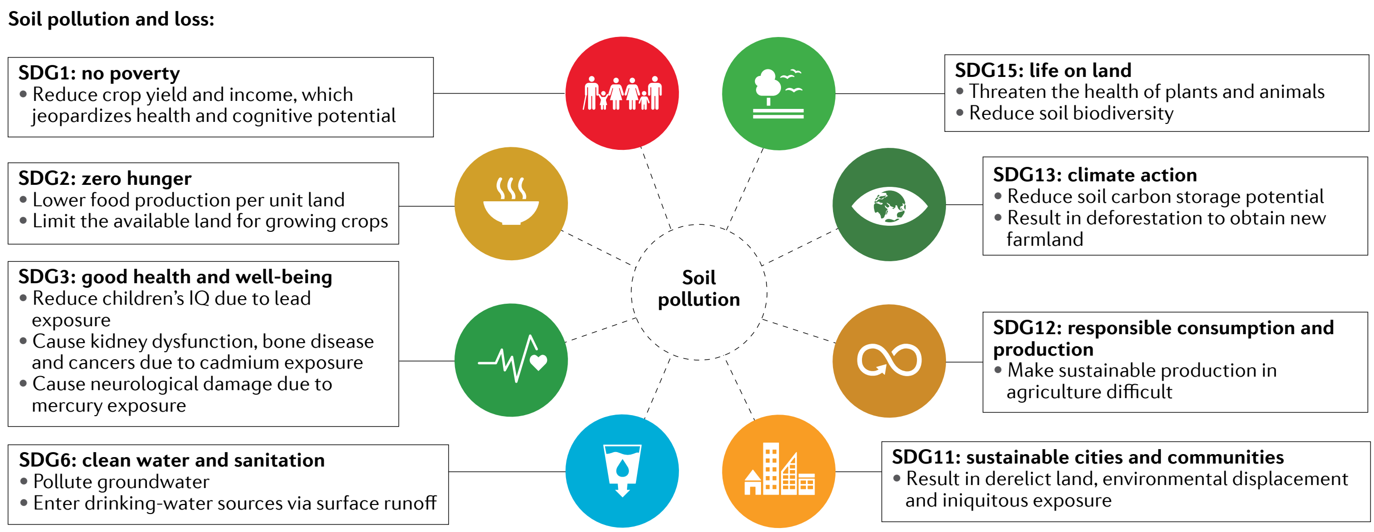


Fig. 1 | **The impact of soil pollution on SDGs.** Soil pollution negatively impacts sustainability, specifically, hindering progress on a number of the Sustainable Development Goals (SDGs) set out by the United Nations.

In England and Wales, atmospheric deposition is the main contributor of heavy metal(loid)s in most agricultural areas, accounting annually for 85% of the mercury, 78% of the lead, 60% of the nickel, 56% of the arsenic and 53% of the cadmium deposited in agricultural soils[46].

Heavy metal(loid)s in agricultural soils can also derive from geogenic sources[42,47,48]. Most existing studies suggest that soil parent material contributes to heavy metal(loid)s concentrations in agricultural soils. Certain heavy metal(loid)s tend to coexist in natural minerals; cadmium, for example, is often associated with zinc, lead or copper in sulfide forms[23]. Geogenic heavy metal(loid)s levels also correlate with soil properties, including clay content, carbonates and soil organic carbon[49]. The coexistence of chromium, cobalt and manganese in soil is indicative of soil heavy metal(loid)s being of lithogenic origin[40]. Some regional studies show that chromium contamination originating from soil parent materials can be associated with nickel[50,51], cadmium[52] and arsenic[53].

### *Heavy metal(loid)s distribution*

Both geogenic and anthropogenic contaminants can accumulate over large spatial areas. Soil pollution in large areas of south-western China is mostly attributable to geogenic sources, and contamination in large areas of China's eastern developed coastal zone is mostly attributable to anthropogenic activity, for instance. However, differentiating the two remains a significant challenge[54]. Spatial distribution can also occur at a smaller scale, even within the same field. The spatial distribution of heavy metal(loid)s is dependent not only upon their sources but also natural factors that generate heterogeneity in soil properties, such as wet–dry cycles and anthropogenic processes such as soil tilling[40].

Variability in heavy metal(loid)s concentrations between fields in the same region can be attributed to chemical transformation[55,56], transportation[57,58], dilution[59] and accumulation[60]. Such processes tend to operate on larger scales than single agricultural plots, reflecting the heterogeneous mineral composition among fields and random distribution of exogenous soil particles with elevated heavy metal(loid)s contents[15,61–63]. Inter-plot variation can also depend on plot distance from pollution sources[64,65], with heavy metal(loid)s concentrations being highest in plots closest to polluted sources of irrigation water, such as contaminated wastewater conveyance channels or polluted natural watercourses[45]. Vehicular pollutants also cause elevated heavy metal(loid)s concentrations in agricultural fields adjacent to major roads[66]. Fields used for growing different crops can differ in heavy metal(loid)s constituents, owing to variations in agricultural cropping and irrigation practices; for example, vineyards in the Piedmont region of Italy have elevated copper and zinc levels due to the application of copper-containing and zinc-containing foliage spray to combat fungal disease[50]. Similarly, sewage-sludge application to individual agricultural plots in England and Wales, despite being conducted under regulatory constraints, was found to supply high levels of heavy metal(loid)s, including zinc, copper, nickel, lead, cadmium, chromium, arsenic and mercury, to the soil[46].

The spatial distribution of heavy metal(loid)s at the regional scale is driven by factors that differ from those controlling variability within and between plots. Such factors include geogenic differences[40], regional atmospheric deposition[67], land-use distribution[68] and the presence of major anthropogenic-emission sources. Heavy metal(loid)s occur naturally in the Earth's crust, and tectonic and weathering processes that result in stratigraphic and sedimentological features largely explain regional variability in topsoil heavy metal(loid)s levels[69]. Atmospheric deposition of parent materials, for example, by dust storms, coupled with anthropogenic atmospheric emissions can result in unique spatial distribution features along wind channels; a study in a major metropolitan area of northern China suggested that the south-eastern winds during the summer season were a key source of heavy metals for the region[67]. Indeed, agricultural fields close to large metropolitan areas are

Geogenic
Resulting from natural geological processes.

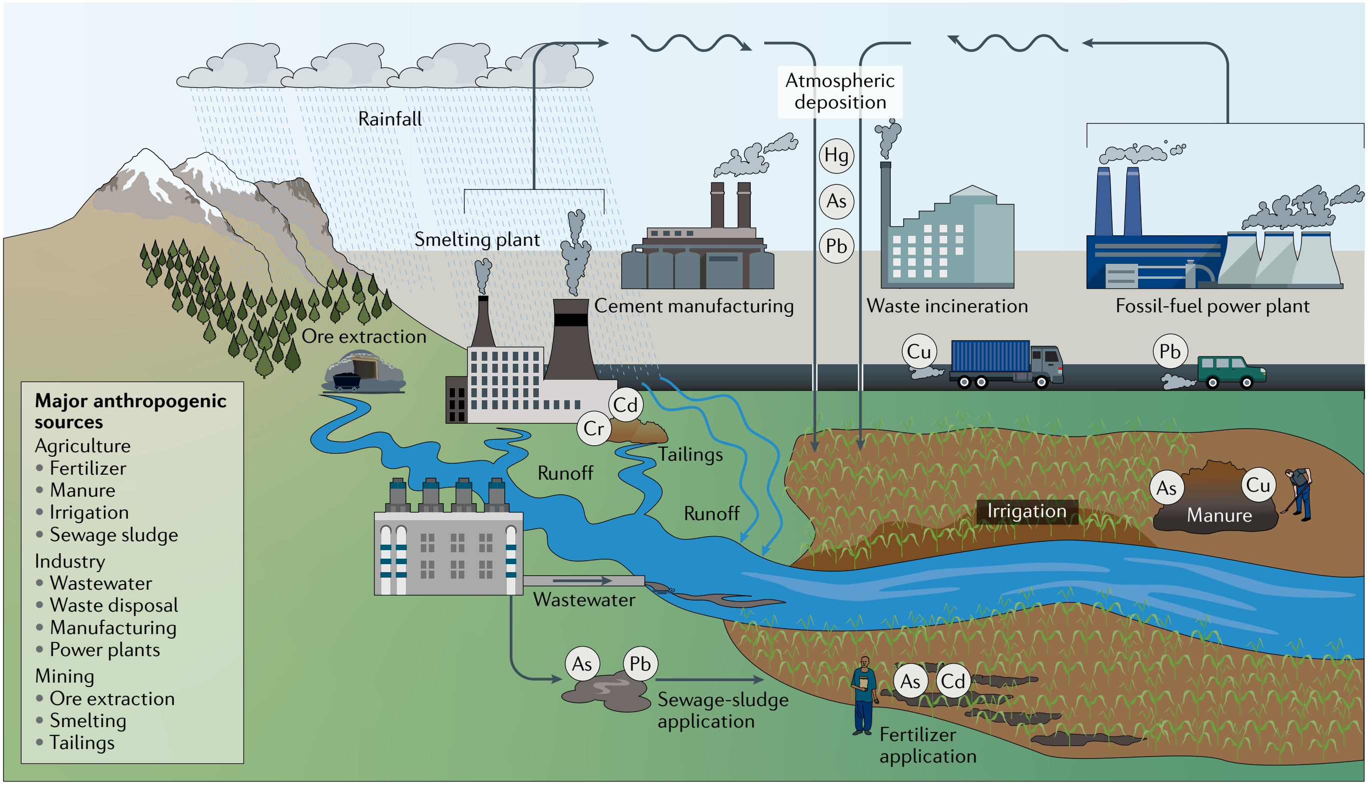


Fig. 2 | **Sources of heavy metal(loid)s pollution in agricultural soil.** Major anthropogenic sources can be classified into three categories: agricultural, industrial and mining. Heavy metal(loid)s can enter agricultural soil through atmospheric deposition, following release into the atmosphere from fossil-fuel burning, waste incineration or cement manufacture. Heavy-metal(loid)s-contaminated runoff from mining and industry can enter waterways and reach agricultural land. The use of manure or sewage contaminated with heavy metal(loid)s to fertilize crops can also contaminate agricultural land.

expected to have higher heavy metal(loid)s concentrations than those in remote areas, owing to more intensive anthropogenic emissions[42,66]. Moreover, major anthropogenic heavy-metal(loid)s-emission sources, such as mega-mining sites, large smelting facilities and large power plants and incinerators without adequate emission control, can result in elevated heavy metal(loid)s in agricultural soil on a regional scale[70,71].

Global mapping of heavy metal(loid)s distribution in agricultural soils is lacking, but airborne heavy metal(loid)s distributions provide an indication of their global distribution in soil. Studies conducted under the United Nations' Convention on Long-Range Transboundary Air Pollution show that atmospheric concentrations of lead, cadmium and mercury are the highest over China, followed by India, the Middle East and northern Africa (FIG. 3a–c). Densely populated areas of Europe and North and South America also have high atmospheric concentrations of these metal(loid)s[72].

Studies mapping soil heavy metal(loid)s concentrations have been conducted on both national and continental scales, though global mapping has not been undertaken. The first harmonized sampling programme of agricultural soils in the EU found that 137,000 km$^2$ of agricultural land requires further local assessment and remediation[27]. A meta-analysis of compiled regional data from south-western China indicates that high heavy metal(loid)s concentrations are present in agricultural fields in a region with high geogenic background concentrations and extensive mining activities[63,73]. In eastern China, industrial facilities that have operated for several decades have caused high heavy metal(loid)s concentrations in agricultural soils[74]. Models using stable mercury isotopic analyses and geospatial climate and vegetation data suggest that South America and East and Southeast Asia have relatively high mercury concentrations in soil[75] (FIG. 3d).

### *Heavy metal(loid)s bioavailability*

Although spatial distributions of metal(loid)s provide useful data on their impact in agricultural soils, the toxicity of heavy metal(loid)s is contingent on their bioavailability, which is, itself, dependent on the oxidation state and specific chemical form of the metal(loid)s[76]. The bioavailability of a given metal(loid) can vary widely depending on the soil type. Only a small fraction of the heavy metal(loid)s in soils are freely available in soil pore water for plant uptake, and dissolved heavy metal(loid)s (usually present as free hydrated ions or complexed ligands) often reach a dynamic equilibrium with the bulk of heavy metal(loid)s existing in the solid phase of the soil[77]. The distribution equilibrium is affected by soil pH, moisture, organic-carbon content, redox conditions, carbonate content, sulfide content, clay minerals and metal-oxide content[23,78–80], factors that can be modified by anthropogenic pollution. For example, irrigation with wastewater can reduce soil pH and increase soil organic matter[45].

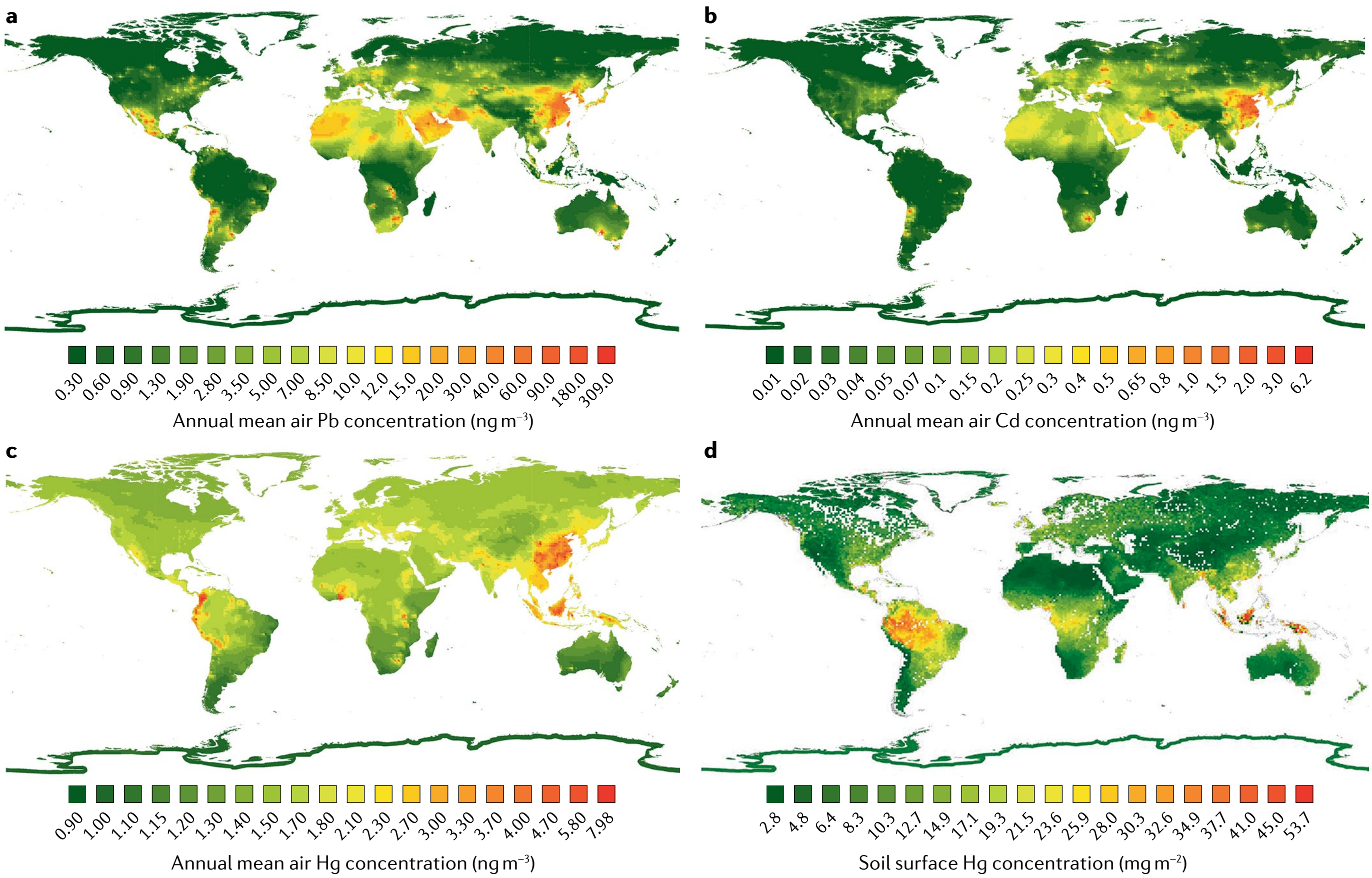


Fig. 3 | **Spatial distribution of heavy metal(loid)s.** The annual mean distribution of heavy metals in the air (panels **a**–**c**) and concentration in the soil (panel **d**). **a** | Lead concentration in air[72]. **b** | Cadmium concentration in air[72]. **c** | Mercury concentration in air[72]. **d** | Mercury concentration in the top 20 cm of soil based on model simulations[75]. Panels **a**–**c** adapted with permission from REF.[72], EMEP. Panel **d** adapted with permission from REF.[75], ACS.

The inherent bioavailability of different heavy metal(loid)s also varies substantially. For instance, the concentration of lead in soil tends to be much higher than that of other heavy metal(loid)s, nearly 40 times higher than cadmium and 100 times higher than mercury, due to the high natural background and high lead-emission levels[23,78]. However, lead has a low inherent bioavailability because it forms insoluble compounds such as pyromorphite and adsorbs strongly to soil minerals such as manganese oxides[81]. In contrast, cadmium has a much higher bioavailability than lead, arsenic and mercury because it exists mainly in exchangeable phases and has a comparatively low adsorption potential[82]. The oxidation state of the heavy metal(loid)s can also change their bioavailability to the plants. For example, the As(III) oxidation state of arsenic causes a decrease in plant growth, stomatal conductance and photosystem II efficiency of *Atriplex atacamensis*, whereas As(V) does not impact these processes[83].

Soil pH is among the most important environmental factors controlling the bioavailability of heavy metal(loid)s[84,85] and can drastically influence the solubility of soil metal(loid)s; for example, cadmium forms insoluble compounds under alkaline conditions (above pH 7.5) but is highly soluble in acidic pH while increasing bioavailability[82]. Local pH and, thus, heavy metal(loid)s bioavailability, are impacted by dynamic biological systems, as in the rhizosphere, where local pH is influenced by root activities and soil amendments[86–88].

Soil organic matter also influences the bioavailability of heavy metal(loid)s, but its effects are complicated. For instance, cadmium can adsorb onto carboxylic, phosphoryl, sulfhydryl and phenolic hydroxyl groups present in organic matter, reducing its bioavailability. However, dissolved humic substances can also form soluble complexes with cadmium, increasing its bioavailability[82]. Humic acids can form complexes with mercury that are highly stable and have low mobility[89,90], whereas compounds of mercury and fulvic acids are more labile and, thus, more bioavailable than mercury–humic-acid complexes[78,91]. Roots release low-molecular-weight organic compounds such as oxalic acid that act as metal chelators, which increase the bioavailability of certain heavy metal(loid)s[47]. Binding to non-organic matter can also influence bioavailability. Among different geochemical fractions of heavy metal(loid)s, exchangeable and easily mobilized metal(loid)s species, such as carbonate-bound metal(loid)s, are more bioavailable and toxic than those species that are less easily mobilized, such as Fe and Mn oxide bound, organic matter bound and residual fractions[92].

Rhizosphere
The very narrow region of soil in the vicinity of plant roots.

Humic acids
Soil organic substances that coagulate when strong base extracts are acidified.

Fulvic acids
Soil organic substances that remain soluble when strong base extracts are acidified.

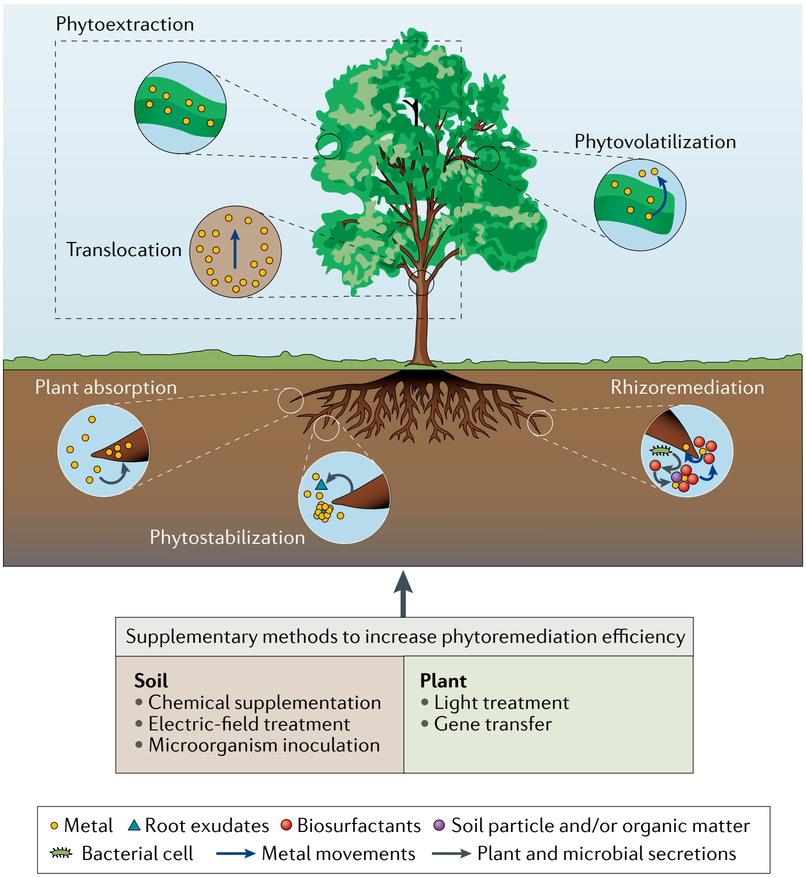


Fig. 4 | **Phytoremediation.** Natural methods of removing or detoxifying soil metal(loid)s, and supplementary methods to increase phytoremediation efficiency[257]. Roots can absorb heavy metal(loid)s from the soil or release factors to stabilize heavy metal(loid)s and prevent bioaccumulation (phytostabilization). Soil microbes can release factors to aid absorption of heavy metal(loid)s (rhizoremediation). Once taken up by roots, heavy metal(loid)s can translocate to the above-ground biomass of the plant, where they can be lost by transpiration (phytovolatilization) or be removed from the field by harvesting the plant. Adapted from REF.[257], CC BY 3.0.

## Bioremediation

The interactions between plants, microbes and heavy metal(loid)s are exploited in bioremediation strategies, which use living organisms for soil decontamination[93,94]. These organisms can be plant or microbial species that are resistant to toxic heavy metal(loid)s and capable of thriving in highly contaminated agricultural soil. Some of these species can adsorb heavy metal(loid)s or release compounds that bind with them[95,96], thus, affecting contaminant bioavailability and toxicity. Other species can extract and remove heavy metal(loid)s from the soil environment[97].

Bioremediation tends to be more sustainable than traditional thermal or physico-chemical techniques such as soil washing, which can remove or destroy living organisms and soil organic matter, jeopardizing long-term soil health and diminishing post-remediation soil productivity[98]. Bioremediation also brings other sustainability benefits, including decreased cost, increased worker safety and smaller life cycle environmental footprints compared with traditional remediation methods[98], maximizing the economic, social and environmental benefits of soil remediation[99]. These benefits have prompted the remediation industry to move towards such nature-based solutions[100,101]. In this section, we discuss three bioremediation approaches: phytoremediation, microbial bioremediation and integrated methods.

### *Phytoremediation*

Phytoremediation for soil decontamination employs indigenous[102] or imported species[103] of plants, including ones that are genetically modified[85,104]. This approach is adaptable to different plot sizes through planting and cultivating an appropriate number of selected phytoremediation plants, and considering intrinsic biogeochemical processes associated with plant growth, metal(loid)s speciation and changes in soil. Phytoremediation techniques include phytostabilization, in which root exudates reduce metal bioavailability in the rhizosphere, and phytovolatilization, which exploits plant evapotranspiration systems to transfer contaminants from the soil to the atmosphere[105] (FIG. 4). However, the most commonly used and well-studied phytoremediation technique is 'phytoextraction'[105]. In this approach, plant species take up heavy metal(loid)s from the soil through their roots; the heavy metals then accumulate in the plant's above-ground biomass, which is harvested. The biomass is typically incinerated, leaving behind a metal-concentrated bottom ash usually disposed of in landfills[106,107]. Particles that result from biomass combustion can pose a health risk and incineration requires appropriate filtration or scrubbing techniques. However, harvested biomass can be used as a feedstock for bioenergy production[108–110] or pyrolyzed to form biochar[111], with appropriate safety considerations.

***Soil–plant–metal interactions.*** Heavy metal(loid)s enter plant tissue through various pathways (FIG. 4). For example, on a molecular level, plant-root systems are not completely selective and will take up heavy metal(loid)s from interstitial soil water, such as cadmium and arsenic, that have properties similar to nutrients required by the plant, such as zinc and calcium[112–114]. After entering root systems, heavy metal(loid)s are translocated from the roots to shoots and leaves, and then to fruits or seeds. Studies have also shown that heavy metal(loid)s will also enter plants from the atmosphere via foliar transfer[115], although unlike soil–root transfer, the molecular mechanisms involved in atmosphere–leaf transfer are not well understood.

Heavy metal(loid)s concentrations, the presence of chelating compounds, plant characteristics and soil properties all affect soil–plant–metal interactions and plant uptake rates[116–120] and, therefore, the effectiveness of phytoremediation. For instance, metal(loid) ions can form insoluble complexes, causing precipitation on soil-particle surfaces that inhibits their uptake by plants. Complexation of heavy metal(loid)s with larger molecules of soil organic matter can also hinder plant uptake. Plant $Fe^{2+}$ uptake systems are upregulated in iron-deficient soil, allowing more $Cd^{2+}$ to be taken up by root cells through $Fe^{2+}$ channels[121]. Similarly, elevated concentrations of $Zn^{2+}$ and $Ca^{2+}$ can act as competitors to $Cd^{2+}$ for plant uptake[112], mitigating cadmium toxicity towards the plant or enhancing essential mineral-element uptake[113]. Methods to augment biomass generation and heavy metal(loid)s uptake have been developed, including the application of artificial light[122,123], chemical supplementation[124,125], electrical-field treatment[126], microorganism inoculation[127] and gene transfer[128].

***Plant selection.*** Plant selection is a critical step in phytoremediation, as species vary widely in their ability to uptake or immobilize different contaminants. Although indigenous species are preferred because they are adept at surviving in local environmental conditions, the use of introduced species might be necessary to speed up remediation[105]. Regardless of origin, hyperaccumulators[129,130] (plant species that extract large amounts of heavy metal(loid)s) are advantageous to use as they can speed up remediation of sites contaminated with high levels of heavy metal(loid)s. However, as they tend to take up a limited variety of heavy metal(loid)s[129], hyperaccumulators might not be suitable for soils contaminated with many different metal(loid)s species[131]. In these cases, fast-growing and high-biomass phytoremediation plants including willow, eucalyptus and poplar trees can be used to extract a wide range of heavy metal(loid)s from soil, although application of such plants may prevent agricultural production for years or possibly decades due to their relatively slow metal-extraction rates[132–134]. However, it is feasible for farmers to apply intercropping techniques, which enable the growth of phytoremediation plants alongside agricultural crops[135,136].

Hyperaccumulators are deemed most suitable for use on occupied agricultural sites to reduce heavy metal(loid)s contamination in developing countries, where there is great pressure for crop production[137]. To assist in this context, research has focused on the identification of hyperaccumulator species that are able to grow alongside crops[138–140], allowing remediation of the soil and the prevention of contaminants from endangering food crops simultaneously. Other research has focused on the use of crop plants that are able to take up contaminants but do not bioaccumulate them in edible parts; for example, crops may produce a grain that is suitable for animal consumption, while contaminants are enriched in the shoots or roots, which can then be removed as part of a phytoextraction strategy[141]. The main drawback to this approach is that heavy-metal(loid)s-extraction rates can be relatively low and use of these plants can prevent the growth of more valuable crops.

A wide variety of hyperaccumulator species specific for a range of metal(loid)s species have been identified, including the Cretan brake fern *Pteris cretica* for arsenic[142], *Sedum plumbizincicola* of the Crassulaceae family for cadmium and zinc[143,144], the grass species *Pogonatherum crinitum* for lead[145], *Celosia argentea* (the plumed cockscomb or silver cock's comb) for manganese[146] and *Pronephrium simplex* of the Thelypteridaceae family for rare-earth elements[147]. The increasing pool of hyperaccumulators, screened and selected from nature, offers new options to tackle difficult-to-treat sites. Meanwhile, researchers are developing transgenic plants to enhance the resistance, volatilization and accumulation of heavy metals in selected plants[148,149]. However, biosafety remains a concern due to potential transfer of conditional lethality and antibiotic-resistance markers to higher levels of the food chain[150,151].

**Hyperaccumulators**
Plant species that extract and concentrate certain heavy metal(loid)s within their biomass when grown in metal-contaminated soils.

***Field successes and challenges.*** Many laboratory studies on phytoremediation have been conducted on spiked soils containing concentrations of heavy metal(loid)s hundreds or thousands of times higher than those found at contaminated sites[152–154]. The rationale for this method is to identify hyperaccumulator species more easily and better elucidate the molecular mechanisms at work by subjecting plants to high stress levels. However, care must be taken in extrapolating the data from such experiments and applying it to field operations.

In recent years, there has been an increasing number of field trials to verify the effectiveness of phytoremediation strategies at more environmentally relevant concentrations, as well as to determine field-related factors influencing their efficiency[155]. Phytoremediation field trials have been carried out globally, with the majority conducted in China[105] and preliminary large-scale (>500 $m^2$) phytoremediation field trials have been carried out for various heavy metal(loid)s contaminants in China, Switzerland, Germany, France and so on[105]. These studies provide evidence to evaluate the resilience, stability, suitability and effectiveness of phytoremediation plants under various environmental conditions. The initial large-scale phytoremediation field trials on heavy-metal(loid)s-contaminated soils were conducted in the early 1990s[156], when it was suggested that this approach could reduce metal concentrations to acceptable ranges on otherwise productive land. Since then, many greenhouse pot studies and small-scale outdoor field trials have been conducted, which have confirmed various hyperaccumulator species as being effective in reducing the soil concentration of a range of heavy metal(loid)s and helped identify practices to increase uptake levels. Plant density[157], initial plant size[119], cropping and harvesting strategies such as double cropping[158,159], transplantation and double harvesting[158,159] have been identified as crucial factors affecting success in these studies; however, these small-scale field studies, often conducted at the metre scale, suffer from inconsistency owing to their small size. Phytoremediation efficiency is also affected by soil heterogeneity[160]; hyperaccumulators are often identified and selected for highly contaminated soils, but may be less effective in soils with a lower degree of contamination. In addition, influencing parameters can vary during field treatments, meaning that long-term studies that report annualized treatment efficiencies are preferable to shorter trials[161,162].

More recently, larger, hectare-scale field trials have been performed. Variability in the results from these larger trials tends to be lower than that seen among smaller studies (TABLE 1). An agricultural trial across 11.1 hectares grew the hyperaccumulator species *P. vittata* and *S. alfredii* at a site previously contaminated with lead (351 ppm), cadmium (320 ppb) and arsenic (37 ppm) in the Guangxi Zhuang Autonomous Region in

Table 1 | **Results of large-scale phytoremediation field studies in agricultural soil polluted by heavy metal(loid)s**

| Plant species | Plot size ($m^2$) | Soil texture | Soil pH | Initial soil OM ($g\,kg^{-1}$) | Initial soil HM ($mg\,kg^{-1}$) | BCF[a] | | TF[b] | Metal(loid)s removal[c] | Key findings | Ref. |
|---|---|---|---|---|---|---|---|---|---|---|---|
| *Morus alba* | 600 | – | 6.9 | – | Cd (3.2) | 1st year | <0.09 (s) | <0.3 (s) | 3–7 g ha$^{-1}$ year$^{-1}$ | Cd and Pb mostly accumulate in root tissue, but not in fruits, indicating the trees could be used as a crop substitute | 251 |
| | | | | | | 2nd year | <0.08 (s) | <0.3 (s) | 2–8 g ha$^{-1}$ year$^{-1}$ | | |
| | | | | | Pb (181.2) | 1st year | <0.02 (s) | <0.6 (s) | 40–85 g ha$^{-1}$ year$^{-1}$ | | |
| | | | | | | 2nd year | <0.008 (s) | <0.2 (s) | 10–42 g ha$^{-1}$ year$^{-1}$ | | |
| *Zea mays* | 675 | Silt loam | 5.8 | 53 | Pb (5,844.2) | 0.06 (r) | | – | 7,181 g ha$^{-1}$ year$^{-1}$ | Each hectare can produce ~25 tonnes of corn grain for animal feed; biomass can generate bioenergy fuel equivalent to 1,545 GJ | 141 |
| | | | | | | 0.01 (s) | | 0.25 (s) | | | |
| | | | | | | 0.04 (l) | | 0.69 (l) | | | |
| *Solanum nigrum* | 1,500 | Sandy loam | 6.2 | 138 | Cd (1.91) | 5.2 (ap) | | – | <233 g ha$^{-1}$ | The plants accumulated Cd in their biomass, enhanced by double cropping and sequential harvesting | 158 |
| *Averrhoa carambola* | 1,500 | Loam | 6.1 | 43 | Cd (1.6) | – | | – | 213 g ha$^{-1}$ | High-density *A. carambola* removed 5.3% of the total Cd within one season; this decreased Cd bioavailability and uptake (63–69%) by vegetables grown afterwards | 252 |
| *Salix* sp. | 1,710 | Sandy loam | 4.0 | 30 | Cd (2.8) | 3.61 (ap) | | 0.60 (ap) | 95 g ha$^{-1}$ | Repeated harvesting of the woody plants prior to leaf fall ensured effective soil decontamination | 253 |
| | | | | | Pb (283) | 0.02 (ap) | | 0.38 (ap) | 55 g ha$^{-1}$ | | |
| | | | | | Zn (295) | 1.16 (ap) | | 0.29 (ap) | 3,320 g ha$^{-1}$ | | |
| *Salix* sp. | 2,100 | Sand | 6.6 | – | Cd (6.5) | 4.3 (s) | | – | 88 g ha$^{-1}$ year$^{-1}$ | Certain *Salix* species produced up to 12.5 tonnes of dry biomass per hectare per year; Cd and Zn removal increased by 40% with leaf harvest | 254 |
| | | | | | | 9.2 (l) | | | | | |
| | | | | | Zn (377) | 1.8 (s) | | | 3,497 g ha$^{-1}$ year$^{-1}$ | | |
| | | | | | | 10.8 (l) | | | | | |
| *Zea mays* and *Pteris vittata* | 400 | – | 6.4 | – | As (93.6) | 5.51 (l) | | 8.1 (l) | 113 g ha$^{-1}$ | Phytoaccumulators grown with maize, limiting As accumulation in maize grains; planting crops in different angular directions improved soil nutrient availability and As uptake | 255 |
| *Zea mays* | 4,050 | Sand | 6.0 | 50 | Cd (67) | 0.01 (s) | | – | 6.4–10.4 g ha$^{-1}$ | Produced biomass for generating 33,000–46,000 kWh of renewable energy per hectare per year | 256 |
| | | | | | Pb (184) | 0.02 (s) | | | 28–46 g ha$^{-1}$ | | |
| | | | | | Zn (355) | 0.41(s) | | | 1,447–2,826 g ha$^{-1}$ | | |
| *Salix* sp. | 10,000 | – | 5.6 | 19 | Cd (5.7) | 9.82 (l) | | – | 82–113 g ha$^{-1}$ year$^{-1}$ | Several decades of phytoremediation with *Salix* required to reduce the Cd content of the soil from 5 to 2 mg kg$^{-1}$, but could be used for bioenergy feedstock | 164 |
| *Pteris vittata* and *Sedum alfredii* | 111,000 | – | – | – | Cd (0.32) | – | | – | 85.8% (re) | Phytoremediation decreased soil HM concentrations below national standards at a cost of US\$75,375.20 ha$^{-1}$ or US\$37.70 m$^{-3}$ of soil, lower than traditional remediation technologies | 163 |
| | | | | | Pb (350.5) | | | | 30.4% (re) | | |
| | | | | | As (36.66) | | | | 55.3% (re) | | |

–, data not available; (ap), above-ground part; As, arsenic; BCF, bioaccumulation factor; Cd, cadmium; HM, heavy metal(loid); (l), leaf; OM, organic matter; Pb, lead; (r), root; (re), removal efficiency; (s), stem; TF, translocation factor; Zn, zinc. [a]The BCF represents the ratio of pollutant concentration in the organism to the soil. [b]TF is the ratio of HMs in the shoots and roots of a plant. It represents the ability of a plant to translocate the metal(loid)s from roots to shoots and/or leaves. Only trials with plot sizes larger than 500 $m^2$ are shown. Heavy-metal concentrations represent the mean total concentration for the whole plant, unless stated otherwise. [c]Removal represents grams of HMs removed per hectare, unless stated otherwise.

Siderophores
Chelating compounds secreted by microorganisms that bind with iron and other metals, increasing their bioavailability.

south-western China[163]. After two years, soluble concentrations of lead, cadmium and arsenic were reduced by 30.4%, 85.8% and 55.3%, respectively. Other large field trials have generally demonstrated that phytoremediation is a promising remedial approach, which can be far more cost-effective than traditional alternatives[163].

Recent studies have aimed to evaluate and enhance the sustainability of phytoremediation. The assessment of a short-rotation willow coppice phytoremediation trial at a heavy metal(loid)s-contaminated agricultural site in Belgium showed this approach to be the most sustainable alternative among various remediation options, due to its capacity to capture atmospheric carbon in plant biomass, while simultaneously treating soil contamination[164]. The use of organic amendments derived from biological waste (such as compost[165], sewage sludge[166] and manure[167]) and industrial waste (such as fly ash and red mud) to enhance phytoremediation have been explored, but some debate remains as to whether these amendments contain harmful levels of contaminants themselves[168–170] and if they actually improve phytoremediation performance[105].

Overall, field trials have been somewhat inconsistent in effectiveness, even among plants of the same species grown in the same plot[171,172]. There are several plausible reasons for this inconsistency. First, phytoremediation efficiency is highly influenced by environmental conditions such as contamination level, soil clay content, weather patterns, soil moisture, organic matter content, pH and salinity[173,174]. Although these parameters are generally homogenous and set at favourable levels in indoor pot experiments, field environments can be highly heterogeneous. Extrapolation of laboratory results might have caused overly optimistic predictions of uptake levels of heavy metal(loid)s in the field[175]. Second, the success of phytoremediation depends on the location of field sampling points, but field heterogeneity causes large differences from one sampling point to the next, often making the interpretation of results challenging.

### *Microbial bioremediation*

Microorganisms exist at high concentrations in agricultural soils[176–178] and possess genes enabling their survival in contaminated soil environments. Many microorganisms are genetically resistant to heavy metal(loid)s[179] and some can survive even under extreme heavy metal(loid)s stress. Native microbes can facilitate the reduction of soil pollution levels or microbes (sometimes, ones that have been genetically engineered) can be introduced to polluted sites to reduce soil metal(loid)s concentrations in a process known as microbial biomediation[180]. Microbe–heavy-metal(loid)s interactions have been studied intensively[181] and a diverse range of microbial species and mechanisms that transform metal(loid)s to chemical species of lower solubility for immobilization, or species of higher solubility for removal[181,182], have now been identified[180,183,184] (FIG. 5). Here, we discuss soil–microorganism–metal interactions and two microbial bioremediation approaches: monitored natural attenuation and engineered microbial bioremediation.

***Soil–microorganism–metal interactions.*** Biogeochemical processes facilitated by microbial activities form the basis of microbial bioremediation[185,186]. A crucial mediator of remediation[187] is the bacterial secretion of siderophores, which primarily transport iron from low-iron soils to cells through specific receptor and transport systems[188]. Fortuitously, siderophores also bind to heavy metal(loid)s[187,189–191]; for example, the bacteria *Alcaligenes eutrophus* secretes siderophores that bind with cadmium, zinc and lead[192]. Bacteria then protect themselves from siderophore-bound heavy metals by producing outer-membrane proteins that facilitate the formation of bioprecipitates, which have low environmental risk

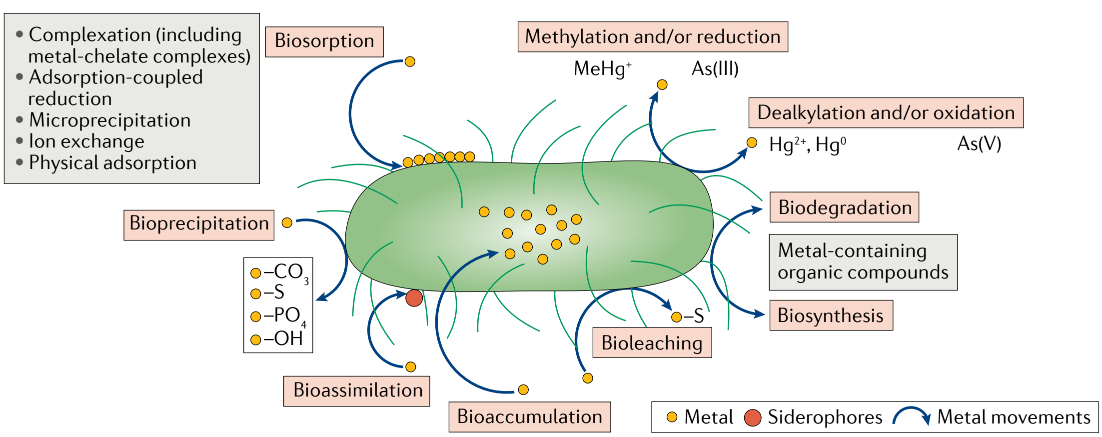


Fig. 5 | **Microbial bioremediation.** Processes by which bacteria can mediate the removal or detoxification of heavy metal(loid)s from agricultural soil. Bacteria can interact with heavy metal(loid)s directly, accumulating them on the cell surface (biosorption). They can also reduce or oxidize metal(loid) species and synthesize or degrade metal-containing organic compounds via catalytic reactions (biosynthesis or biodegradation). Sulfur-oxidizing bacteria can release acids and dissolve metal-containing compounds for leaching of metals (bioleaching). Sulfate-reducing bacteria can precipitate metals by formation of low-mobility sulfides (bioprecipitation). Bacteria can also accumulate metals in the intracellular space by using proteins in their cellular processes (bioaccumulation). Bacteria assimilate metals via iron-assimilation pathways using siderophores (bioassimilation). $CO_3$, carbonate $CO_3^{2-}$; OH, hydroxyl $OH^-$; $PO_4$, phosphate $PO_4^{3-}$; S, sulfide $S^{2-}$. Adapted with permission from REF.[258], Elsevier.

**Lime**
Calcium-rich alkaline-soil amendments, including marl, chalk, limestone or hydrated lime.

due to low bioavailability[192]. As siderophore production is boosted when iron levels are low, applying lime to contaminated soils can enhance bioremediation efficiency by reducing iron availability to the microbes[193].

Bioleaching and bioprecipitation are mechanisms of microbial bioremediation that rely on the presence of sulfur-oxidizing bacteria (SOB) and sulfate-reducing bacteria, respectively, and play a crucial role in determining the relative abundance of the common oxidation states of sulfur in nature[194]. Bioleaching involves the release of metal ions through the mineral dissolution, for example, by sulfuric acid produced by SOB. Dissolution of metals by sulfuric acid subsequently increases sulfur bioavailability to sulfate-reducing bacteria, which facilitates the bioprecipitation of metals as low-mobility sulfides, effectively removing them from the reactive-metal pool. Laboratory experiments on a multi-metal-contaminated soil by inoculating an SOB showed that metal bioleaching levels were as high as 74% for cobalt, 69% for copper, 69% for manganese and 68% for nickel after six months, the majority of which (80–98%) bioprecipitated as stable sulfide species[195]. Subsequent studies exploited sulfur-cycle bacteria to reduce risks posed by zinc, copper, chromium, lead and nickel in the soil. In this case, the study found that introduced microbial species were able to outperform indigenous species due to their higher sulfur-oxidizing activity[196]. Bioleaching of heavy metal(loid)s from exchangeable, carbonate-bound, Fe-oxide-bound and/or Mn-oxide-bound, organic-matter-bound and residual fractions were observed after only one month because of SOB activity[196]. Several bacterial species associated with the natural sulfur cycle, such as *Acidithiobacillus* spp., *Acetobacter* spp., *Arthrobacter* spp. and *Pseudomonas* spp., can be exploited for bioleaching of heavy metal(loid)s in soils and some fungi species, including *Penicillium* spp., *Aspergillus* spp. and *Fusarium* spp.[197–200]. The activity of *Penicillium chrysogenum*, for example, can mobilize cadmium, copper, lead and zinc in contaminated soils, leading to enhanced bioleaching[201,202].

Biological reduction provides another important route for microbially assisted soil remediation because the toxicity of heavy metal(loid)s depends on their oxidation state. For example, hexavalent chromium (Cr(VI)) is toxic and carcinogenic, whereas trivalent chromium (Cr(III)) is considered non-hazardous, which enables the remediation of Cr(VI)-contaminated soils by reduction. Bacterial strains resistant to and able to reduce elevated Cr(VI) concentrations include *Pseudomonas fluorescens, P. aeruginosa* and *Enterobacter cloacae*[203]. This biological reduction process can be achieved in inundated soils (such as in rice paddies), where oxygen levels are low, or in artificially induced reductive environments, for example, those in which an electron donor is added to induce microbial growth[204,205].

***Monitored natural attenuation.*** The risk posed by heavy metal(loid)s in soil environments can naturally attenuate over time without specific remedial treatment[206]. This phenomenon has been observed at abandoned historic mining sites and adjacent agricultural fields throughout the world[207]. Natural attenuation processes comprise biological, physical and chemical mechanisms[208–210], but the activities of indigenous microbes often drive attenuation; these activities include metal(loid)s sequestration, ion efflux (which can lead to metal precipitation as carbonates near and around cells)[211,212] and extracellular chelation. Indigenous microbes can also mediate biogeochemical reactions that convert mobile heavy metal(loid)s into stable compounds of low bioavailability[208] through adsorption of metal(loid)s to organic matter[213], the formation of carbonates and sulfides (facilitated by *Kocuria flava*, *Sporosarcina pasteurii* and *Terrabacter tumescens*)[214,215], binding to iron and manganese oxides[216], reduction of metal(loid)s to aid the formation of stable compounds (by *Escherichia coli*, *Staphylococcus aureus* and *Staphylococcus xylosus*, for example)[208,217] and the oxidation and hydrolysis of aluminium, iron and manganese species (FIG. 5).

Natural attenuation often takes years or decades to reduce risk levels, although it remains a viable option for remediation when coupled with an appropriate and robust monitoring plan[218]. In some cases, bioremediation based on monitored natural attenuation may be the only practicable option to lower risk, given the difficulties and high costs inherent in treating some agricultural sites, particularly in developing countries[208]. At these sites, agricultural soil-management approaches, such as no-till farming and the use of cover crops, can influence microbial respiration and plant growth[219,220], thus, influencing natural-attenuation rates. For instance, no-till farming increases microbial biomass, soil carbon content and the activity of microbial enzyme activities (such as dehydrogenases, cellulases, xylanases, β-glucosidases, phenol oxidases and peroxidases) in agricultural soil[221], which can accelerate the formation of stable fractions of heavy metal(loid)s.

***Engineered microbial bioremediation.*** Two types of engineered microbial bioremediation exist: biostimulation and bioaugmentation. Biostimulation involves providing indigenous soil microbes with additional nutrients, electron donors or electron acceptors in order to increase their capacity for immobilizing or degrading contaminants in the soil. This approach has been used in remediating heavy metal(loid)s[222], gasoline additives[223], broad-range hydrocarbons[224] and radionuclides[225]. Although indigenous microbes are often excellent candidates for bioremediation because they are acclimated to site conditions[226], laboratory-grown microbial strains can be added to soil, a process known as bioaugmentation[227]. Current commercial bioaugmentation applications use microorganisms collected from environmental samples and enriched in laboratories. There are many successful biofertilizers produced from plant-growth-promoting microorganisms and applied safely in the field[228,229], but, often, microorganisms cultivated under controlled conditions do not survive once placed in competition with indigenous microorganisms in field conditions[230]. Further research is needed to improve the performance of these cultivated microorganisms under field conditions for heavy metal(loid)s immobilization.

Genetic-engineering techniques can be used to improve microbial mechanisms for heavy metal(loid)s resistance[231]. For example, the introduction of a gene encoding phytochelatin synthase from *Schizosaccharomyces pombe*, *SpPCS*, into *E. coli* can enhance the cadmium uptake of *E. coli* by 7.5-fold[232]. Phytochelatins, the protein products of phytochelatin synthase activity, bind strongly to toxic elements such as cadmium, arsenic, lead and mercury[232–234], and render them non-toxic. Insertion of a gene encoding arsenite S-adenosyl methionine methyltransferase (*arsM*) into *Sphingomonas desiccabilis* and *Bacillus idriensis* similarly enables a tenfold increase in the production of methylated arsenic gas[235], allowing arsenic to volatize from the soil. Transferring genes from heavy-metal(loid)s-resistant microbes to other microbial species suitable for microbial bioremediation has potential to increase bioremediation effectiveness, subject to regulatory approval and oversight[212]. However, biosafety issues, including the possibility of horizontal gene transfer, must be taken into account before introducing these organisms into the environment[230].

### *Integrated methods and phytomanagement*

Microbially mediated processes can enhance the efficiency of phytoremediation[236] by transforming heavy metal(loid)s, rendering metabolic nutrients and minerals more bioavailable to aid plant growth, stimulating systems that regulate plant heavy metal(loid)s stress responses or aiding the production of plant hormones that increase plant growth[237] (FIG. 4). The bacterial species *Pseudomonas aeruginosa, Pseudomonas fluorescens* and *Ralstonia metallidurans* produce siderophores that increase contaminant bioavailability to roots, leading to enhanced phytoextraction efficiency[238]. For example, augmentation of soil with these strains can increase chromium and lead uptake by plants by as much as 5.4-fold[238]. Moreover, microorganisms such as species of *Bacillus*, *Achromobacter*, *Stenotrophomonas*, *Brevundimonas*, *Ochrobactrum*, *Pseudomonas*, *Microbacterium*, *Comamonas* and *Sinorhizobium* can lower the toxicity and increase bioavailability of arsenic to plants by increasing arsenic mobilization[239]. A bacterial consortium of *Bacillus methylotrophicus*, *Bacillus aryabhattai* and *Bacillus licheniformis* applied to phytoremediation sites promotes plant growth through enhancing nitrogen fixation and phosphate solubilization, and producing siderophores and other molecules that affect plant hormonal processes. This consortium, when applied to *Spartina maritima*, effectively improved root growth by approximately 60% and bioaccumulation of cadmium, arsenic, copper, lead and zinc by between 17% and 65%[240].

The most significant drawback to bioremediation is the time required to complete treatment, which is sometimes overcome through its coupling with other remediation technologies to shorten treatment length. For example, the production of $H^+$ and $OH^-$ ions during electrokinetic treatment of soil can produce potential gradients that cause unwanted bands of high residual metal concentration[241]; these issues are mitigated by phytoremediation techniques, as plant roots can extract $H^+$ and $OH^-$ and residual heavy metal(loid)s[242]. Moreover, electrical fields induced by electrokinetic treatment can transport pollutants from deep in the soil up to the rhizosphere, enhancing phytoremediation effectiveness[126]. *Solanum tuberosum* showed higher zinc and copper accumulation in plant root under supplement of alternating current compared with the control in a laboratory study[243].

The integration of remediation technologies provides a scenario where ecosystem services such as nutrient cycling, carbon sequestration and water storage are restored[244]. Moreover, plants grown in contaminated agricultural fields undergoing bioremediation can be sold as bioenergy products or other profitable products[245]. Moreover, in comparison with traditional remediation strategies, phytomanagement focuses on both risk mitigation and commercial viability by using plants to control contamination while producing marketable biomass, and has been suggested as a viable strategy that can be carried out in large-scale applications[246]. Phytomanagement is considered as either a low cost or a profitable strategy for producing valuable plant biomass such as bioenergy or timber crops, or it can be used to prevent decreased food production on contaminated lands[246,247].

## Summary and future perspectives

The accumulation of heavy metal(loid)s in agricultural soils is an obstacle to achieving global food safety and security. Bioremediation is a promising nature-based solution for treating heavy metal(loid)s contamination; however, several issues must be addressed before it can be more broadly implemented.

First, it will be beneficial to accelerate global soil mapping and establish regional models that can adequately predict contaminant distributions and identify pollution sources[248]. Second, the measured effectiveness of bioremediation in the field has been somewhat inconsistent, attributed to heterogeneity in field conditions and artefacts caused by evaluating treatments on a spot-by-spot basis, rather than employing field-wide assessment. Importantly, variability tends to decrease with increasing plot size[249], showing the importance of large-scale field trials. Third, field stations are needed to provide valuable insights into the mechanisms that render heavy-metal(loid)s-contaminated sites resistant to treatment. We suggest there is a need for improved monitoring instrumentation to measure trends in microbial dynamics, metal speciation and fractions, and soil environmental conditions (pH, temperature, redox potential and soil gases), as all these factors can mediate bioremediation effectiveness. Fourth, further research is required in order to decrease clean-up time and expand the applicability of bioremediation techniques to include more sites. Seeking out new natural species for this purpose and developing new genetic technologies that can modify and design the functionality of plant species and microbial strains could play a leading role in future development.

Global agricultural soil pollution by heavy metal(loid)s represents one of the biggest challenges for sustainable development, and developing countries are particularly vulnerable to this threat to food, health and livelihoods. By the 5th session of the United Nations Environment Assembly in 2021, institutions including the WHO and the FAO will elaborate on guidelines for the prevention and minimization of soil contamination, specifically including the use of nature-based solutions[250]. In this context, it is imperative that the international community realizes the seriousness of the threat, takes actions to prevent further pollution and instigates the remediation of contaminated sites with environmentally friendly techniques. Policymakers should foster a bioremediation-enabling environment through policy instruments and increased field-based research funding.

Published online 23 June 2020


1. Montgomery, D. R. Soil erosion and agricultural sustainability. *Proc. Natl Acad. Sci. USA* **104**, 13268–13272 (2007).
**Provides evidence from 201 different field studies globally that the soil-erosion rates in no-till agriculture is similar to soil-production rates and can be considered as a sustainable agricultural practice**.
2. Cassidy, E. S., West, P. C., Gerber, J. S. & Foley, J. A. Redefining agricultural yields: from tonnes to people nourished per hectare. *Environ. Res. Lett.* **8**, 034015 (2013).
3. Jansson, J. K. & Hofmockel, K. S. Soil microbiomes and climate change. *Nat. Rev. Microbiol.* **18**, 35–46 (2020).
4. Foley, J. A. et al. Solutions for a cultivated planet. *Nature* **478**, 337–342 (2011).
5. Borrelli, P. et al. An assessment of the global impact of 21st century land use change on soil erosion. *Nat. Commun.* **8**, 2013 (2017).
6. Pérez, A. P. & Eugenio, N. R. Status of local soil contamination in Europe (European Commission, 2018).
7. Zhao, F. J., Ma, Y., Zhu, Y. G., Tang, Z. & McGrath, S. P. Soil contamination in China: current status and mitigation strategies. *Environ. Sci. Technol.* **49**, 750–759 (2015).
8. Le Quéré, C. et al. Global carbon budget 2018. *Earth Syst. Sci. Data* **10**, 2141–2194 (2018).
9. Kopittke, P. M., Menzies, N. W., Wang, P., McKenna, B. A. & Lombi, E. Soil and the intensification of agriculture for global food security. *Environ. Int.* **132**, 105078 (2019).
10. Türkdoğan, M. K., Kilicel, F., Kara, K., Tuncer, I. & Uygan, I. Heavy metals in soil, vegetables and fruits in the endemic upper gastrointestinal cancer region of Turkey. *Environ. Toxicol. Pharmacol.* **13**, 175–179 (2003).
11. O'Connor, D., Hou, D., Ok, Y. S. & Lanphear, B. P. The effects of iniquitous lead exposure on health. *Nat. Sustain.* **3**, 77–79 (2020).
**A comprehensive comment on human-health impacts of lead exposure, which deserves global attention to reduce its concentrations in consumer products and soil environment**.
12. Rodríguez Eugenio, N., McLaughlin, M. & Pennock, D. Soil pollution: a hidden reality (FAO, 2018).
13. Tilman, D., Balzer, C., Hill, J. & Befort, B. L. Global food demand and the sustainable intensification of agriculture. *Proc. Natl Acad. Sci. USA* **108**, 20260–20264 (2011).
**Found that the global crop demand of 2050 could be achieved by attaining high yields on existing croplands of underyielding nations**.
14. Ministry of Environmental Protection. China national soil contamination survey report (CWR, 2014).
15. Hou, D. & Li, F. Complexities surrounding China's soil action plan. *Land Degrad. Dev.* **28**, 2315–2320 (2017).
16. Hu, Y., Cheng, H. & Tao, S. The challenges and solutions for cadmium-contaminated rice in China: a critical review. *Environ. Int.* **92–93**, 515–532 (2016).
17. Antoniadis, V. et al. A critical prospective analysis of the potential toxicity of trace element regulation limits in soils worldwide: Are they protective concerning health risk assessment? - a review. *Environ. Int.* **127**, 819–847 (2019).
18. United Nations Environment Assembly of the United Nations Environment Programme. Managing soil pollution to achieve sustainable development (UNEP, 2018).
19. Food and Agriculture Organization of the United Nations. The state of food security and nutrition in the world (FAO, 2019).
20. Järup, L. Hazards of heavy metal contamination. *Br. Med. Bull.* **68**, 167–182 (2003).
**Outlines the serious human-health impacts of heavy metal(loid)s exposure**.
21. British Geological Survey. World mineral statistics data. *BGS* https://www.bgs.ac.uk/mineralsuk/statistics/home.html (2019).
22. Dominish, E., Teske, S. & Florin, N. Responsible minerals sourcing for renewable energy (UTS, 2019).
23. Mulligan, C. N., Yong, R. N. & Gibbs, B. F. Remediation technologies for metal-contaminated soils and groundwater: an evaluation. *Eng. Geol.* **60**, 193–207 (2001).
24. Rogich, D. G. & Matos, G. R. The global flows of metals and minerals (USGS, 2008).
25. Yin, W. J., Yang, J. H., Kang, J., Yan, Y. & Wei, S. H. Halide perovskite materials for solar cells: a theoretical review. *J. Mater. Chem. A* **3**, 8926–8942 (2015).
26. Li, B., Zheng, M., Xue, H. & Pang, H. High performance electrochemical capacitor materials focusing on nickel based materials. *Inorg. Chem. Front.* **3**, 175–202 (2016).
27. Tóth, G., Hermann, T., Da Silva, M. R. & Montanarella, L. Heavy metals in agricultural soils of the European Union with implications for food safety. *Environ. Int.* **88**, 299–309 (2016).
**Provides evidence that only 6.24% of European agricultural lands needs local assessment and remediation action to reduce the heavy metal(loid)s contaminations**.
28. Boussen, S., Soubrand, M., Bril, H., Ouerfelli, K. & Abdeljaouad, S. Transfer of lead, zinc and cadmium from mine tailings to wheat (*Triticum aestivum*) in carbonated Mediterranean (Northern Tunisia) soils. *Geoderma* **192**, 227–236 (2013).
29. Navarro, M. C. et al. Abandoned mine sites as a source of contamination by heavy metals: a case study in a semi-arid zone. *J. Geochem. Explor.* **96**, 183–193 (2008).
30. Atafar, Z. et al. Effect of fertilizer application on soil heavy metal concentration. *Environ. Monit. Assess.* **160**, 83–89 (2010).
31. Srivastava, S. K., Tyagi, R. & Pant, N. Adsorption of heavy metal ions on carbonaceous material developed from the waste slurry generated in local fertilizer plants. *Water Res.* **23**, 1161–1165 (1989).
32. Walter, I., Martínez, F. & Cala, V. Heavy metal speciation and phytotoxic effects of three representative sewage sludges for agricultural uses. *Environ. Pollut.* **139**, 507–514 (2006).
33. Chaoua, S., Boussaa, S., El Gharmali, A. & Boumezzough, A. Impact of irrigation with wastewater on accumulation of heavy metals in soil and crops in the region of Marrakech in Morocco. *J. Saudi Soc. Agric. Sci.* **18**, 429–436 (2019).
34. Balkhair, K. S. & Ashraf, M. A. Field accumulation risks of heavy metals in soil and vegetable crop irrigated with sewage water in western region of Saudi Arabia. *Saudi J. Biol. Sci.* **23**, S32–S44 (2016).
35. Bi, X. et al. Environmental contamination of heavy metals from zinc smelting areas in Hezhang County, western Guizhou, China. *Environ. Int.* **32**, 883–890 (2006).
36. Noli, F. & Tsamos, P. Concentration of heavy metals and trace elements in soils, waters and vegetables and assessment of health risk in the vicinity of a lignite-fired power plant. *Sci. Total Environ.* **563–564**, 377–385 (2016).
37. Zhang, F. et al. Influence of traffic activity on heavy metal concentrations of roadside farmland soil in mountainous areas. *Int. J. Environ. Res. Public Health* **9**, 1715–1731 (2012).
38. Ogunkunle, C. O. & Fatoba, P. O. Pollution loads and the ecological risk assessment of soil heavy metals around a mega cement factory in southwest Nigeria. *Pol. J. Environ. Stud.* **22**, 487–493 (2013).
39. Luo, C. et al. Heavy metal contamination in soils and vegetables near an e-waste processing site, south China. *J. Hazard. Mater.* **186**, 481–490 (2011).
40. Hou, D., O'Connor, D., Nathanail, P., Tian, L. & Ma, Y. Integrated GIS and multivariate statistical analysis for regional scale assessment of heavy metal soil contamination: a critical review. *Environ. Pollut.* **231**, 1188–1200 (2017).
41. Motamen Salehi, F., Khaemba, D. N., Morina, A. & Neville, A. Corrosive–abrasive wear induced by soot in boundary lubrication regime. *Tribol. Lett.* **63**, 19 (2016).
42. Lu, A. et al. Multivariate and geostatistical analyses of the spatial distribution and origin of heavy metals in the agricultural soils in Shunyi, Beijing, China. *Sci. Total Environ.* **425**, 66–74 (2012).
43. Nziguheba, G. & Smolders, E. Inputs of trace elements in agricultural soils via phosphate fertilizers in European countries. *Sci. Total Environ.* **390**, 53–57 (2008).
44. Wang, X. et al. Evolving wastewater infrastructure paradigm to enhance harmony with nature. *Sci. Adv.* **4**, eaaq0210 (2018).
45. Khan, S., Cao, Q., Zheng, Y. M., Huang, Y. Z. & Zhu, Y. G. Health risks of heavy metals in contaminated soils and food crops irrigated with wastewater in Beijing, China. *Environ. Pollut.* **152**, 686–692 (2008).
46. Nicholson, F. A., Smith, S. R., Alloway, B. J., Carlton-Smith, C. & Chambers, B. J. An inventory of heavy metals inputs to agricultural soils in England and Wales. *Sci. Total Environ.* **311**, 205–219 (2003).
47. Clemens, S. Toxic metal accumulation, responses to exposure and mechanisms of tolerance in plants. *Biochimie* **88**, 1707–1719 (2006).
48. Marrugo-Negrete, J., Pinedo-Hernández, J. & Díez, S. Assessment of heavy metal pollution, spatial distribution and origin in agricultural soils along the Sinú River Basin, Colombia. *Environ. Res.* **154**, 380–388 (2017).
49. Micó, C., Recatalá, L., Peris, M. & Sánchez, J. Assessing heavy metal sources in agricultural soils of an European Mediterranean area by multivariate analysis. *Chemosphere* **65**, 863–872 (2006).
50. Facchinelli, A., Sacchi, E. & Mallen, L. Multivariate statistical and GIS-based approach to identify heavy metal sources in soils. *Environ. Pollut.* **114**, 313–324 (2001).
51. Shan, Y. et al. Identification of sources of heavy metals in agricultural soils using multivariate analysis and GIS. *J. Soils Sediments* **13**, 720–729 (2013).
52. Li, X., Lee, S., Wong, S., Shi, W. & Thornton, I. The study of metal contamination in urban soils of Hong Kong using a GIS-based approach. *Environ. Pollut.* **129**, 113–124 (2004).
53. Zhou, J., Feng, K., Pei, Z., Meng, F. & Sun, J. Multivariate analysis combined with GIS to source identification of heavy metals in soils around an abandoned industrial area, Eastern China. *Ecotoxicology* **25**, 380–388 (2016).
54. Barbieri, M., Sappa, G. & Nigro, A. Soil pollution: anthropogenic versus geogenic contributions over large areas of the Lazio region. *J. Geochem. Explor.* **195**, 78–86 (2018).
55. Palumbo, B. et al. Influence of inheritance and pedogenesis on heavy metal distribution in soils of Sicily, Italy. *Geoderma* **95**, 247–266 (2000).
56. Cao, X., Ma, L. Q., Chen, M., Hardison, D. W. & Harris, W. G. Lead transformation and distribution in the soils of shooting ranges in Florida, USA. *Sci. Total Environ.* **307**, 179–189 (2003).
57. Knechtenhofer, L. A., Xifra, I. O., Scheinost, A. C., Flühler, H. & Kretzschmar, R. Fate of heavy metals in a strongly acidic shooting-range soil: small-scale metal distribution and its relation to preferential water flow. *J. Plant Nutr. Soil Sci.* **166**, 84–92 (2003).
58. Fang, W., Wei, Y. & Liu, J. Comparative characterization of sewage sludge compost and soil: heavy metal leaching characteristics. *J. Hazard. Mater.* **310**, 1–10 (2016).
59. Düring, R. A., Hoβ, T. & Gäth, S. Sorption and bioavailability of heavy metals in long-term differently tilled soils amended with organic wastes. *Sci. Total Environ.* **313**, 227–234 (2003).
60. Angelova, V., Ivanova, R., Delibaltova, V. & Ivanov, K. Bio-accumulation and distribution of heavy metals in fibre crops (flax, cotton and hemp). *Ind. Crop. Prod.* **19**, 197–205 (2004).
61. Greinert, A. The heterogeneity of urban soils in the light of their properties. *J. Soils Sediments* **15**, 1725–1737 (2015).

62. Fisher-Power, L. M., Cheng, T. & Rastghalam, Z. S. Cu and Zn adsorption to a heterogeneous natural sediment: Influence of leached cations and natural organic matter. *Chemosphere* **144**, 1973–1979 (2016).
63. Zhang, Y. et al. Lead contamination in Chinese surface soils: source identification, spatial-temporal distribution and associated health risks. *Crit. Rev. Environ. Sci. Technol.* **49**, 1386–1423 (2019).
**The soil lead pollution in China is more severe, with hotspots due to anthropogenic activities**.
64. Dimitrijević, M. D., Nujkić, M. M., Alagić, S., Milić, S. M. & Tošić, S. B. Heavy metal contamination of topsoil and parts of peach-tree growing at different distances from a smelting complex. *Int. J. Environ. Sci. Technol.* **13**, 615–630 (2016).
65. Alsbou, E. M. E. & Al-Khashman, O. A. Heavy metal concentrations in roadside soil and street dust from Petra region, Jordan. *Environ. Monit. Assess.* **190**, 48 (2017).
66. Romic, M. & Romic, D. Heavy metals distribution in agricultural topsoils in urban area. *Environ. Geol.* **43**, 795–805 (2003).
67. Jin, Y. et al. Assessment of sources of heavy metals in soil and dust at children's playgrounds in Beijing using GIS and multivariate statistical analysis. *Environ. Int.* **124**, 320–328 (2019).
68. Li, C., Li, F., Wu, Z. & Cheng, J. Effects of landscape heterogeneity on the elevated trace metal concentrations in agricultural soils at multiple scales in the Pearl River Delta, South China. *Environ. Pollut.* **206**, 264–274 (2015).
69. Rodríguez Martín, J. A., Arias, M. L. & Grau Corbí, J. M. Heavy metals contents in agricultural topsoils in the Ebro basin (Spain). Application of the multivariate geoestatistical methods to study spatial variations. *Environ. Pollut.* **144**, 1001–1012 (2006).
70. Feng, X. & Qiu, G. Mercury pollution in Guizhou, Southwestern China — an overview. *Sci. Total Environ.* **400**, 227–237 (2008).
71. Zhao, H., Xia, B., Fan, C., Zhao, P. & Shen, S. Human health risk from soil heavy metal contamination under different land uses near Dabaoshan Mine, Southern China. *Sci. Total Environ.* **417–418**, 45–54 (2012).
72. Gusev, A. et al. Assessment of transboundary pollution by toxic substances: heavy metals and POPs (EMEP, 2019).
73. Chen, H., Teng, Y., Lu, S., Wang, Y. & Wang, J. Contamination features and health risk of soil heavy metals in China. *Sci. Total Environ.* **512–513**, 143–153 (2015).
74. Li, W. et al. The identification of 'hotspots' of heavy metal pollution in soil–rice systems at a regional scale in eastern China. *Sci. Total Environ.* **472**, 407–420 (2014).
75. Wang, X. et al. Climate and vegetation as primary drivers for global mercury storage in surface soil. *Environ. Sci. Technol.* **53**, 10665–10675 (2019).
76. Caporale, A. G. & Violante, A. Chemical processes affecting the mobility of heavy metals and metalloids in soil environments. *Curr. Pollut. Rep.* **2**, 15–27 (2016).
77. Antoniadis, V., Golia, E. E., Shaheen, S. M. & Rinklebe, J. Bioavailability and health risk assessment of potentially toxic elements in Thriasio Plain, near Athens, Greece. *Environ. Geochem. Health* **39**, 319–330 (2017).
78. O'Connor, D. et al. Mercury speciation, transformation, and transportation in soils, atmospheric flux, and implications for risk management: a critical review. *Environ. Int.* **126**, 747–761 (2019).
79. Kim, R. Y. et al. Bioavailability of heavy metals in soils: definitions and practical implementation — a critical review. *Environ. Geochem. Health* **37**, 1041–1061 (2015).
80. Okoro, H. K., Fatoki, O. S., Adekola, F. A., Ximba, B. J. & Snyman, R. G. A review of sequential extraction procedures for heavy metals speciation in soil and sediments. *Open Access. Sci. Rep.* **1**, 1–9 (2012).
81. Hettiarachchi, G. M. & Pierzynski, G. M. Soil lead bioavailability and in situ remediation of lead-contaminated soils: a review. *Environ. Prog.* **23**, 78–93 (2004).
82. Shahid, M., Dumat, C., Khalid, S., Niazi, N. K. & Antunes, P. M. C. in *Reviews of Environmental Contamination and Toxicology* (eds Gunther, F. A. & de Voogt, P.) 73–137 (Springer, 2016).
83. Vromman, D., Martínez, J. P., Kumar, M., Šlejkovec, Z. & Lutts, S. Comparative effects of arsenite (As(III)) and arsenate (As(V)) on whole plants and cell lines of the arsenic-resistant halophyte plant species *Atriplex atacamensis*. *Environ. Sci. Pollut. Res.* **25**, 34473–34486 (2018).
84. Rieuwerts, J. S., Thornton, I., Farago, M. E. & Ashmore, M. R. Factors influencing metal bioavailability in soils: preliminary investigations for the development of a critical loads approach for metals. *Chem. Speciat. Bioavailab.* **10**, 61–75 (1998).
85. Antoniadis, V. et al. Trace elements in the soil–plant interface: phytoavailability, translocation, and phytoremediation–a review. *Earth-Sci. Rev.* **171**, 621–645 (2017).
86. Wang, X. & Tang, C. The role of rhizosphere pH in regulating the rhizosphere priming effect and implications for the availability of soil-derived nitrogen to plants. *Ann. Bot.* **121**, 143–151 (2018).
87. Tao, H., Pan, W. L., Carter, P. & Wang, K. Addition of lignin to lime materials for expedited pH increase and improved vertical mobility of lime in no-till soils. *Soil Use Manag.* **35**, 314–322 (2019).
88. Jing, F. et al. Biochar effects on soil chemical properties and mobilization of cadmium (Cd) and lead (Pb) in paddy soil. *Soil Use Manag.* **36**, 320–327 (2020).
89. Aijun, Y., Changle, Q., Shusen, M. & Reardon, E. J. Effects of humus on the environmental activity of mineral-bound Hg: Influence on Hg volatility. *Appl. Geochem.* **21**, 446–454 (2006).
90. Beckers, F. & Rinklebe, J. Cycling of mercury in the environment: Sources, fate, and human health implications: a review. *Crit. Rev. Environ. Sci. Technol.* **47**, 693–794 (2017).
91. Wallschläger, D., Desai, M. V. M., Spengler, M. & Wilken, R.-D. Mercury speciation in floodplain soils and sediments along a contaminated river transect. *J. Environ. Qual.* **27**, 1034 (1998).
92. Shaheen, S. M. & Rinklebe, J. Geochemical fractions of chromium, copper, and zinc and their vertical distribution in floodplain soil profiles along the Central Elbe River, Germany. *Geoderma* **228–229**, 142–159 (2014).
93. Park, J. H. et al. Role of organic amendments on enhanced bioremediation of heavy metal(loid) contaminated soils. *J. Hazard. Mater.* **185**, 549–574 (2011).
94. Chibuike, G. U. & Obiora, S. C. Heavy metal polluted soils: effect on plants and bioremediation methods. *Appl. Environ. Soil Sci.* **2014**, 752708 (2014).
95. Congeevaram, S., Dhanarani, S., Park, J., Dexilin, M. & Thamaraiselvi, K. Biosorption of chromium and nickel by heavy metal resistant fungal and bacterial isolates. *J. Hazard. Mater.* **146**, 270–277 (2007).
96. Dary, M., Chamber-Pérez, M. A., Palomares, A. J. & Pajuelo, E. "In situ" phytostabilisation of heavy metal polluted soils using *Lupinus luteus* inoculated with metal resistant plant-growth promoting rhizobacteria. *J. Hazard. Mater.* **177**, 323–330 (2010).
97. Sarma, H. Metal hyperaccumulation in plants: a review focusing on phytoremediation technology. *J. Environ. Sci. Technol.* **4**, 118–138 (2011).
98. Hou, D. et al. A sustainability assessment framework for agricultural land remediation in China. *Land Degrad. Dev.* **29**, 1005–1018 (2018).
99. Hou, D. *Sustainable Remediation of Contaminated Soil and Groundwater: Materials, Processes, and Assessment* (Butterworth-Heinemann, 2019).
100. Song, Y. et al. Nature based solutions for contaminated land remediation and brownfield redevelopment in cities: a review. *Sci. Total Environ.* **663**, 568–579 (2019).
101. Hou, D. & Al-Tabbaa, A. Sustainability: a new imperative in contaminated land remediation. *Environ. Sci. Policy* **39**, 25–34 (2014).
102. Visoottiviseth, P., Francesconi, K. & Sridokchan, W. The potential of Thai indigenous plant species for the phytoremediation of arsenic contaminated land. *Environ. Pollut.* **118**, 453–461 (2002).
103. Zhao, F. J., Lombi, E. & McGrath, S. P. Assessing the potential for zinc and cadmium phytoremediation with the hyperaccumulator *Thlaspi caerulescens*. *Plant Soil* **249**, 37–43 (2003).
104. Angassa, K., Leta, S., Mulat, W., Kloos, H. & Meers, E. Evaluation of pilot-scale constructed wetlands with *Phragmites karka* for phytoremediation of municipal wastewater and biomass production in Ethiopia. *Environ. Process.* **6**, 65–84 (2019).
105. Wang, L. et al. Field trials of phytomining and phytoremediation: A critical review of influencing factors and effects of additives. *Crit. Rev. Environ. Sci. Technol.* https://doi.org/10.1080/10643389.2019.1705724 (2019).
**Suggests that phytomining and phytoremediation are nature-based solutions for soil pollution, but there are several limitations that have to be addressed**.
106. Keller, C., Ludwig, C., Davoli, F. & Wochele, J. Thermal treatment of metal-enriched biomass produced from heavy metal phytoextraction. *Environ. Sci. Technol.* **39**, 3359–3367 (2005).
**Found that the biochar production is a sound technique to increase recovery of Cd and Zn from plant biomass**.
107. Chalot, M., Blaudez, D., Rogaume, Y., Provent, A.-S. & Pascual, C. Fate of trace elements during the combustion of phytoremediation wood. *Environ. Sci. Technol.* **46**, 13361–13369 (2012).
108. Schreurs, E., Voets, T. & Thewys, T. GIS-based assessment of the biomass potential from phytoremediation of contaminated agricultural land in the Campine region in Belgium. *Biomass Bioenergy* **35**, 4469–4480 (2011).
109. Schröder, P. et al. Intensify production, transform biomass to energy and novel goods and protect soils in Europe — a vision how to mobilize marginal lands. *Sci. Total Environ.* **616–617**, 1101–1123 (2018).
110. Andersson-Sköld, Y., Hagelqvist, A., Crutu, G. & Blom, S. Bioenergy grown on contaminated land – a sustainable bioenergy contributor? *Biofuels* **5**, 487–498 (2014).
111. Huang, H. et al. Effect of pyrolysis temperature on chemical form, behavior and environmental risk of Zn, Pb and Cd in biochar produced from phytoremediation residue. *Bioresour. Technol.* **249**, 487–493 (2018).
112. Murtaza, G., Javed, W., Hussain, A., Qadir, M. & Aslam, M. Soil-applied zinc and copper suppress cadmium uptake and improve the performance of cereals and legumes. *Int. J. Phytoremediat.* **19**, 199–206 (2017).
113. Ahmad, P. et al. Alleviation of cadmium toxicity in *Brassica juncea* L. (Czern. & Coss.) by calcium application involves various physiological and biochemical strategies. *PLoS One* **10**, e0114571 (2015).
114. Meharg, A. A. & Macnair, M. R. Suppression of the high affinity phosphate uptake system: a mechanism of arsenate tolerance in *Holcus lanatus* L. *J. Exp. Bot.* **43**, 519–524 (1992).
115. Shahid, M. et al. Foliar heavy metal uptake, toxicity and detoxification in plants: a comparison of foliar and root metal uptake. *J. Hazard. Mater.* **325**, 36–58 (2017).
116. Che-Castaldo, J. P. & Inouye, D. W. Interspecific competition between a non-native metal-hyperaccumulating plant (*Noccaea caerulescens*, Brassicaceae) and a native congener across a soil-metal gradient. *Aust. J. Bot.* **63**, 141 (2015).
117. Moreno, F. N., Anderson, C. W. N., Stewart, R. B. & Robinson, B. H. Phytoremediation of mercury-contaminated mine tailings by induced plant-mercury accumulation. *Environ. Pract.* **6**, 165–175 (2004).
118. Zhang, L., Rylott, E. L., Bruce, N. C. & Strand, S. E. Genetic modification of western wheatgrass (*Pascopyrum smithii*) for the phytoremediation of RDX and TNT. *Planta* **249**, 1007–1015 (2019).
119. Guidi Nissim, W. & Labrecque, M. Planting microcuttings: an innovative method for establishing a willow vegetation cover. *Ecol. Eng.* **91**, 472–476 (2016).
120. Bhuiyan, M. S. I., Raman, A. & Hodgkins, D. S. Plants in remediating salinity-affected agricultural landscapes. *Proc. Indian Natl Sci. Acad.* **83**, 51–66 (2017).
121. Cohen, C. K., Fox, T. C., Garvin, D. F. & Kochian, L. V. The role of iron-deficiency stress responses in stimulating heavy-metal transport in plants. *Plant Physiol.* **116**, 1063–72 (1998).
**Provides evidence that plant accumulation of heavy metal(loid)s can be improved under iron deficiency in soil**.
122. Luo, J., He, W., Xing, X., Wu, J. & Sophie Gu, X. W. The variation of metal fractions and potential environmental risk in phytoremediating multiple metal polluted soils using *Noccaea caerulescens* assisted by LED lights. *Chemosphere* **227**, 462–469 (2019).

123. Fu, Y. et al. Interaction effects of light intensity and nitrogen concentration on growth, photosynthetic characteristics and quality of lettuce (*Lactuca sativa* L. Var. youmaicai). *Sci. Hortic.* **214**, 51–57 (2017).
124. Niazi, N. K. et al. Phosphate-assisted phytoremediation of arsenic by *Brassica napus* and *Brassica juncea*: Morphological and physiological response. *Int. J. Phytoremediat.* **19**, 670–678 (2017).
125. Iqbal, M., Puschenreiter, M., Oburger, E., Santner, J. & Wenzel, W. W. Sulfur-aided phytoextraction of Cd and Zn by *Salix smithiana* combined with in situ metal immobilization by gravel sludge and red mud. *Environ. Pollut.* **170**, 222–231 (2012).
126. Cameselle, C. & Gouveia, S. Phytoremediation of mixed contaminated soil enhanced with electric current. *J. Hazard. Mater.* **361**, 95–102 (2019).
127. Rehman, K., Imran, A., Amin, I. & Afzal, M. Inoculation with bacteria in floating treatment wetlands positively modulates the phytoremediation of oil field wastewater. *J. Hazard. Mater.* **349**, 242–251 (2018).
128. Wang, X. et al. Transgenic tobacco plants expressing a P1B-ATPase gene from *Populus tomentosa* Carr. (*PtoHMA5*) demonstrate improved cadmium transport. *Int. J. Biol. Macromol.* **113**, 655–661 (2018).
129. Ma, L. Q. et al. A fern that hyperaccumulates arsenic. *Nature* **409**, 579 (2001).
**Found that fern *Pteris vittata* (brake fern) is highly efficient in phytoremediation of arsenic in contaminated soils**.
130. Rascio, N. & Navari-Izzo, F. Heavy metal hyperaccumulating plants: How and why do they do it? And what makes them so interesting? *Plant Sci.* **180**, 169–181 (2011).
131. Li, Z., Wu, L., Luo, Y. & Christie, P. Changes in metal mobility assessed by EDTA kinetic extraction in three polluted soils after repeated phytoremediation using a cadmium/zinc hyperaccumulator. *Chemosphere* **194**, 432–440 (2018).
132. Tőzsér, D. et al. Phytoextraction with *Salix viminalis* in a moderately to strongly contaminated area. *Environ. Sci. Pollut. Res.* **25**, 3275–3290 (2018).
133. Chandra, R. & Kang, H. Mixed heavy metal stress on photosynthesis, transpiration rate, and chlorophyll content in poplar hybrids. *For. Sci. Technol.* **12**, 55–61 (2016).
134. Luo, J., Qi, S., Gu, X. W. S., Wang, J. & Xie, X. Evaluation of the phytoremediation effect and environmental risk in remediation processes under different cultivation systems. *J. Clean. Prod.* **119**, 25–31 (2016).
135. Hu, R. et al. Intercropping with hyperaccumulator plants decreases the cadmium accumulation in grape seedlings. *Acta Agric. Scand. B Soil Plant Sci.* **69**, 304–310 (2019).
136. Praveen, A., Mehrotra, S. & Singh, N. Mixed plantation of wheat and accumulators in arsenic contaminated plots: a novel way to reduce the uptake of arsenic in wheat and load on antioxidative defence of plant. *Ecotoxicol. Environ. Saf.* **182**, 109462 (2019).
137. Ewel, J. J., Schreeg, L. A. & Sinclair, T. R. Resources for crop production: accessing the unavailable. *Trends Plant Sci.* **24**, 121–129 (2019).
138. Wan, X., Lei, M., Chen, T. & Yang, J. Intercropped *Pteris vittata* L. and *Morus alba* L. presents a safe utilization mode for arsenic-contaminated soil. *Sci. Total Environ.* **579**, 1467–1475 (2017).
139. Smith, E., Juhasz, A. L., Weber, J. & Naidu, R. Arsenic uptake and speciation in rice plants grown under greenhouse conditions with arsenic contaminated irrigation water. *Sci. Total Environ.* **392**, 277–283 (2008).
140. ur Rehman, M. Z. et al. Remediation of heavy metal contaminated soils by using *Solanum nigrum*: a review. *Ecotoxicol. Environ. Saf.* **143**, 236–248 (2017).
141. Cheng, S. F., Huang, C. Y., Lin, Y. C., Lin, S. C. & Chen, K. L. Phytoremediation of lead using corn in contaminated agricultural land-An in situ study and benefit assessment. *Ecotoxicol. Environ. Saf.* **111**, 72–77 (2015).
142. Jeong, S., Moon, H. S. & Nam, K. Increased ecological risk due to the hyperaccumulation of As in *Pteris cretica* during the phytoremediation of an As-contaminated site. *Chemosphere* **122**, 1–7 (2015).
143. Li, Z., Jia, M., Wu, L., Christie, P. & Luo, Y. Changes in metal availability, desorption kinetics and speciation in contaminated soils during repeated phytoextraction with the Zn/Cd hyperaccumulator *Sedum plumbizincicola*. *Environ. Pollut.* **209**, 123–131 (2016).
144. Jiang, J. et al. Effects of multiple heavy metal contamination and repeated phytoextraction by *Sedum plumbizincicola* on soil microbial properties. *Eur. J. Soil Biol.* **46**, 18–26 (2010).
145. Hou, X. et al. Pb stress effects on leaf chlorophyll fluorescence, antioxidative enzyme activities, and organic acid contents of *Pogonatherum crinitum* seedlings. *Flora* **240**, 82–88 (2018).
146. Shen, Z., Wang, Y., Chen, Y. & Zhang, Z. Transfer of heavy metals from the polluted rhizosphere soil to *Celosia argentea* L. in copper mine tailings. *Hortic. Environ. Biotechnol.* **58**, 93–100 (2017).
147. Pratas, J., Favas, P. J. C., Varun, M., D'Souza, R. & Paul, M. S. Distribution of rare earth elements, thorium and uranium in streams and aquatic mosses of Central Portugal. *Environ. Earth Sci.* **76**, 156 (2017).
148. Bennett, L. E. et al. Analysis of transgenic Indian mustard plants for phytoremediation of metal-contaminated mine tailings. *J. Environ. Qual.* **32**, 432–440 (2003).
149. Van Huysen, T. et al. Overexpression of cystathionine-γ-synthase enhances selenium volatilization in *Brassica juncea*. *Planta* **218**, 71–78 (2003).
150. Davison, J. Risk mitigation of genetically modified bacteria and plants designed for bioremediation. *J. Ind. Microbiol. Biotechnol.* **32**, 639–650 (2005).
151. Al-Ahmad, H., Galili, S. & Gressel, J. Tandem constructs to mitigate transgene persistence: tobacco as a model. *Mol. Ecol.* **13**, 697–710 (2004).
152. Shrestha, P., Bellitürk, K. & Görres, J. H. Phytoremediation of heavy metal-contaminated soil by switchgrass: a comparative study utilizing different composts and coir fiber on pollution remediation, plant productivity, and nutrient leaching. *Int. J. Environ. Res. Public Health* **16**, 1261 (2019).
153. Din, B. U. et al. Assisted phytoremediation of chromium spiked soils by *Sesbania sesban* in association with *Bacillus xiamenensis* PM14: a biochemical analysis. *Plant Physiol. Biochem.* **146**, 249–258 (2020).
154. Zhang, X. et al. Effect of plant-growth-promoting rhizobacteria on phytoremediation efficiency of *Scirpus triqueter* in pyrene-Ni co-contaminated soils. *Chemosphere* **241**, 125027 (2020).
155. Willscher, S. et al. Field scale phytoremediation experiments on a heavy metal and uranium contaminated site, and further utilization of the plant residues. *Hydrometallurgy* **131–132**, 46–53 (2013).
156. Baker, A. J. M., McGrath, S. P., Sidoli, C. M. D. & Reeves, R. D. The possibility of in situ heavy metal decontamination of polluted soils using crops of metal-accumulating plants. *Resour. Conserv. Recycl.* **11**, 41–49 (1994).
157. Kidd, P. et al. Agronomic practices for improving gentle remediation of trace element-contaminated soils. *Int. J. Phytoremediat.* **17**, 1005–1037 (2015).
158. Ji, P., Sun, T., Song, Y., Ackland, M. L. & Liu, Y. Strategies for enhancing the phytoremediation of cadmium-contaminated agricultural soils by *Solanum nigrum* L. *Environ. Pollut.* **159**, 762–768 (2011).
159. Li, N. et al. Effects of double harvesting on heavy metal uptake by six forage species and the potential for phytoextraction in field. *Pedosphere* **26**, 717–724 (2016).
160. Lim, J. E. et al. Impact of natural and calcined starfish (*Asterina pectinifera*) on the stabilization of Pb, Zn and As in contaminated agricultural soil. *Environ. Geochem. Health* **39**, 431–441 (2017).
161. Mertens, J., Vervaeke, P., Meers, E. & Tack, F. M. G. Seasonal changes of metals in willow (*Salix* sp.) stands for phytoremediation on dredged sediment. *Environ. Sci. Technol.* **40**, 1962–1968 (2006).
162. Sampanpanish, P. & Nanthavong, K. Effect of EDTA and NTA on arsenic bioaccumulation and translocation using phytoremediation by *Mimosa pudica* L. from contaminated soils. *Bull. Environ. Contam. Toxicol.* **102**, 140–145 (2019).
163. Wan, X., Lei, M. & Chen, T. Cost–benefit calculation of phytoremediation technology for heavy-metal-contaminated soil. *Sci. Total Environ.* **563–564**, 796–802 (2016).
**Illustrates that the phytoremediation renders low environmental impacts and costs compared with other remediation technologies**.
164. Ruttens, A. et al. Short rotation coppice culture of willows and poplars as energy crops on metal contaminated agricultural soils. *Int. J. Phytoremediat.* **13**, 194–207 (2011).
**Suggests phytoremediation with tolerant plant species for adverse environmental conditions is a promising remediation method for mine tailings**.
165. Touceda-González, M. et al. Aided phytostabilisation reduces metal toxicity, improves soil fertility and enhances microbial activity in Cu-rich mine tailings. *J. Environ. Manage.* **186**, 301–313 (2017).
166. Grobelak, A. et al. Effects of single sewage sludge application on soil phytoremediation. *J. Clean. Prod.* **155**, 189–197 (2017).
167. Galende, M. A. et al. Field assessment of the effectiveness of organic amendments for aided phytostabilization of a Pb-Zn contaminated mine soil. *J. Geochem. Explor.* **145**, 181–189 (2014).
168. Panda, D., Panda, D., Padhan, B. & Biswas, M. Growth and physiological response of lemongrass (*Cymbopogon citratus* (D.C.) Stapf.) under different levels of fly ash-amended soil. *Int. J. Phytoremediat.* **20**, 538–544 (2018).
169. Kursun Unver, I. & Terzi, M. Distribution of trace elements in coal and coal fly ash and their recovery with mineral processing practices: a review. *J. Min. Environ.* **9**, 641–655 (2018).
170. Blissett, R. S. & Rowson, N. A. A review of the multi-component utilisation of coal fly ash. *Fuel* **97**, 1–23 (2012).
171. Li, X., Wang, X., Chen, Y., Yang, X. & Cui, Z. Optimization of combined phytoremediation for heavy metal contaminated mine tailings by a field-scale orthogonal experiment. *Ecotoxicol. Environ. Saf.* **168**, 1–8 (2019).
172. Khaokaew, S. & Landrot, G. A field-scale study of cadmium phytoremediation in a contaminated agricultural soil at Mae Sot district, Tak Province, Thailand: (1) Determination of Cd-hyperaccumulating plants. *Chemosphere* **138**, 883–887 (2015).
173. Chen, Y., Shen, Z. & Li, X. The use of vetiver grass (*Vetiveria zizanioides*) in the phytoremediation of soils contaminated with heavy metals. *Appl. Geochem.* **19**, 1553–1565 (2004).
174. Wang, L., Ji, B., Hu, Y., Liu, R. & Sun, W. A review on in situ phytoremediation of mine tailings. *Chemosphere* **184**, 594–600 (2017).
175. Vangronsveld, J. et al. Phytoremediation of contaminated soils and groundwater: lessons from the field. *Environ. Sci. Pollut. Res.* **16**, 765–794 (2009).
176. Kallmeyer, J., Pockalny, R., Adhikari, R. R., Smith, D. C. & D'Hondt, S. Global distribution of microbial abundance and biomass in subseafloor sediment. *Proc. Natl Acad. Sci. USA* **109**, 16213–16216 (2012).
177. Anantharaman, K. et al. Thousands of microbial genomes shed light on interconnected biogeochemical processes in an aquifer system. *Nat. Commun.* **7**, 13219 (2016).
**Provides evidence that inter-organism interactions of soil microorganisms are significant in biogeochemical cycling of carbon, nitrogen, sulfur and hydrogen**.
178. Serna-Chavez, H. M., Fierer, N. & Van Bodegom, P. M. Global drivers and patterns of microbial abundance in soil. *Glob. Ecol. Biogeogr.* **22**, 1162–1172 (2013).
**Illustrates that soil microbial biomass is not primarily driven by soil temperature but by soil moisture and nutrients availability**.
179. Madsen, E. L. Microorganisms and their roles in fundamental biogeochemical cycles. *Curr. Opin. Biotechnol.* **22**, 456–464 (2011).
**A comprehensive view of the microbially mediated biogeochemical processes that transform and recycle organic and inorganic substances in soils, sediments and waters**.
180. Lovley, D. R. Cleaning up with genomics: applying molecular biology to bioremediation. *Nat. Rev. Microbiol.* **1**, 35–44 (2003).
**A comprehensive view of the application of genome-enabled techniques for bioremediation**.
181. Gadd, G. M. Bioremedial potential of microbial mechanisms of metal mobilization and immobilization. *Curr. Opin. Biotechnol.* **11**, 271–279 (2000).
**The environmental fate of toxic metals and radionuclides is determined by diverse mechanisms of microorganisms**.
182. Rajendran, P., Muthukrishnan, J. & Gunasekaran, P. Microbes in heavy metal remediation. *Indian. J. Exp. Biol.* **41**, 935–944 (2003).
183. Pushpanathan, M., Jayashree, S., Gunasekaran, P. & Rajendhran, J. in *Microbial Biodegradation and Bioremediation* (ed. Das, S.) 407–419 (Elsevier, 2014).

184. Kavamura, V. N. & Esposito, E. Biotechnological strategies applied to the decontamination of soils polluted with heavy metals. *Biotechnol. Adv.* **28**, 61–69 (2010).
185. Gadd, G. M. Microbial influence on metal mobility and application for bioremediation. *Geoderma* **122**, 109–119 (2004).
186. Gadd, G. M. Metals, minerals and microbes: geomicrobiology and bioremediation. *Microbiology* **156**, 609–643 (2010).
**Microbial transformations of metals and minerals are vital biosphere processes, with both beneficial and detrimental impacts on human society**.
187. Hesse, E. et al. Anthropogenic remediation of heavy metals selects against natural microbial remediation. *Proc. R. Soc. B Biol. Sci.* **286**, 20190804 (2019).
188. Luján, A. M., Gómez, P. & Buckling, A. Siderophore cooperation of the bacterium *Pseudomonas fluorescens* in soil. *Biol. Lett.* **11**, 20140934 (2015).
189. Hider, R. C. & Kong, X. Chemistry and biology of siderophores. *Nat. Prod. Rep.* **27**, 637 (2010).
190. Hesse, E. et al. Ecological selection of siderophore-producing microbial taxa in response to heavy metal contamination. *Ecol. Lett.* **21**, 117–127 (2018).
191. O'Brien, S. & Buckling, A. The sociality of bioremediation: Hijacking the social lives of microbial populations to clean up heavy metal contamination. *EMBO Rep.* **16**, 1241–5 (2015).
192. Diels, L., De Smet, M., Hooyberghs, L. & Corbisier, P. Heavy metals bioremediation of soil. *Mol. Biotechnol.* **12**, 149–158 (1999).
193. Shenker, M. & Chen, Y. Increasing iron availability to crops: fertilizers, organo-fertilizers, and biological approaches. *Soil Sci. Plant Nutr.* **51**, 1–17 (2005).
194. Blowes, D. W., Ptacek, C. J., Jambor, J. L. & Weisener, C. G. in *Treatise on Geochemistry* (eds Holland, H. D. & Turekian, K. K.) 149–204 (Elsevier, 2003).
195. White, C., Shaman, A. K. & Gadd, G. M. An integrated microbial process for the bioremediation of soil contaminated with toxic metals. *Nat. Biotechnol.* **16**, 572–575 (1998).
196. Chang, C.-Y., Chen, S.-Y., Klipkhayai, P. & Chiemchaisri, C. Bioleaching of heavy metals from harbor sediment using sulfur-oxidizing microflora acclimated from native sediment and exogenous soil. *Environ. Sci. Pollut. Res.* **26**, 6818–6828 (2019).
197. Rasoulnia, P., Mousavi, S. M., Rastegar, S. O. & Azargoshasb, H. Fungal leaching of valuable metals from a power plant residual ash using *Penicillium simplicissimum*: Evaluation of thermal pretreatment and different bioleaching methods. *Waste Manag.* **52**, 309–317 (2016).
198. Ren, W.-X., Li, P.-J., Geng, Y. & Li, X.-J. Biological leaching of heavy metals from a contaminated soil by *Aspergillus niger*. *J. Hazard. Mater.* **167**, 164–169 (2009).
199. Jadhav, U., Su, C. & Hocheng, H. Leaching of metals from printed circuit board powder by an *Aspergillus niger* culture supernatant and hydrogen peroxide. *RSC Adv.* **6**, 43442–43452 (2016).
200. Liang, X. & Gadd, G. M. Metal and metalloid biorecovery using fungi. *Microb. Biotechnol.* **10**, 1199–1205 (2017).
201. Deng, X. et al. Bioleaching of heavy metals from a contaminated soil using indigenous *Penicillium chrysogenum* strain F1. *J. Hazard. Mater.* **233–234**, 25–32 (2012).
202. Deng, X., Yang, Z. & Chen, R. Study of characteristics on metabolism of *Penicillium chrysogenum* F1 during bioleaching of heavy metals from contaminated soil. *Can. J. Microbiol.* **65**, 629–641 (2019).
203. Zayed, A. M. & Terry, N. Chromium in the environment: Factors affecting biological remediation. *Plant Soil* **249**, 139–156 (2003).
204. Alessi, D. S. et al. Speciation and reactivity of uranium products formed during in situ bioremediation in a shallow alluvial aquifer. *Environ. Sci. Technol.* **48**, 12842–12850 (2014).
205. Bargar, J. R. et al. Uranium redox transition pathways in acetate-amended sediments. *Proc. Natl Acad. Sci. USA* **110**, 4506–4511 (2013).
206. Damian, G. E., Micle, V., Sur, I. M. & Chirilă Băbău, A. M. From environmental ethics to sustainable decision-making: assessment of potential ecological risk in soils around abandoned mining areas-case study "Larga de Sus mine" (Romania). *J. Agric. Environ. Ethics* **32**, 27–49 (2019).
207. Xie, Z. et al. Conservation opportunities on uncontested lands. *Nat. Sustain.* **3**, 9–15 (2020).
208. Favas, P. J. C., Pratas, J., Paul, M. S. & Prasad, M. N. V. in *Phytomanagement of Polluted Sites* (eds Pandey, V. C. & Bauddh, K.) 277–300 (Elsevier, 2019).
209. Gadd, G. M. Heavy metal accumulation by bacteria and other microorganisms. *Experientia* **46**, 834–840 (1990).
210. Kasemodel, M. C., Sakamoto, I. K., Varesche, M. B. A. & Rodrigues, V. G. S. Potentially toxic metal contamination and microbial community analysis in an abandoned Pb and Zn mining waste deposit. *Sci. Total Environ.* **675**, 367–379 (2019).
211. Nies, D. H. Heavy metal-resistant bacteria as extremophiles: Molecular physiology and biotechnological use of *Ralstonia* sp. CH34. *Extremophiles* **4**, 77–82 (2000).
212. Valls, M. & de Lorenzo, V. Exploiting the genetic and biochemical capacities of bacteria for the remediation of heavy metal pollution. *FEMS Microbiol. Rev.* **26**, 327–338 (2002).
213. Liang, Y. et al. Molecular characteristics, proton dissociation properties, and metal binding properties of soil organic matter: A theoretical study. *Sci. Total Environ.* **656**, 521–530 (2019).
214. Elghali, A. et al. The role of hardpan formation on the reactivity of sulfidic mine tailings: a case study at Joutel mine (Québec). *Sci. Total Environ.* **654**, 118–128 (2019).
215. Achal, V., Pan, X. & Zhang, D. Remediation of copper-contaminated soil by *Kocuria flava* CR1, based on microbially induced calcite precipitation. *Ecol. Eng.* **37**, 1601–1605 (2011).
216. Wang, J. et al. Iron–manganese (oxyhydro)oxides, rather than oxidation of sulfides, determine mobilization of Cd during soil drainage in paddy soil systems. *Environ. Sci. Technol.* **53**, 2500–2508 (2019).
217. Chen, Y.-W., Yu, X. & Belzile, N. Arsenic speciation in surface waters and lake sediments in an abandoned mine site and field observations of arsenic eco-toxicity. *J. Geochem. Explor.* **205**, 106349 (2019).
218. Wilkin, R. T. Contaminant attenuation processes at mine sites. *Mine Water Environ.* **27**, 251–258 (2008).
219. Kumar, V. et al. Impact of tillage and crop establishment methods on crop yields, profitability and soil physical properties in rice–wheat system of Indo-Gangetic Plains of India. *Soil Use Manag.* **35**, 303–313 (2019).
220. Farhate, C. V. V. et al. Soil tillage and cover crop on soil $CO_2$ emissions from sugarcane fields. *Soil Use Manag.* **35**, 273–282 (2019).
221. Mangalassery, S., Mooney, S. J., Sparkes, D. L., Fraser, W. T. & Sjögersten, S. Impacts of zero tillage on soil enzyme activities, microbial characteristics and organic matter functional chemistry in temperate soils. *Eur. J. Soil Biol.* **68**, 9–17 (2015).
222. Lovley, D. R. & Coates, J. D. Bioremediation of metal contamination. *Curr. Opin. Biotechnol.* **8**, 285–289 (1997).
223. Thornton, S. F., Nicholls, H. C. G., Rolfe, S. A., Mallinson, H. E. H. & Spence, M. J. Biodegradation and fate of ethyl *tert*-butyl ether (ETBE) in soil and groundwater: a review. *J. Hazard. Mater.* **391**, 122046 (2020).
224. Ławniczak, Ł., Woźniak-Karczewska, M., Loibner, A. P., Heipieper, H. J. & Chrzanowski, Ł. Microbial degradation of hydrocarbons—basic principles for bioremediation: a review. *Molecules* **25**, 856 (2020).
225. Newsome, L., Morris, K. & Lloyd, J. R. The biogeochemistry and bioremediation of uranium and other priority radionuclides. *Chem. Geol.* **363**, 164–184 (2014).
226. Donati, E. R., Sani, R. K., Goh, K. M. & Chan, K.-G. Editorial: recent advances in bioremediation/biodegradation by extreme microorganisms. *Front. Microbiol.* **10**, 1851 (2019).
227. Lebeau, T. in *Bioaugmentation, Biostimulation and Biocontrol. Soil Biology* Vol. 108 (eds Singh, A., Parmar, N. & Kuhad, R.) 129–186 (Springer, 2011).
228. Young, C.-C., Rekha, P. D., Lai, W.-A. & Arun, A. B. Encapsulation of plant growth-promoting bacteria in alginate beads enriched with humic acid. *Biotechnol. Bioeng.* **95**, 76–83 (2006).
229. Bashan, Y., Hernandez, J. P., Leyva, L. A. & Bacilio, M. Alginate microbeads as inoculant carriers for plant growth-promoting bacteria. *Biol. Fertil. Soils* **35**, 359–368 (2002).
230. van Elsas, J. D., Trevors, J. T., Rosado, A. S. & Nannipieri, P. *Modern Soil Microbiology* 3rd edn (CRC, 2019).
231. Diep, P., Mahadevan, R. & Yakunin, A. F. Heavy metal removal by bioaccumulation using genetically engineered microorganisms. *Front. Bioeng. Biotechnol.* **6**, 157 (2018).
232. Kang, S. H. et al. Bacteria metabolically engineered for enhanced phytochelatin production and cadmium accumulation. *Appl. Environ. Microbiol.* **73**, 6317–6320 (2007).
233. Yadav, S. K. Heavy metals toxicity in plants: an overview on the role of glutathione and phytochelatins in heavy metal stress tolerance of plants. *South Afr. J. Bot.* **76**, 167–179 (2010).
234. Sneller, F. E. C. et al. Derivatization of phytochelatins from *Silene vulgaris*, induced upon exposure to arsenate and cadmium: comparison of derivatization with Ellman's reagent and monobromobimane. *J. Agric. Food Chem.* **48**, 4014–4019 (2000).
235. Gupta, S. & Singh, D. in *Advances in Environmental Biotechnology* (eds Kumar, R., Sharma, A. & Ahluwalia, S.) 197–214 (Springer, 2017).
236. Yergeau, E., Sanschagrin, S., Maynard, C., St-Arnaud, M. & Greer, C. W. Microbial expression profiles in the rhizosphere of willows depend on soil contamination. *ISME J.* **8**, 344–358 (2014).
237. Rajkumar, M., Sandhya, S., Prasad, M. N. V. & Freitas, H. Perspectives of plant-associated microbes in heavy metal phytoremediation. *Biotechnol. Adv.* **30**, 1562–1574 (2012).
238. Braud, A., Jézéquel, K., Bazot, S. & Lebeau, T. Enhanced phytoextraction of an agricultural Cr- and Pb-contaminated soil by bioaugmentation with siderophore-producing bacteria. *Chemosphere* **74**, 280–286 (2009).
239. Plewniak, F., Crognale, S., Rossetti, S. & Bertin, P. N. A genomic outlook on bioremediation: the case of arsenic removal. *Front. Microbiol.* **9**, 820 (2018).
240. Mesa, J. et al. Moving closer towards restoration of contaminated estuaries: bioaugmentation with autochthonous rhizobacteria improves metal rhizoaccumulation in native *Spartina maritima*. *J. Hazard. Mater.* **300**, 263–271 (2015).
241. Li, D., Niu, Y. Y., Fan, M., Xu, D. L. & Xu, P. Focusing phenomenon caused by soil conductance heterogeneity in the electrokinetic remediation of chromium (VI)-contaminated soil. *Sep. Purif. Technol.* **120**, 52–58 (2013).
242. Liang, L. et al. Phytoremediation of heavy metal contaminated saline soils using halophytes: current progress and future perspectives. *Environ. Rev.* **25**, 269–281 (2017).
243. Aboughalma, H., Bi, R. & Schlaak, M. Electrokinetic enhancement on phytoremediation in Zn, Pb, Cu and Cd contaminated soil using potato plants. *J. Environ. Sci. Health A* **43**, 926–933 (2008).
244. Burges, A., Alkorta, I., Epelde, L. & Garbisu, C. From phytoremediation of soil contaminants to phytomanagement of ecosystem services in metal contaminated sites. *Int. J. Phytoremediat.* **20**, 384–397 (2018).
245. Conesa, H. M., Evangelou, M. W. H., Robinson, B. H. & Schulin, R. A critical view of current state of phytotechnologies to remediate soils: still a promising tool? *Sci. World J.* **2012**, 173829 (2012).
246. Robinson, B. H., Bañuelos, G., Conesa, H. M., Evangelou, M. W. H. & Schulin, R. The phytomanagement of trace elements in soil. *Crit. Rev. Plant Sci.* **28**, 240–266 (2009).
247. Evangelou, M. W. H., Papazoglou, E. G., Robinson, B. H. & Schulin, R. in *Phytoremediation: Management of Environmental Contaminants* Vol. 1 (eds Ansari, A. et al.) 115–132 (Springer, 2015).
248. Hou, D. & Ok, Y. S. Soil pollution — speed up global mapping. *Nature* **566**, 455–455 (2019).
249. O'Connor, D. et al. Biochar application for the remediation of heavy metal polluted land: a review of in situ field trials. *Sci. Total Environ.* **619–620**, 815–826 (2018).
250. United Nations Environment Assembly of the United Nations Environment Programme. Implementation plan "Towards a pollution-free planet" (UNEP, 2019).
251. Jiang, Y. et al. Field scale remediation of Cd and Pb contaminated paddy soil using three mulberry (*Morus alba* L.) cultivars. *Ecol. Eng.* **129**, 38–44 (2019).
252. Li, J. T., Liao, B., Dai, Z. Y., Zhu, R. & Shu, W. S. Phytoextraction of Cd-contaminated soil by carambola (*Averrhoa carambola*) in field trials. *Chemosphere* **76**, 1233–1239 (2009).
253. Mayerová, M. et al. Non-enhanced phytoextraction of cadmium, zinc, and lead by high-yielding crops. *Environ. Sci. Pollut. Res.* **24**, 14706–14716 (2017).
254. van Slycken, S. et al. Field evaluation of willow under short rotation coppice for phytomanagement of

metal-polluted agricultural soils. *Int. J. Phytoremediat.* **15**, 677–689 (2013).
255. Ma, J., Lei, E., Lei, M., Liu, Y. & Chen, T. Remediation of arsenic contaminated soil using malposed intercropping of *Pteris vittata* L. and maize. *Chemosphere* **194**, 737–744 (2018).
256. Meers, E. et al. The use of bio-energy crops (*Zea mays*) for 'phytoattenuation' of heavy metals on moderately contaminated soils: a field experiment. *Chemosphere* **78**, 35–41 (2010).
257. Favas, P. J. C., Pratas, J., Varun, M., D'Souza, R. & Paul, M. S. in *Environmental Risk Assessment of Soil Contamination* (ed. Hernandez Soriano, M. C.) 485–518 (InTech, 2014).
258. Ahemad, M. Remediation of metalliferous soils through the heavy metal resistant plant growth promoting bacteria: paradigms and prospects. *Arab. J. Chem.* **12**, 1365–1377 (2019).

**Acknowledgements**
This work was carried out with the support of the Cooperative Research Program for Agriculture Science and Technology Development (project no. PJ01475801), Rural Development Administration, Republic of Korea. This work was also supported by the National Research Foundation of Korea (NRF) (NRF-2015R1A2A2A11001432) and the NRF Germany-Korea Partnership Program (GEnKO Program) (2018–2020). Y.S.O. and A.D.I. were partly supported by the KU Future Research Grant (KU FRG) Fund, Korea Biochar Research Center (KBRC) Fund and the Association of Pacific Rim Universities (APRU) Sustainable Waste Management Program from the Korea University, Republic of Korea. D.H. and D.O.C. were supported by the National Key Research and Development Program of China (grant no. 2018YFC1801300), the National Water Pollution Control and Treatment Science and Technology Major Project (no. 2018ZX07109-003) and the Ministry of Ecology and Environment's National Soil Pollution Investigation Project in China.

**Author contributions**
Y.S.O., D.H., D.O.C., A.D.I., J.L. and D.C.W.T. researched data for the article. Y.S.O., D.H. and D.C.W.T. made a substantial contribution to the discussion of content. Y.S.O., D.H., D.O.C., A.D.I., D.S.A. and J.L. contributed to the writing of the review. Y.S.O., D.H., D.O.C., A.D.I., D.S.A., D.C.W.T., D.L.S., Y.Y. and J.R. reviewed and edited the manuscript before submission.

**Competing interests**
The authors declare no competing interests.

**Publisher's note**
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.