

M1D1:

Learn best practices for mammalian cell culture

1. Prelab discussion
2. Orientation quiz
3. Cell culture exercises



Mark your calendar!

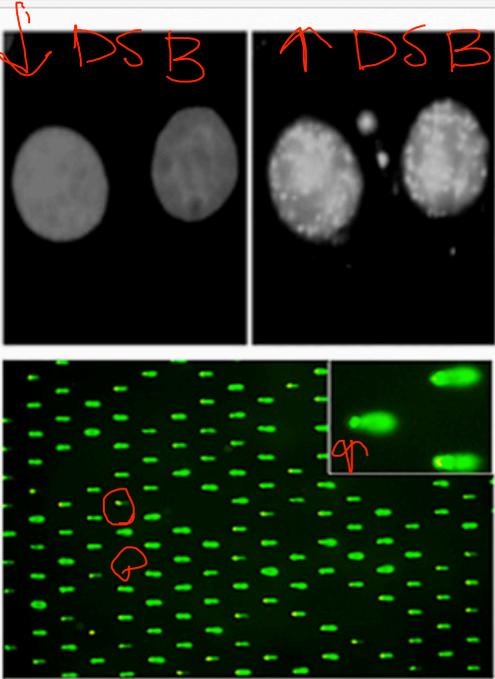
- **Data summary** (15%)
 - completed in teams and submitted via Stellar
 - draft due 10/4, final revision due 10/14
- **Mini-presentation** (5%)
 - completed individually and submitted via Gmail
 - due 10/11
- **Laboratory quizzes** (collectively 5%)
 - scheduled for M1D4 and M1D7
- **Notebook** (collectively 5%)
 - one entry will be graded by Aimee 24 hr after M1D7
- **Blog** (part of 5% Participation)
 - due 10/5 via Blogspot



Overview of M1: genomic instability

Research question: Does exposure to As inhibit, or decrease, repair of H_2O_2 -induced DNA damage, raising the possibility that combined exposure is an important risk to public health?

H_2O_2 oxidizing agent.



1. Use repair foci experiment to measure DNA breaks

- Examine effect of H_2O_2 +/- As on double strand DNA breaks by measuring $\gamma H2AX$ foci formation

$\gamma H2AX$ assay

2. Use high-throughput genome damage assay to measure DNA damage

- Measure effects of H_2O_2 +/- As on DNA damage by measuring DNA migration in agarose matrix

Comet Chip

We will use human lymphoblastoid cells

- Specifically, what cell line are we using in M1?

M25

- What are primary cells? Why are they difficult to use in experiments?

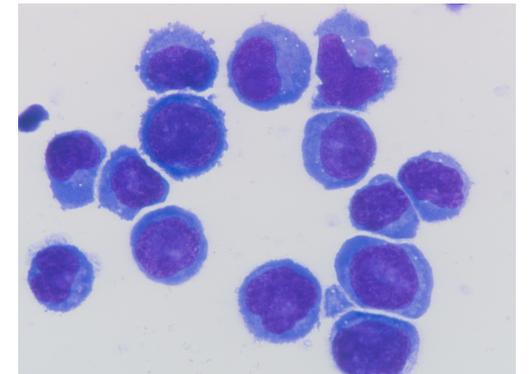
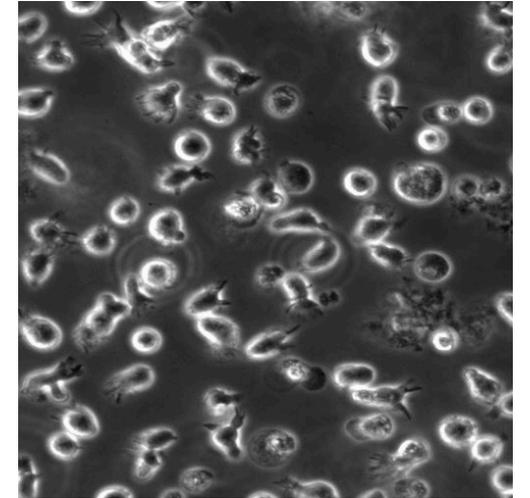
taken directly from tissue
division is not indefinite

- Why are cancer cells easier to use in experiments?

immortal

- What growth conditions are important when culturing mammalian cells?

Space, growth factors, temperature, pH



Growth medium is used to culture cells

Food



- RPMI 1640 (Roswell Park Memorial Institute) **DEFINED**
Sugars / amino acids / pH indicator
butter / salt / water



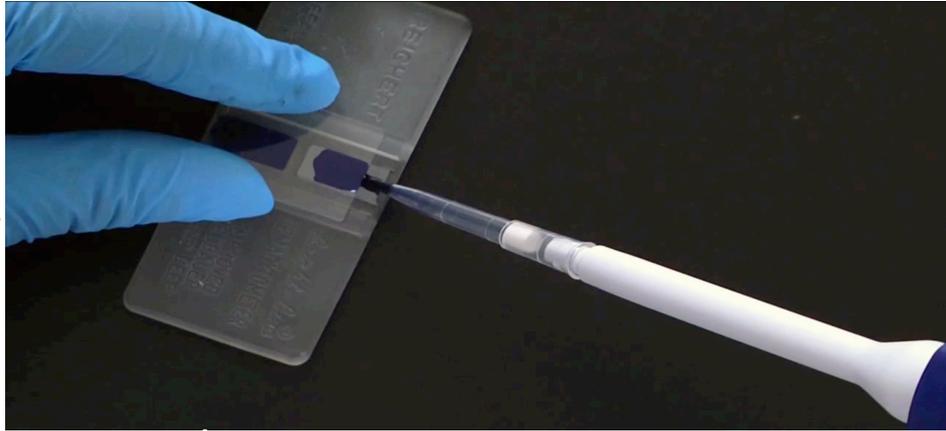
- FBS (fetal bovine serum) **UNDEFINED**
albumin / growth factors / cytokines
lipids / fatty acids

Non-food

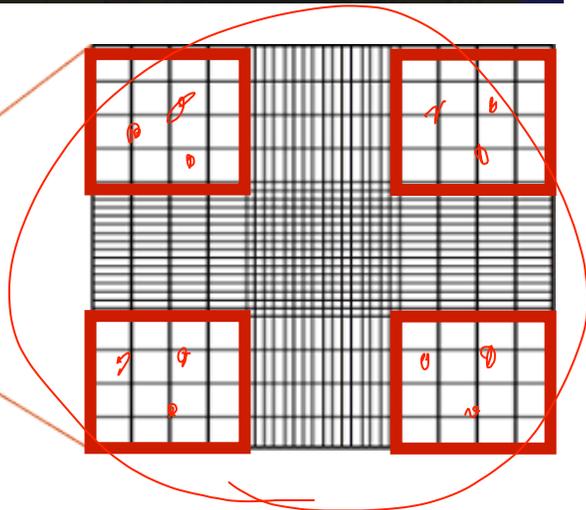


- Antibiotic solution: penicillin and streptomycin

Hemocytometers are used to count cells



- Trypan blue mixed with cell suspension at 1:10 ratio, then 10 μL added to hemocytometer
- Cells within highlighted sections of the hemocytometer grid are counted



cells / mL = average # of cells in the 4 highlighted boxes * 10,000

$$3 \times 10,000$$

The language of cell culture

- Confluence

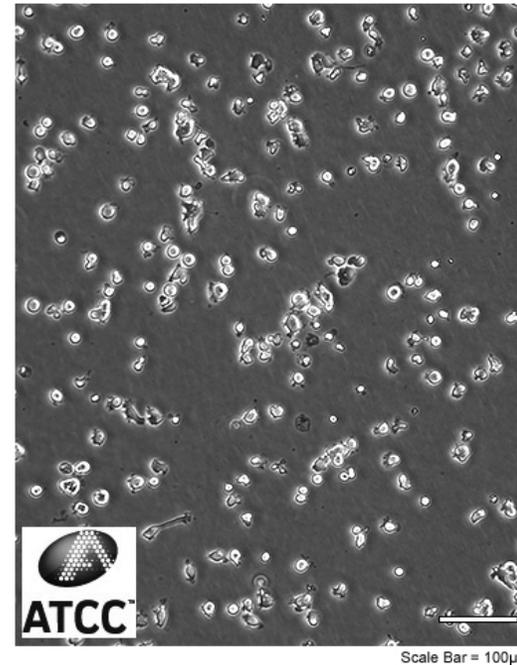
Density / # of cells

- Splitting / Sub-culturing

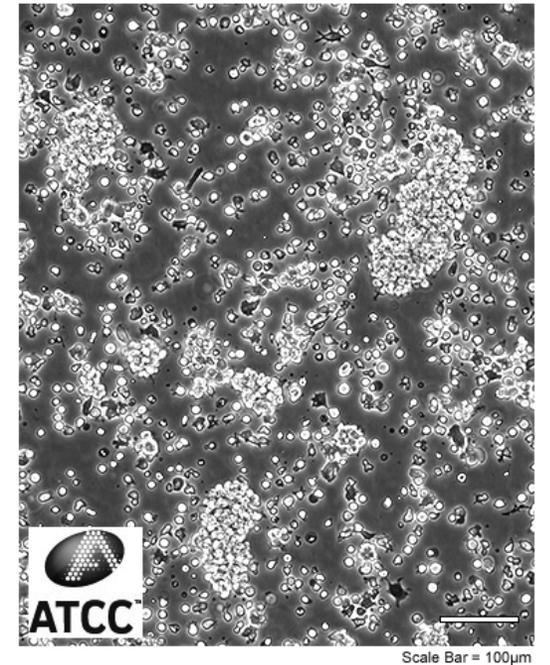
*diluting cell culture
→ nutrients / space*

- Seeding

for experiment → exact # of cells



Low density



High density

For today...

- Choose a team name!
- Complete Orientation quiz
 - Submit to Stellar by 10 pm
- Work through cell culture exercises
 - Be sure to record your notes in your laboratory notebook



For M1D2...

- Prepare a template for Benchling laboratory notebook entries
- Be sure to share your Benchling laboratory notebook

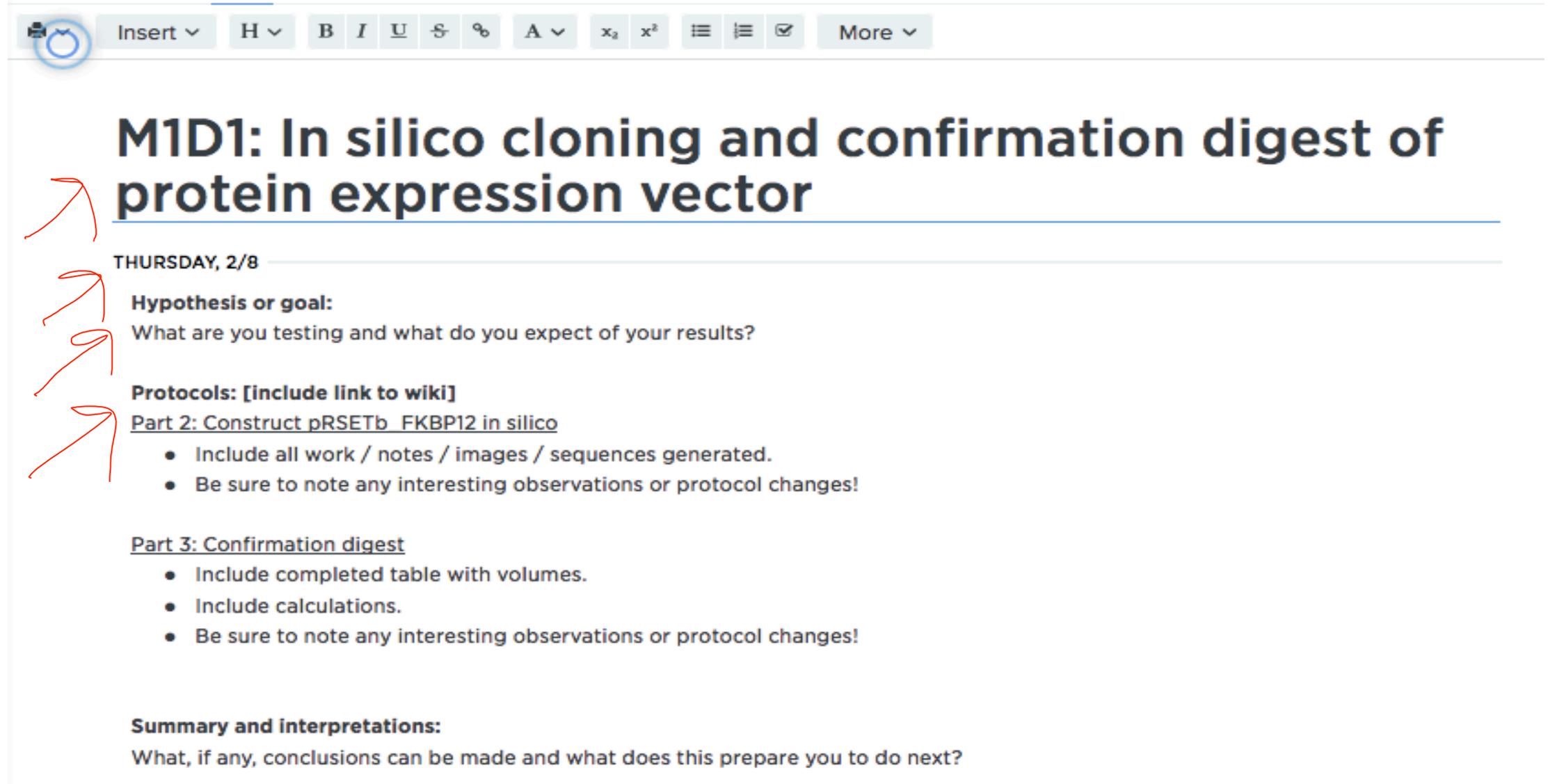
What should go in your notebook?

Assignments
tab

| Laboratory notebook entry component: | Points: | | |
|--|----------|---------|---------|
| | Complete | Partial | Missing |
| Date of experiment (include Module#/Day#) and Title for experiment | 1 | 0.5 | 0 |
| Hypothesis or goal / purpose | 2 | 1 | 0 |
| Protocols (link to appropriate wiki sections) | 1 | 0.5 | 0 |
| Answering questions embedded in wiki sections | 5 | 3 | 0 |
| Observations from demonstrations and video tutorials | 3 | 2 | 0 |
| *Visual details | | | |
| *Qualitative information | | | |
| *Raw data | | | |
| Data analysis | 3 | 2 | 0 |
| *Calculations | | | |
| *Graphs and Tables | | | |
| Summary and interpretation of data | 3 | 2 | 0 |
| *What did you learn? | | | |
| *How does this information fit into the larger scope of the project? | | | |
| Information is clear | 2 | 1 | 0 |
| All days represented | 5 | 3 | 0 |
| OVERALL /25 | | | |

Be sure to include your responses to the prompts within the laboratory exercises!

How should you format your notebook?



The image shows a screenshot of a digital notebook interface. At the top, there is a toolbar with various editing options like 'Insert', 'H', 'B', 'I', 'U', 'A', 'x₂', 'x²', and 'More'. Below the toolbar, the notebook content is displayed. The main heading is 'M1D1: In silico cloning and confirmation digest of protein expression vector', which is underlined. Below this, the date 'THURSDAY, 2/8' is shown. The entry is organized into sections: 'Hypothesis or goal:' with the question 'What are you testing and what do you expect of your results?'; 'Protocols: [include link to wiki]' with two sub-sections: 'Part 2: Construct pRSETb FKBP12 in silico' and 'Part 3: Confirmation digest'. Each sub-section has a list of bullet points. Finally, there is a 'Summary and interpretations:' section with the question 'What, if any, conclusions can be made and what does this prepare you to do next?'. Red arrows on the left side of the page point to the title, the hypothesis section, the protocols section, and the first bullet point of Part 2.

M1D1: In silico cloning and confirmation digest of protein expression vector

THURSDAY, 2/8

Hypothesis or goal:
What are you testing and what do you expect of your results?

Protocols: [include link to wiki]

Part 2: Construct pRSETb FKBP12 in silico

- Include all work / notes / images / sequences generated.
- Be sure to note any interesting observations or protocol changes!

Part 3: Confirmation digest

- Include completed table with volumes.
- Include calculations.
- Be sure to note any interesting observations or protocol changes!

Summary and interpretations:
What, if any, conclusions can be made and what does this prepare you to do next?

How should you organize your notebook?

- Title your project “20.109(F20) YourName”
 - Make each module a new folder
 - Make each day a new entry within module folder
- Share the project with Instructors and Aimee
 - Right-click and choose ‘settings’
 - Add collaborators by email
 - nlyell@mit.edu
 - amoise@mit.edu
 - rcmeyster@mit.edu
 - mebane@mit.edu

