

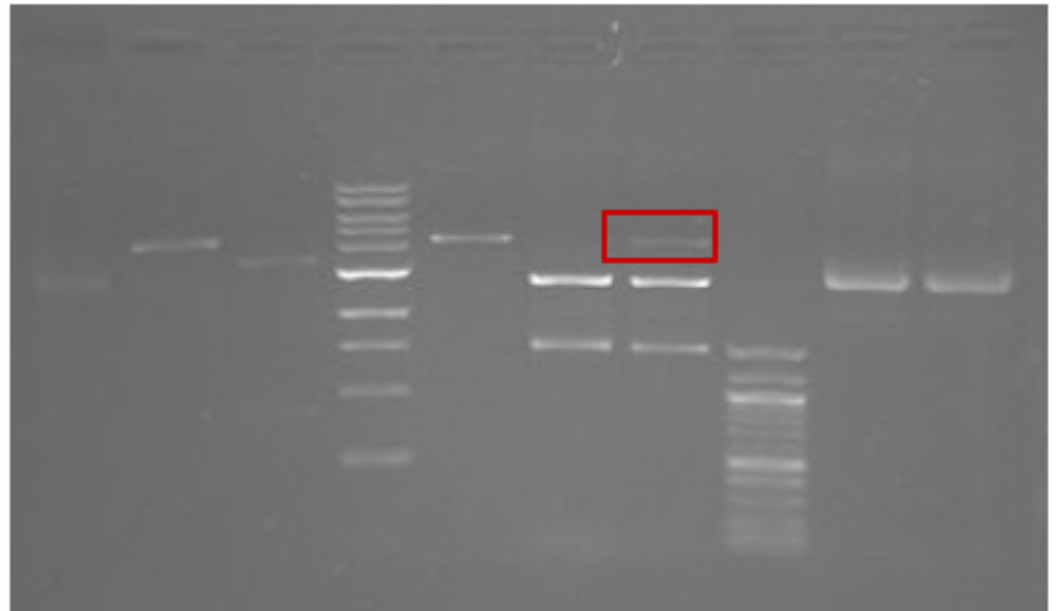
- Announcements
- Quiz
- Pre-lab Lecture
  - ❖ SDS-PAGE Part 1
  - ❖ Affinity purification recap
  - ❖ Today in Lab (M2D6)

# Announcements

- Lab etiquette reminder
  - Upcoming OH
    - Mon, 4/8, 3-4 pm
    - Wed, 4/17, by appt (no class Tue)
    - Sat or Sun Apr 20/21, c. 2:30-4:30 pm
  - Revision due tomorrow at 11 AM
  - Hey, what happened last time?
    - S08, S09, S10: no antibiotics during induction
    - S12: odd cell behavior (plasmid loss?), so used antibiotics, *but* spring break was after D3 not D4
    - S13: combo of week-old plates and fresh antibiotics most likely challenged cell survival+growth
- + discuss evals next time

# Diagnostic digests and ODs

- What might the identities of extra bands be?  
*from partial digestion (rxn pool, not binary)*
- What could *unaccounted for* extra bands indicate?  
*\* activity; contamination; other mutation*
- Besides cell normalization, why might getting -IPTG and +IPTG OD values be useful?



Courtesy T/R Platinum.

Expect: 3300/800 REF; 2700/1500 MUT

*mutant growth phenotypes; [protein] expectations*

# SDS-PAGE preparation

acrylamide - toxic

- You will make whole cell extracts with equal cell #s

- Based on  $OD_{600}$  reading, normalize ①  $OD = 1.0$  ②  $OD = 0.5$

$V_{max} = 15 \mu L$

②  $15 \mu L$

①  $7.5 \mu L + 7.5 \mu L H_2O$



- Gel separates proteins based on size, shape, charge

- Sample preparation (finish next time)

- SDS: coat proteins w/(-) charge

- $\beta$ -Me: break S-S bonds

- Boiling: denaturing higher-order structures

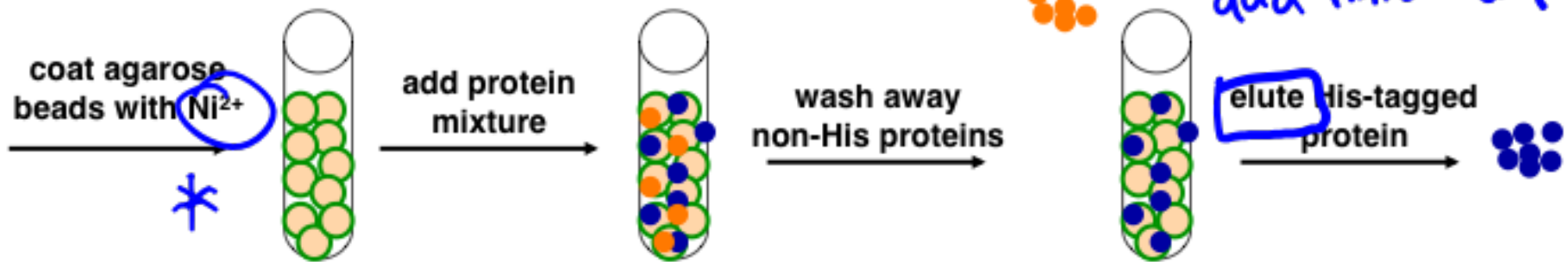
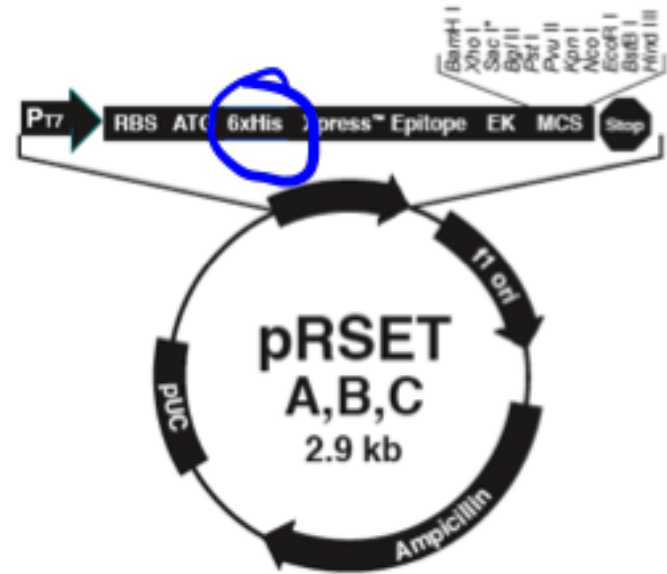
- Sample Buffer has SDS,  $\beta$ -Me, plus glycerol, BPB dye

make uniform

in hood

# Affinity purification

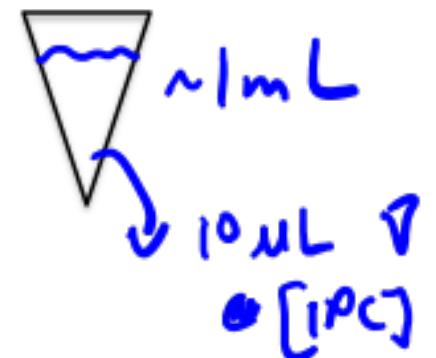
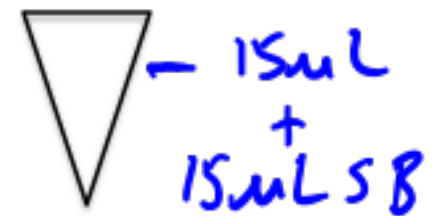
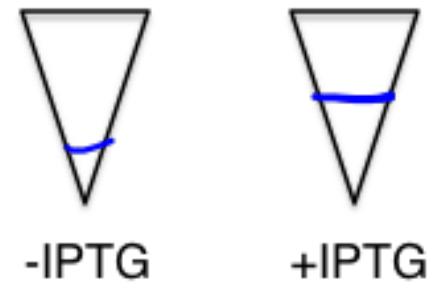
- Basis: His tag (6x)  
binds to metals\*
- elute by competition



⊕ desalting → bye-bye imidazole!

# Today in Lab (M2D6)

- Lyse cell pellets in BPER
  - BSA “carrier,” protease inhibitors
  - Add lysis enzymes  $\times$   $\uparrow$  1:2000
- Prep an aliquot for SDS-PAGE
- Purify IPC protein from the rest (long!)
  - Two steps: affinity purification, desalting
  - Immediately take 10  $\mu$ L aliquot and measure concentration
  - The rest is stabilized w/BSA, to be titrated against calcium next time



\* low [IPC] may still give high enough fluorescence (sensitivity)