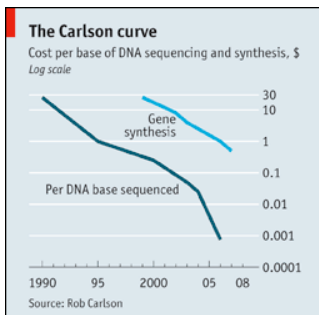


System Engineering

20.109(F09)
M2D5 lecture
10.29.09

Personal Genomics



One more acronym: GWAS



Height = 'model trait'
simple to measure and relatively constant

- in 2007, comparison made of 5000 genomes
- GWAS have revealed >40 loci involved in height

Editorial

Molecular Systems Biology 5 Article number: 273 doi:10.1038/msb.2009.32
 Published online: 19 May 2009
 Citation: *Molecular Systems Biology* 5:273

Personal phenotypes to go with personal genomes

Michael Snyder^{1,2}, Sherman Weissman³ & Mark Gerstein^{2,4,5}

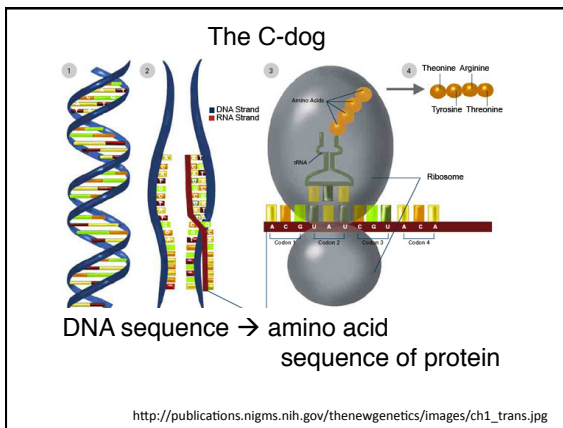
“the phenotyping of large numbers of individuals might well prove to be more expensive, complex and difficult to implement than the genomic sequencing.”

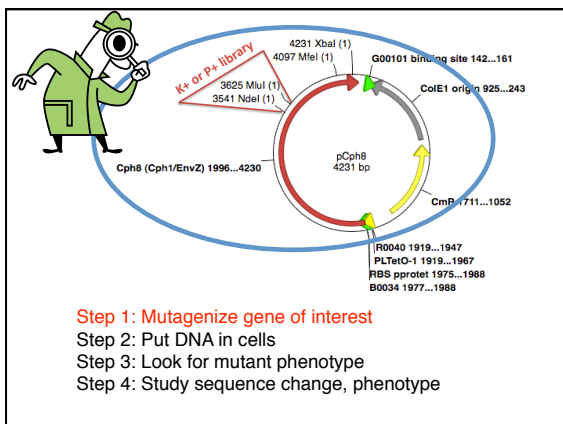
What to phenotype?

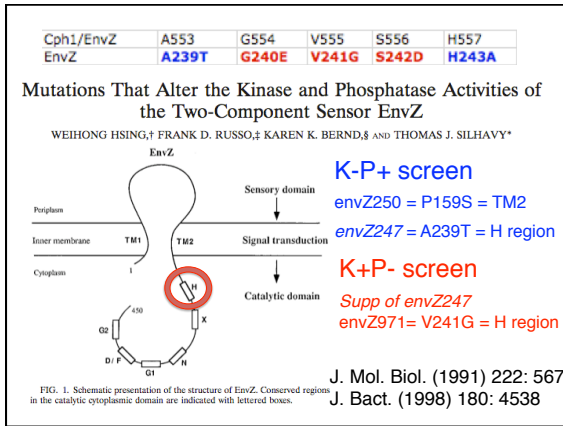
- ✓ height, blood pressure, medical history...
- behavioral (anxiety? Risk taking?)
- cognitive (intelligence?)

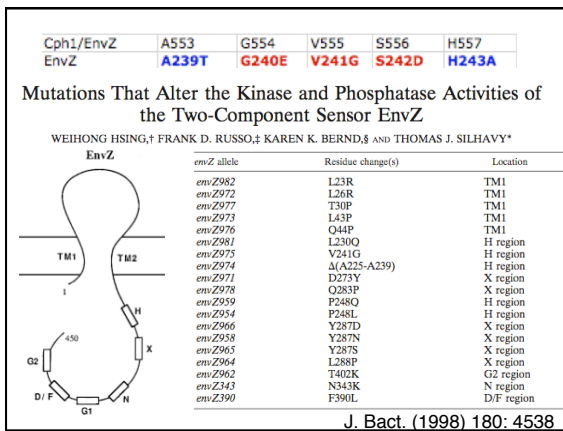
Standards for collection?

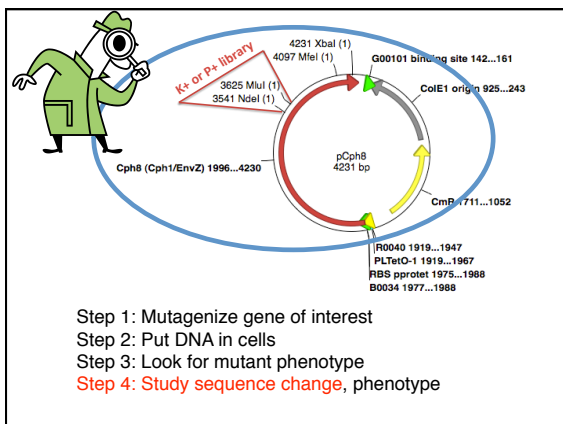
macroscopic VS molecular











Dideoxy Sequencing Method aka Sanger Method

dNTP: can be extended by DNA polymerase

3'-OH required for chain elongation

ddNTP: chain terminating!

No 3'-OH therefore, terminates chain

Dideoxy Sequencing Method aka Sanger Method

primer
→ ATTAGAGTCCG

"A" reaction: A, ATTA, (ATTAGA)
 "T" reaction: AT, ATT, (ATTAGAGT)
 "G" reaction: ATTAG, ATTAGACG, (ATTAGAGTCCG)
 "C" reaction: ATTAGAC, ATTAGACGTC, (ATTAGAGTCC)

sequencing gel

| | A | T | G | C |
|-------------|-----|-----|-----|-----|
| ATTAGAGTCCG | --- | --- | --- | --- |
| ATTAGAGTCC | --- | --- | --- | --- |
| ATTAGAGTCT | --- | --- | --- | --- |
| ATTAGAGCT | --- | --- | --- | --- |
| ATTAGAC | --- | --- | --- | --- |
| ATTAGA | --- | --- | --- | --- |
| ATTAG | --- | --- | --- | --- |
| ATTAA | --- | --- | --- | --- |
| ATT | --- | --- | --- | --- |
| AT | --- | --- | --- | --- |
| A | --- | --- | --- | --- |

Elongation Reactions:
 Template
 Primer
 DNA polymerase
 dNTPs (limiting)
 α P³²-dATP

4 Termination Reactions:
 dNTPs (not limiting)
 ddNTPs (either ddG, ddA, ddT or ddC)

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Sequencing K+ or P+ candidates

- Miniprep DNA as you did in Module 1 (Soln I, Soln II, Soln III, EtOH, wash, dry)
- Resuspend pellets in 40 μ l of water
- For sequencing, mix:
 - 2 μ l plasmid DNA
 - 6.4 μ l of a 1:100 dilution of the primer NO289
 - 15.6 μ l sterile water

pCp8 (CpH1/EnvZ) 1996_4230

ori origin 925_243

ClnR 1771_1052

binding site 142_161

PROK0 1919_1947

PLT001 1319_1357

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Also check your DNA by digest


- Provide 10 ul of each
- Teaching faculty will cut with

A= Nde
B= Mlu


to run on agarose gel and post

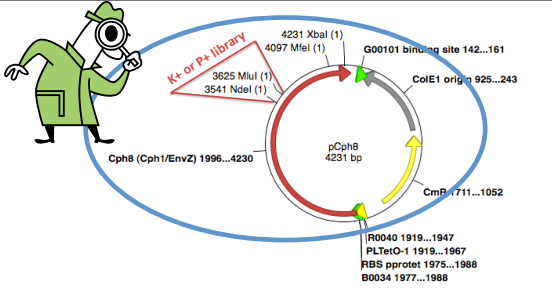
With "stuffer" frag expect: _____
With library frag expect: _____

"Stuffer frag"



Library fragments

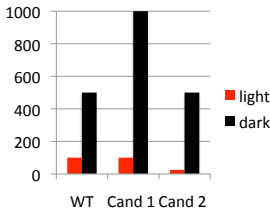




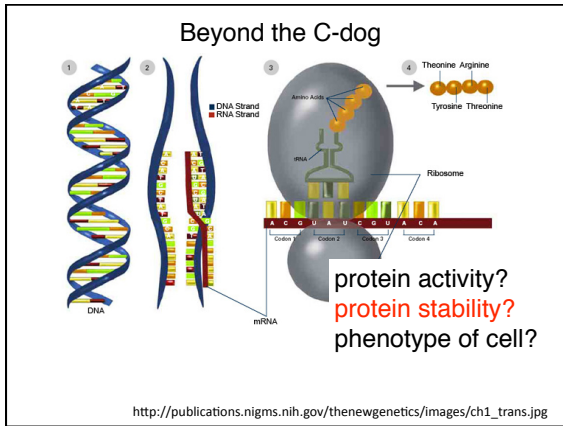
Step 1: Mutagenize gene of interest
Step 2: Put DNA in cells
Step 3: Look for mutant phenotype
Step 4: Study sequence change, phenotype

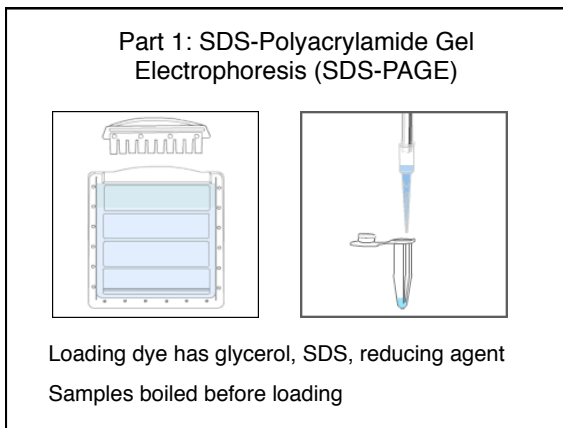
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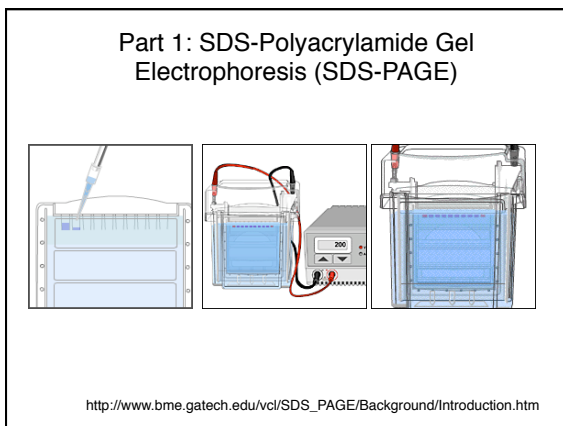
Grow 2 candidates in Light/Dark for β -gal next week



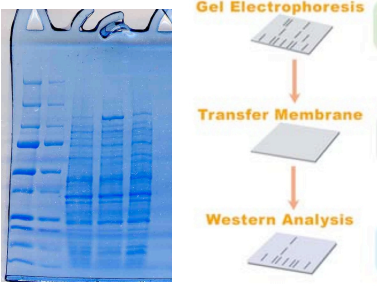
| Strain | Light | Dark |
|--------|-------|-------|
| WT | ~100 | ~500 |
| Cand 1 | ~100 | ~1000 |
| Cand 2 | ~50 | ~500 |







Part 2: Transfer for Western



Next week:
probe
membrane
with an
antibody

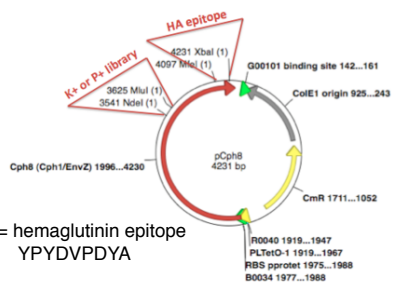
http://www.genscript.com/product_001/western_application/grp_id/60065/op/detail/Uvrag_Antibody_Analysis.html

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In lab you will:

- Measure OD600 of 1:10 of bacterial photography strain, Candidate 1, Candidate 2
- Harvest 4 OD
e.g. if 0.5 OD, harvest 8 ml of 1:10 or 0.8 ml of undiluted
- Isolate protein with lysis kit (enzymatic lysis of cells, spin out debris)
- Mix supernatant with loading dye
- Boil
- Load for SDS-PAGE along with markers, + control lysate

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HA = hemagglutinin epitope
YPYDVPDYA
