

# M2D5: Purify Protein

4/5/13

Mod1 Report

1. Reflections on your ~~journal club~~ re-writes -- send them to me today or sometime this weekend.

2. Lab treat.

3. More happy things:



# Diagnostic Gels: How did you do?

IPC WT

IPC KpnI

D24H KpnI

1 Kb Ladder

IPC StyI

G25S #1 StyI

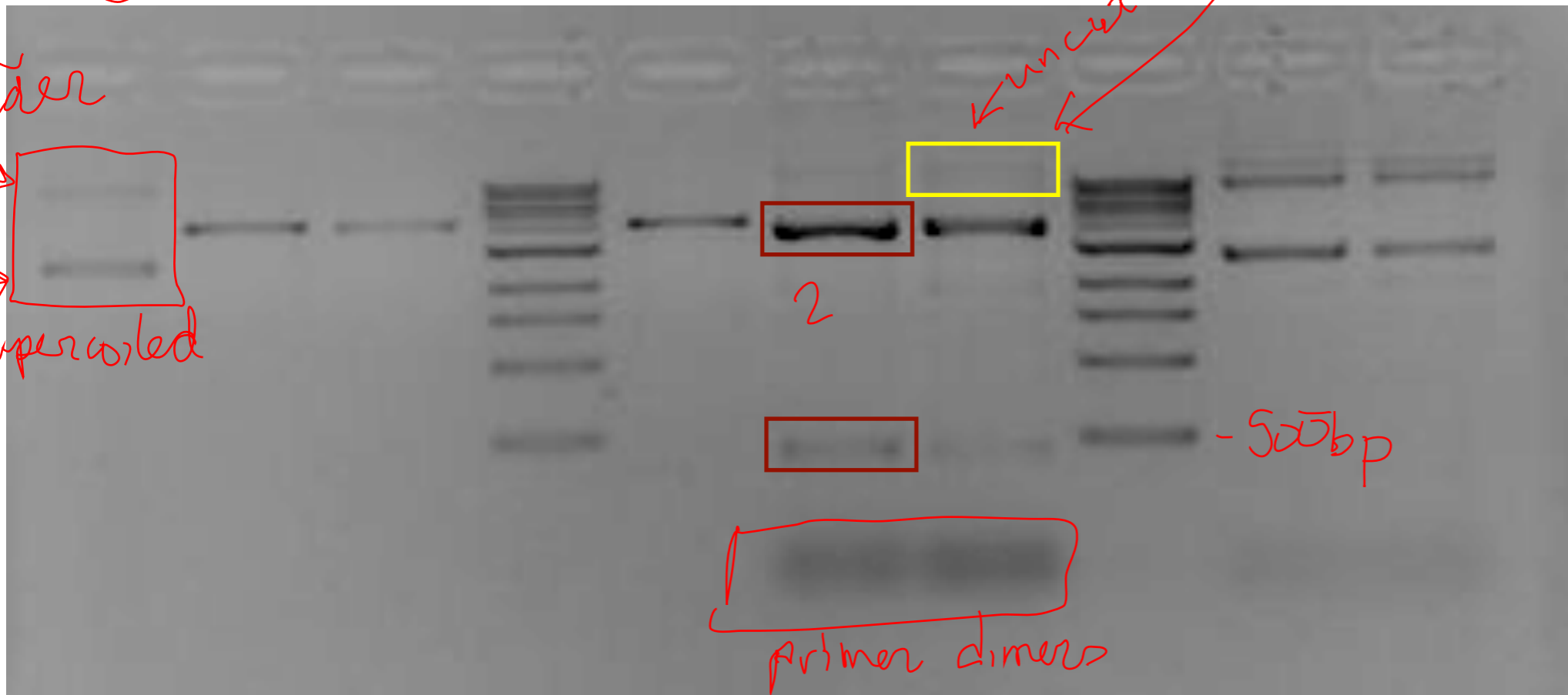
G25S #2 StyI

1 Kb Ladder

G25S #1

G25S #2

- religation v. low  
- inefficient rxn



1. Extra bands?

2. Unaccounted for bands?

contamination  
random mutation  
\*activity

**Pink Team!**

# Protein purification: Protocol overview



lyse cells and  
extract protein

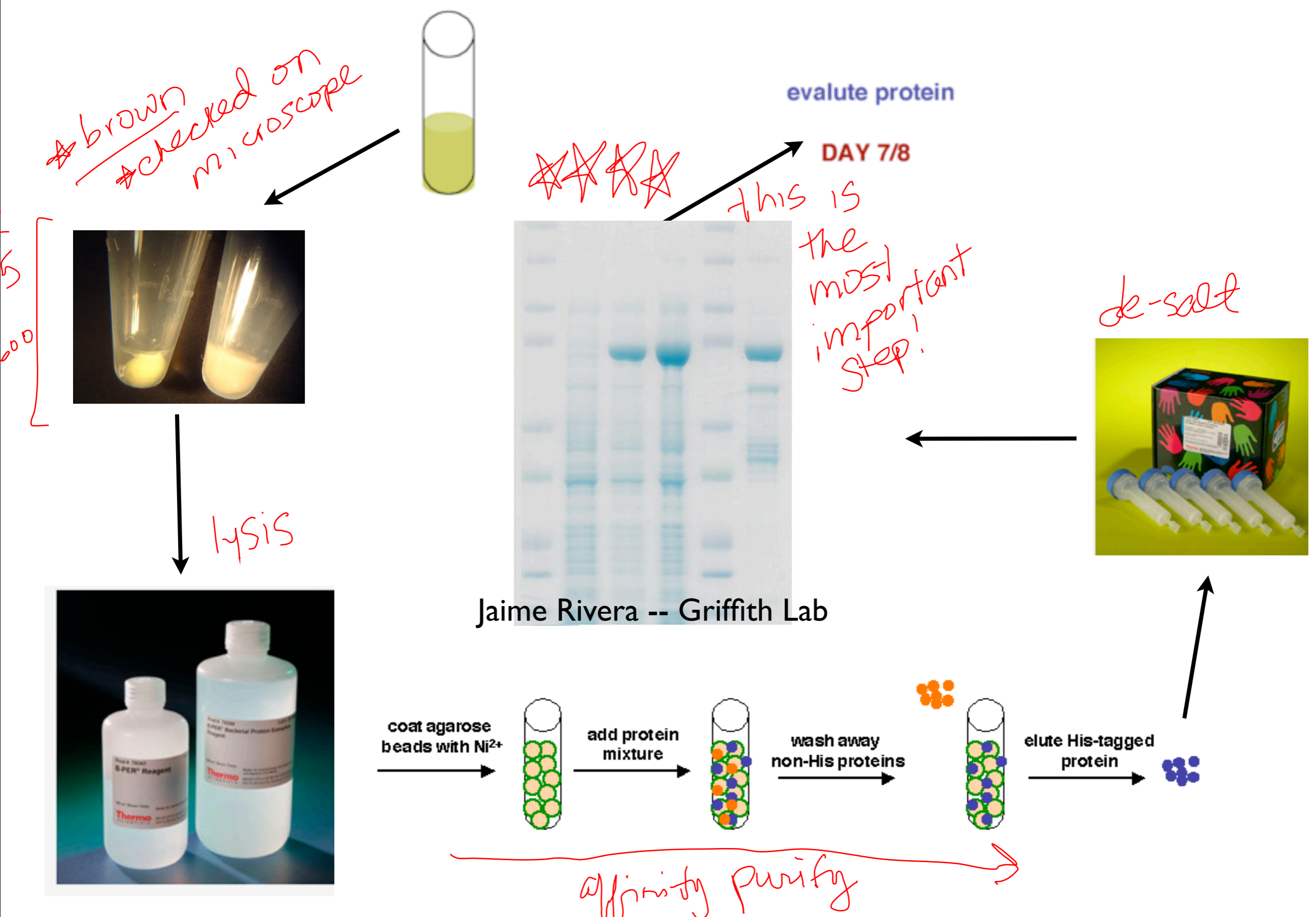


**DAY 6**

evalute protein

**DAY 7/8**

# Protein purification: Protocol overview



# Protein purification: Get it out of there! ← lyse

## Lysis Buffer: BPER from Pierce

- + Lysozyme → blow apart bacteria
- + DNASE I → chew up DNA



## Prepare samples for SDS-PAGE analysis

Normalize samples by cell number. Why?

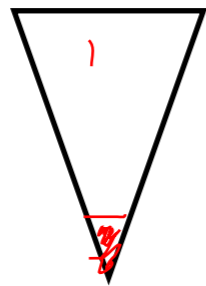
OD600 - compare across samples

OD600 = 0.15

1

0.75

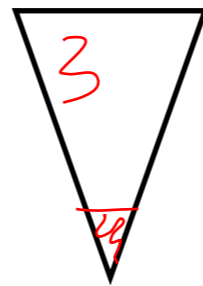
1



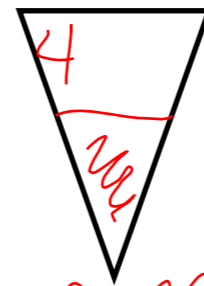
-IPTGx3



WT



D24H



G255

$V_t = 15 \mu\text{L}$

$V_{S1} = 15 \mu\text{L}$  lysate

$V_{S2} = 84 = 7.5 \mu\text{L}$  lysate +  $7.5 \mu\text{L}$  H<sub>2</sub>O

$V_{S3} = 10 \mu\text{L}$  lysate +  $5 \mu\text{L}$  H<sub>2</sub>O

Get your cell counts off the M2D5 talk page (pick your clone! 3x)

Thanks Agi!!



# Protein purification: SDS-PAGE Analysis

size  
charge

Some details:

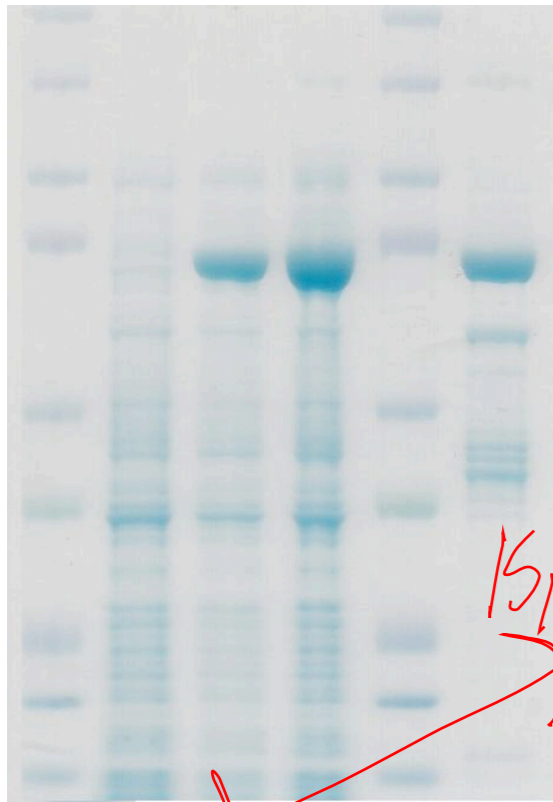
1. Polyacrylamide is Toxic - Neurotoxin

2. What is in the loading buffer? Why?

- ★ SDS: Neg charge coating
- ★  $\beta$ ME: breaks S-S bonds (denature)
- ★ Glycerol: hold sample in well
- ★ Dye: visualize migration

Last step: Boil it!

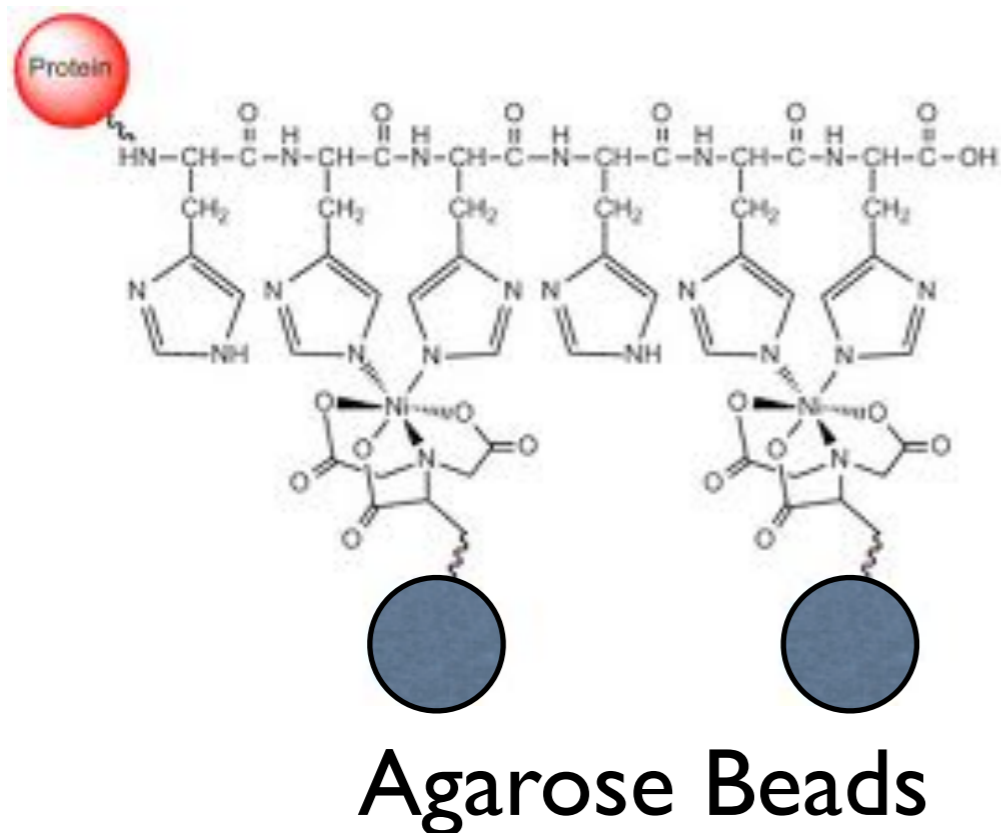
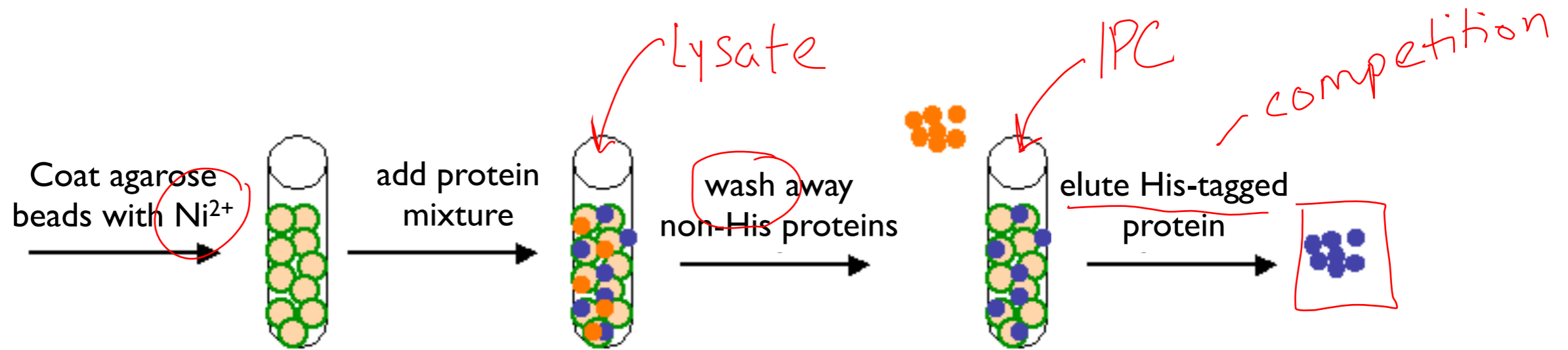
next time



Hood

15µl

# Protein purification: Affinity purification + desalting

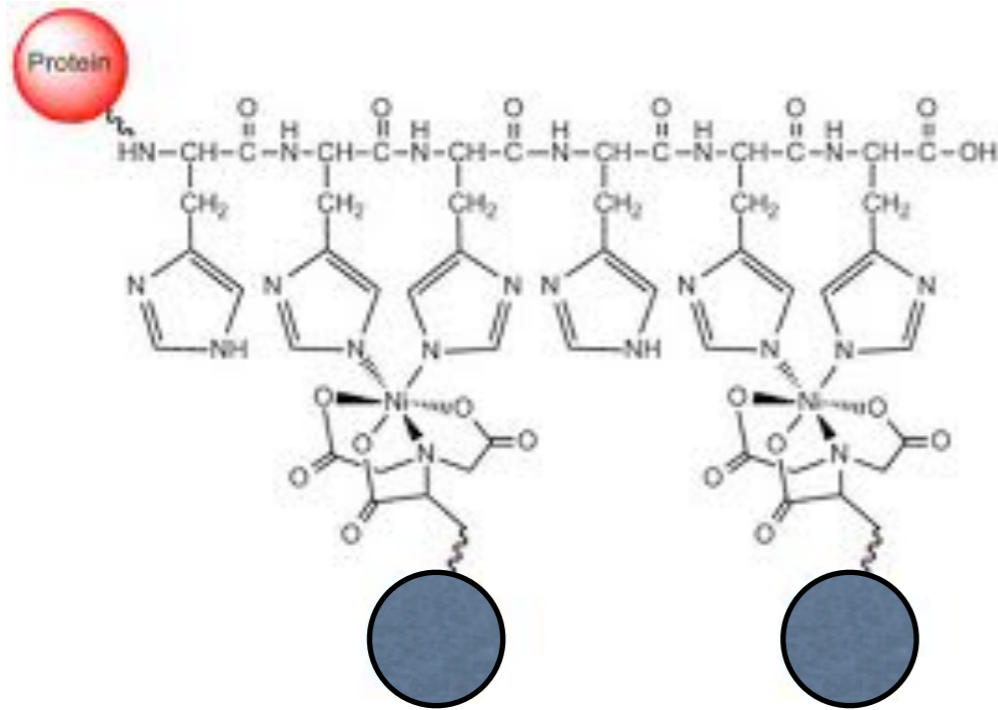


Elute with excess imidazole --

But -- binding and wash buffer also have imidazole. Why?

- weak binding of off-target proteins
- increases purity of His-tagged protein

# Protein purification: Affinity purification + desalting

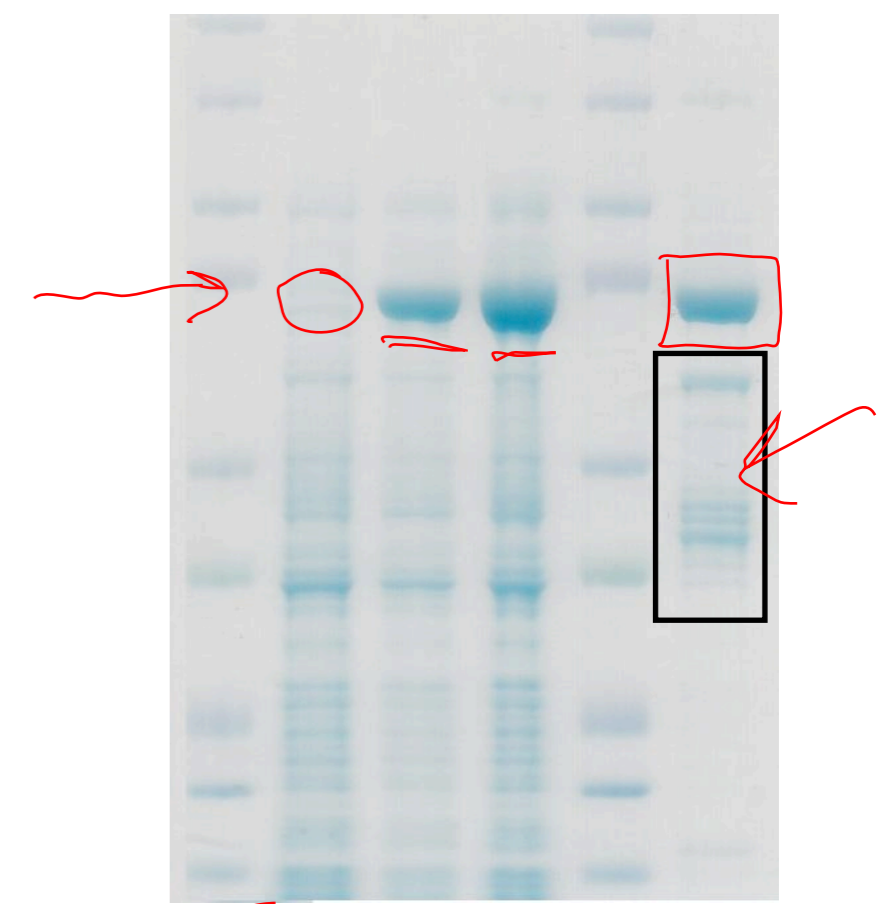


Agarose Beads

\*Do not use sepharose beads!

The CRAPome: A Contaminant Repository for Affinity Purification Mass Spectrometry Data; Alexey Nesvizhskii<sup>1</sup>; Dattatreya Mellacheruvu<sup>1</sup>; Zachary Wright<sup>1</sup>; Amber Couzens<sup>2</sup>; Jean-Philippe Lambert<sup>2</sup>; Nicole St-Denis<sup>2</sup>; Tuo Li<sup>3</sup>; Yana Miteva<sup>3</sup>; Simon Hauri<sup>4</sup>; Mihaela Sardi<sup>5</sup>; Teck Low<sup>6</sup>; Vincentius Halim<sup>6</sup>; Richard Bagshaw<sup>2</sup>; Nina Hubner<sup>7</sup>; Abdallah al-Hakim<sup>2</sup>; Annie Bouchard<sup>8</sup>; Denis Faubert<sup>8</sup>; Damian Fermin<sup>1</sup>; Wade Dunham<sup>2</sup>; Marilyn Goudreault<sup>2</sup>; Zhen-Yuan Lin<sup>2</sup>; Beatriz Gonzalez Badillo<sup>2</sup>; Tony Pawson<sup>2</sup>; Daniel Durocher<sup>2</sup>; Benoit Coulombe<sup>8</sup>; Ruedi Aebersold<sup>4</sup>; Giulio Superti-Furga<sup>9</sup>; Jacques Colinge<sup>9</sup>; Albert Heck<sup>6</sup>; Hyungwon Choi<sup>10</sup>; Matthias Gstaiger<sup>4</sup>; Shabaz Mohammed<sup>6</sup>; Ileana Cristea<sup>3</sup>; Keiryn Bennett<sup>9</sup>; Michael Washburn<sup>5</sup>; Brian Raught<sup>11</sup>; Robert Ewing<sup>12</sup>; Anne-Claude Gingras<sup>2</sup>; <sup>1</sup>University of Michigan, Ann Arbor, MI; <sup>2</sup>Samuel Lunenfeld Research Institute, Toronto, Canada; <sup>3</sup>Princeton University, Princeton, NJ; <sup>4</sup>Institute of Molecular Systems Biology, ETZ, Zurich, Switzerland; <sup>5</sup>Stowers Institute for Medical Research, Kansas City, MO; <sup>6</sup>Utrecht University, Utrecht, The Netherlands; <sup>7</sup>Radboud University, Nijmegen, The Netherlands; <sup>8</sup>Institut de recherches cliniques de Montréal, Montréal, Canada; <sup>9</sup>CeMM Research Center for Molecular Medicine, Vienna, Austria; <sup>10</sup>National University of Singapore, Singapore, Singapore; <sup>11</sup>Ontario Cancer Institute, Toronto, Canada; <sup>12</sup>Case Western Reserve University School of Medicine, Cleveland, OH

-IPTG culture sample  
 +IPTG culture sample  
 lysed bacteria after induction  
 MW Marker  
 purified protein (affinity purification)



Jaimel



# Today in lab:

★ Pick your clone! Lyse samples with BPER + enzymes.

★ Prepare samples for SDS-PAGE

★ Affinity purify your samples (this will take awhile)

★ De-salt. Why?

*- to get rid of imidazole  
- Ca<sup>2+</sup> binding*

*do not add  
BSA until you  
take sample  
for ~~SDS PAGE~~  
Bradford*

★ Immediately save 10 uL for the Bradford Assay

★ Add BSA to the rest as a carrier protein. Why?

 Sleep.