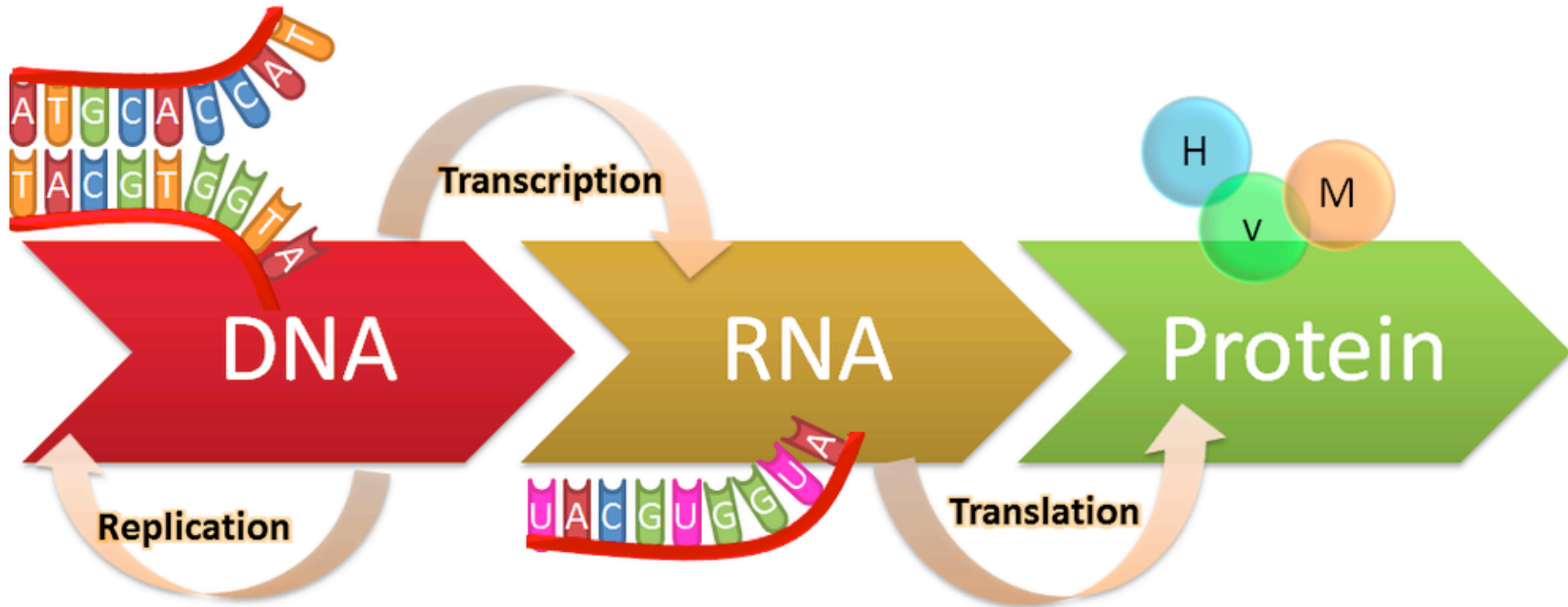
A grayscale microscopic image showing a cluster of cells with irregular, rounded shapes and some internal structure. The cells are arranged in a somewhat circular pattern, with some cells appearing more prominent than others. The background is a light, textured gray.

# **Module 1: Protein engineering**

- I Calcium signaling
- II Calmodulin
- III Protein engineering

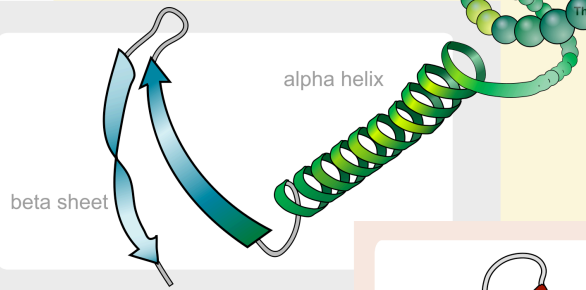
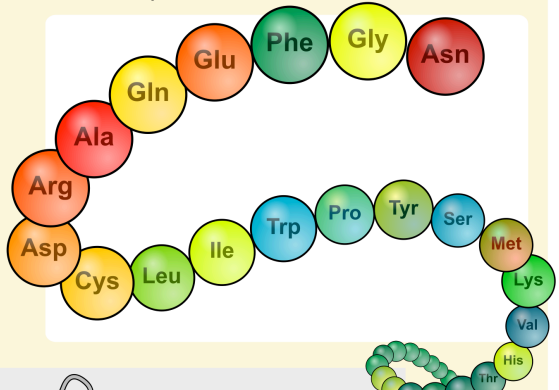
2/9/16

# The central dogma

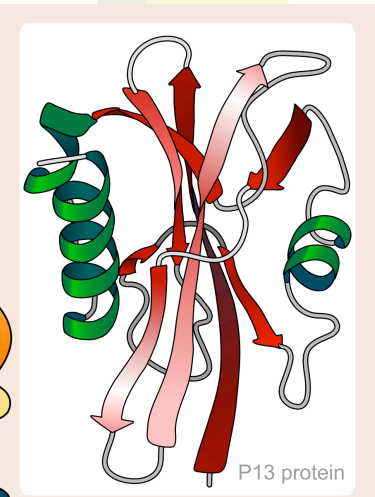


# What are proteins?

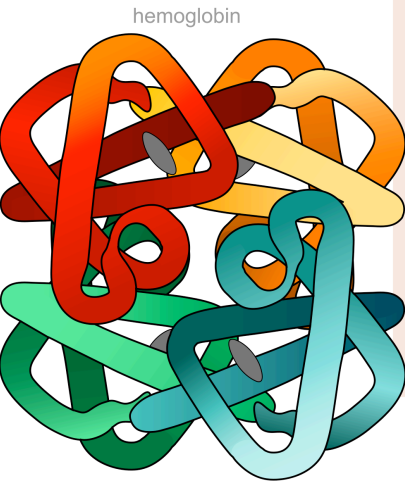
Primary structure  
amino acid sequence



Secondary structure  
regular sub-structures

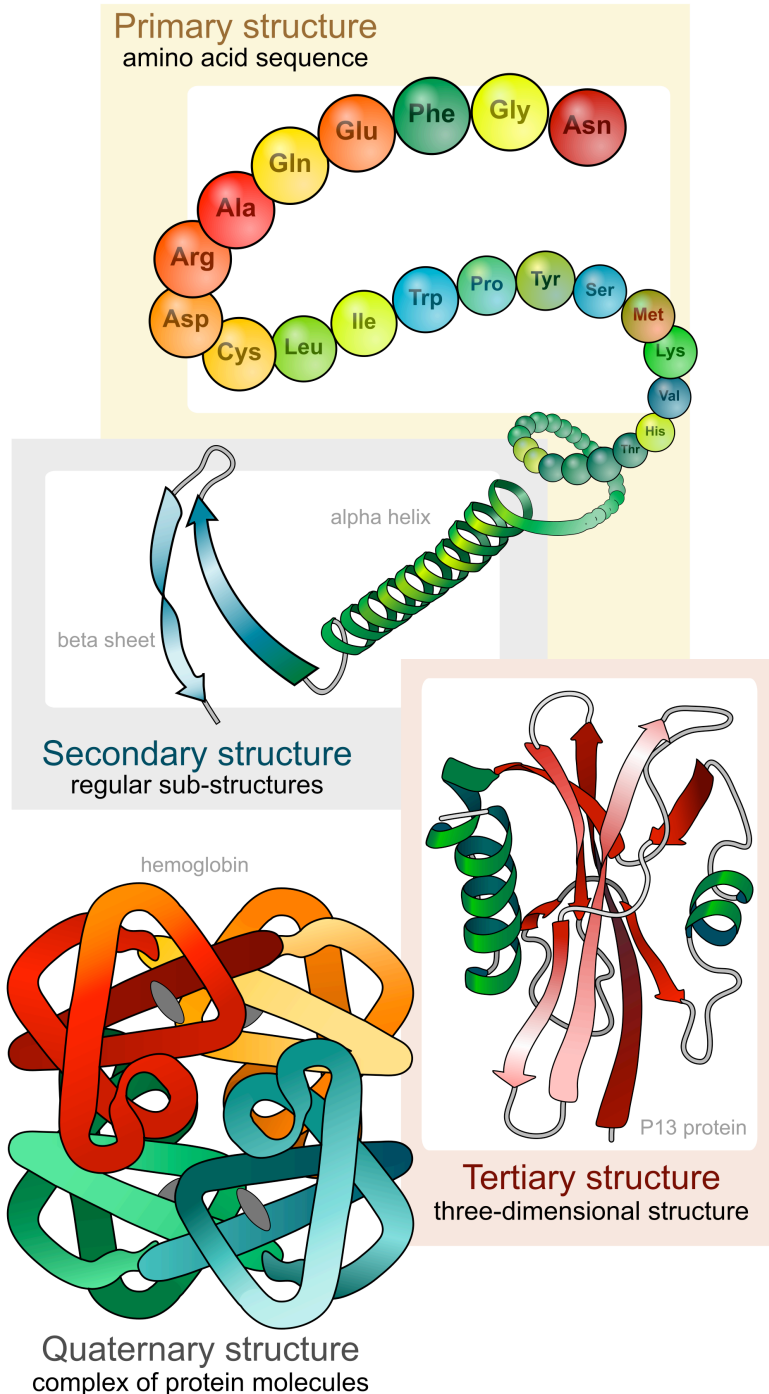


Tertiary structure  
three-dimensional structure



Quaternary structure  
complex of protein molecules

# What are proteins?



- Large macromolecules consisting of one or more long chains of amino acids
- Function related to conformation
  - H-bonding
  - Ionic interactions
  - Van der Waals forces
  - Hydrophobic packing

# Why do cells (we) need proteins?



# Why do cells (we) need proteins?

- Catalyze metabolic reactions
- DNA synthesis and maintenance
- Transport of molecules
- Structural and mechanical functions
- Propagation of and response to stimuli
- ...and all other cellular processes

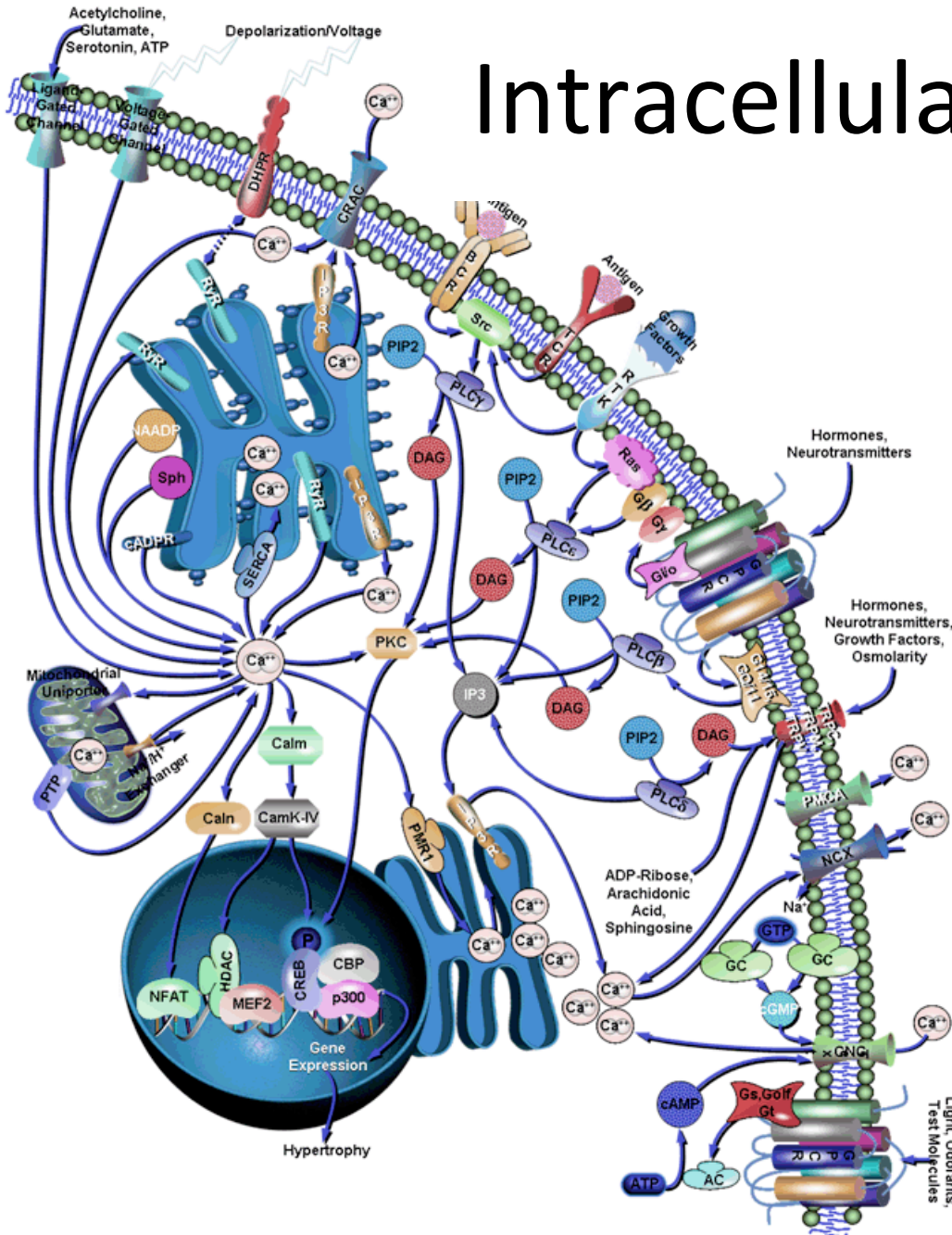
- Food



Your engineering task in Mod 2:

Alter  $\text{Ca}^{2+}$  binding properties of inverse  
pericam protein

# Intracellular Ca<sup>2+</sup> signaling



- Binding triggers change in protein charge and shape
- 20,000-fold gradient between intracellular and extracellular concentrations



# Ca<sup>2+</sup> is not just a structural element



Sidney Ringer, 1835-1910



Contractions



Contractions

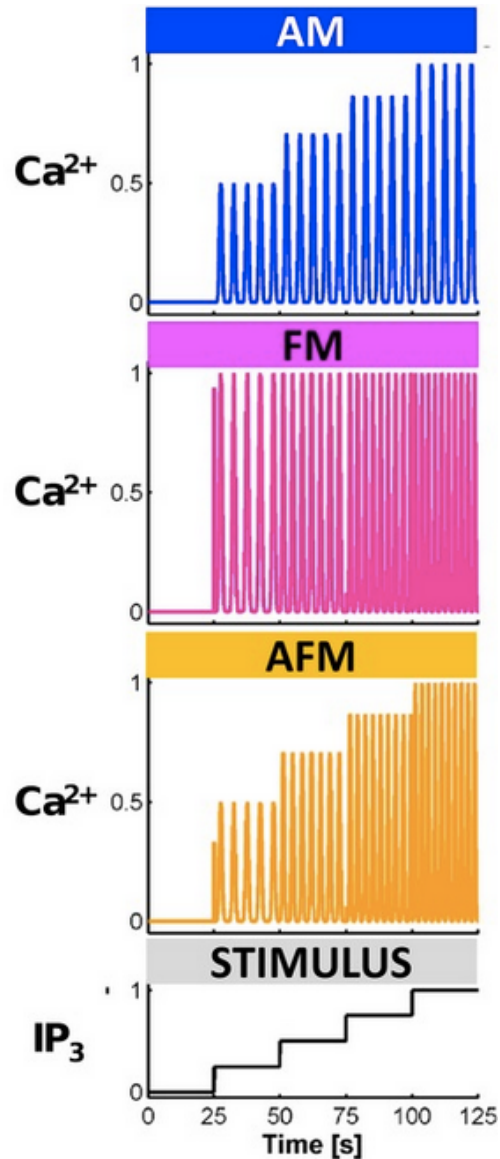
# Calcium is an essential messenger

- Contracts frog muscle fibers
- Activates ATPase activity of myosin
- Binding to myofibrils activates actomyosin
- Accumulated in sarcoplasmic reticulum vesicles
- Receptor, troponin C, mediates myofibrillar contractions
- EDTA chelation relieves muscle contractions

# Calcium signal is tightly controlled

- Regulated by channels and transporters
  - Maintain low cytosolic concentrations
  - Enable dynamic and rapid changes in stores
- Transduces signals from extracellular sources: hormones and growth factors
- Amplitude and frequency code for the signal
  - AM, FM, or AFM

# How is the $\text{Ca}^{2+}$ signal coded?

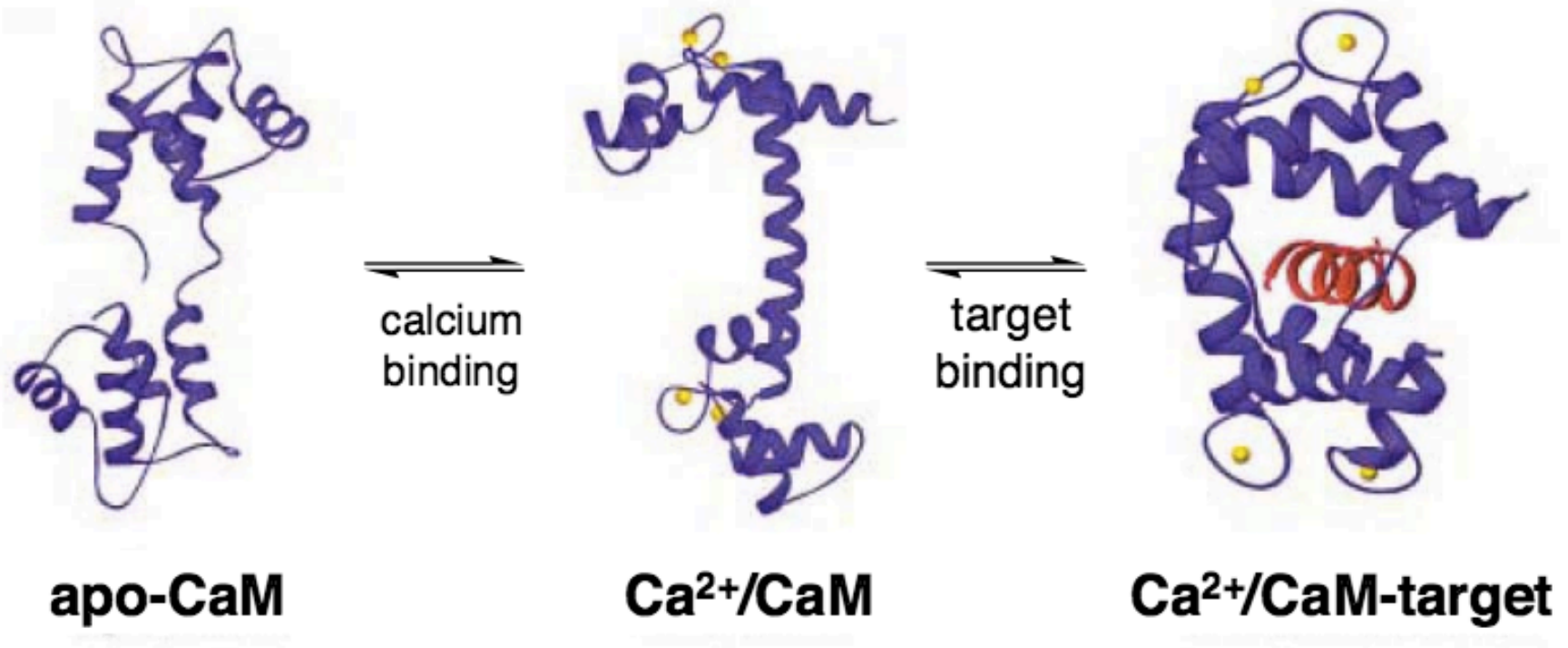


- AM (amplitude modulations)

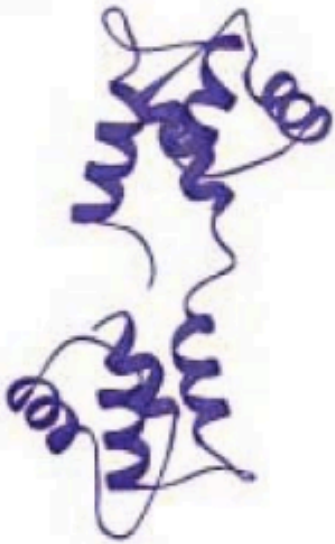
- FM (frequency modulations)

- AFM (amplitude and frequency modulations)

# Calmodulin (CaM) translates $\text{Ca}^{2+}$ code



# Ca<sup>2+</sup>/CaM binding is dynamic

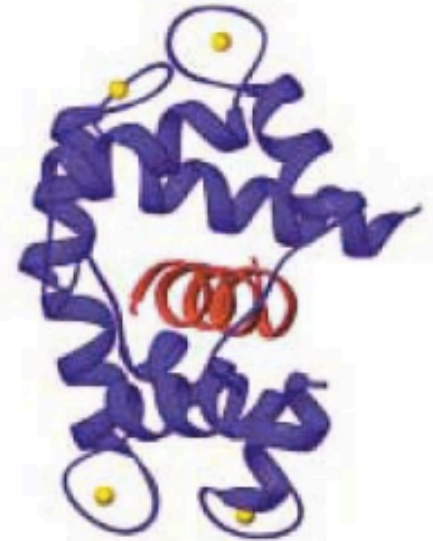


**apo-CaM**

- Apo-CaM N-terminal domain in 'closed' conformation
- Apo-CaM C-terminal domain in 'semi-open' conformation
- C-terminal domain 3-5x higher affinity for Ca<sup>2+</sup>
- Ligands coordinate Ca<sup>2+</sup> binding
- Cooperative binding of Ca<sup>2+</sup>

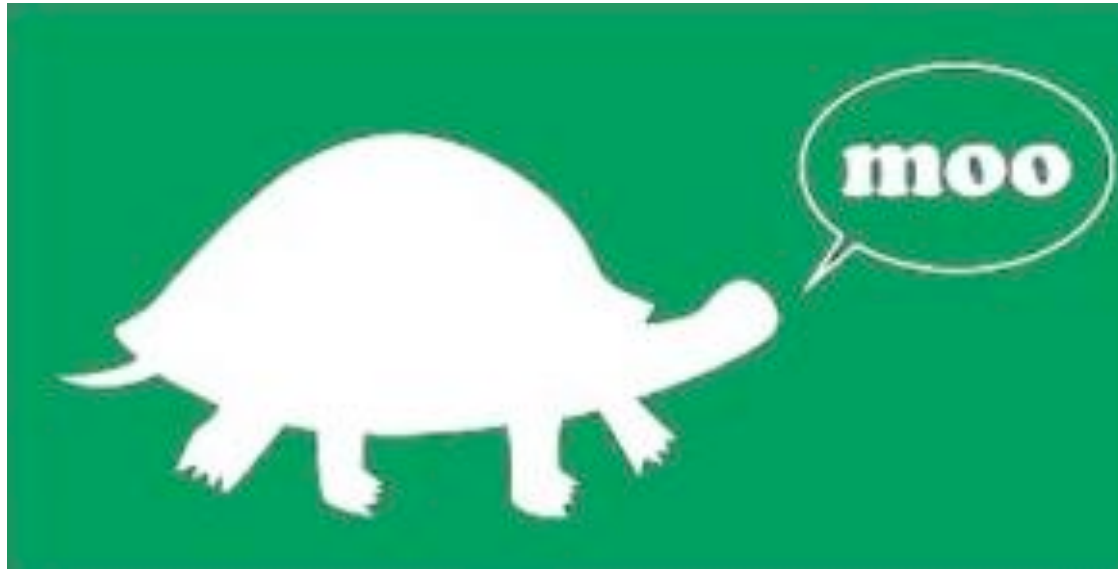
# Ca<sup>2+</sup>/CaM binds peptides

- Relieve autoinhibition
- Remodel active sites
- Promote dimerization



**Ca<sup>2+</sup>/CaM-target**

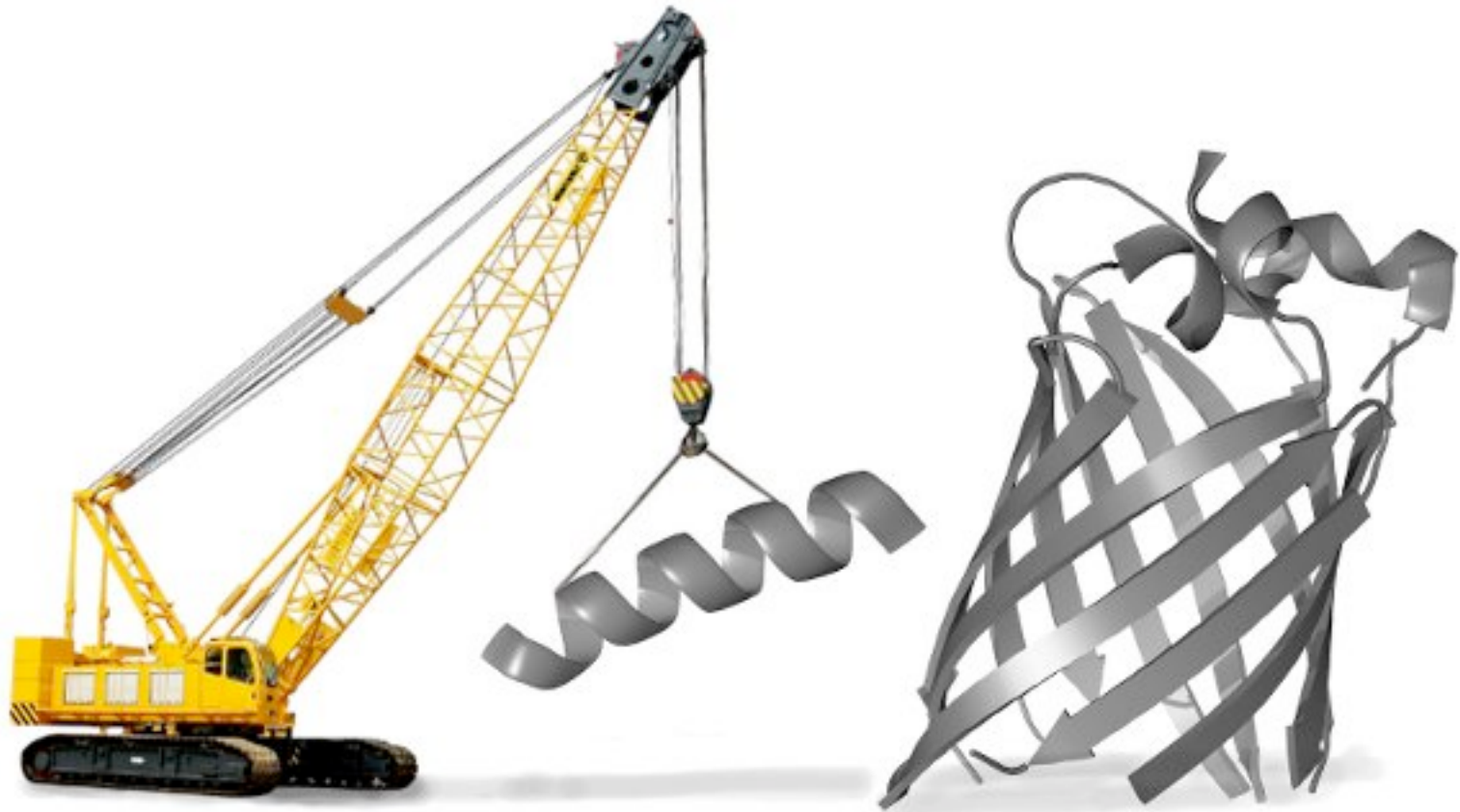
So what. Now what?



We are biological *engineers!*



# Protein engineering

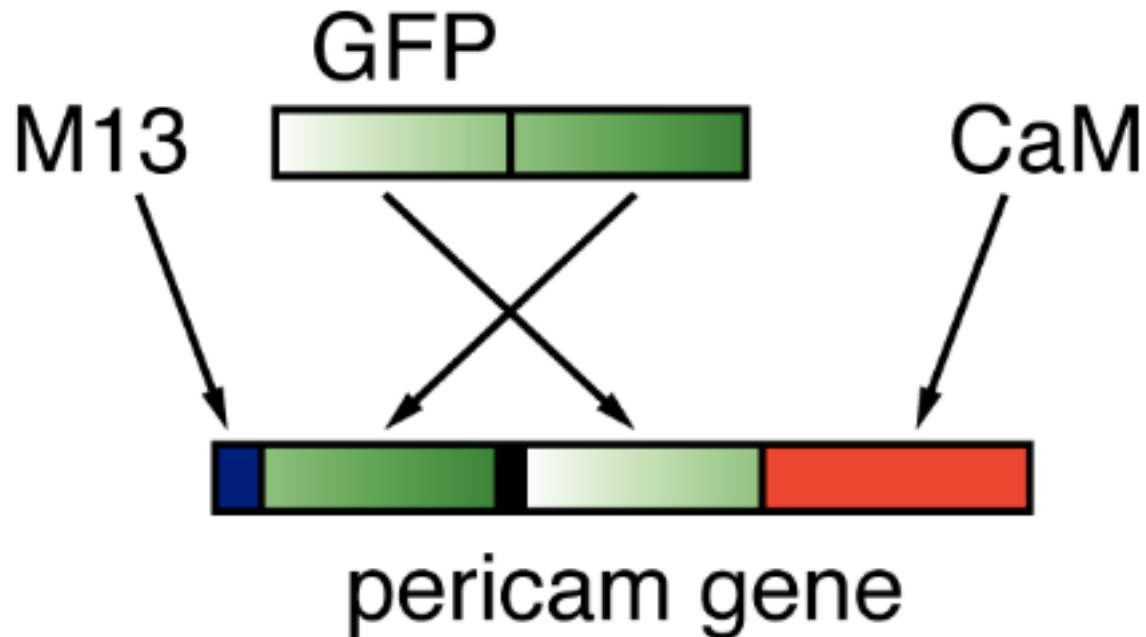


# Mechanisms for engineering proteins

- Rational design
- Random library screens
- Directed evolution

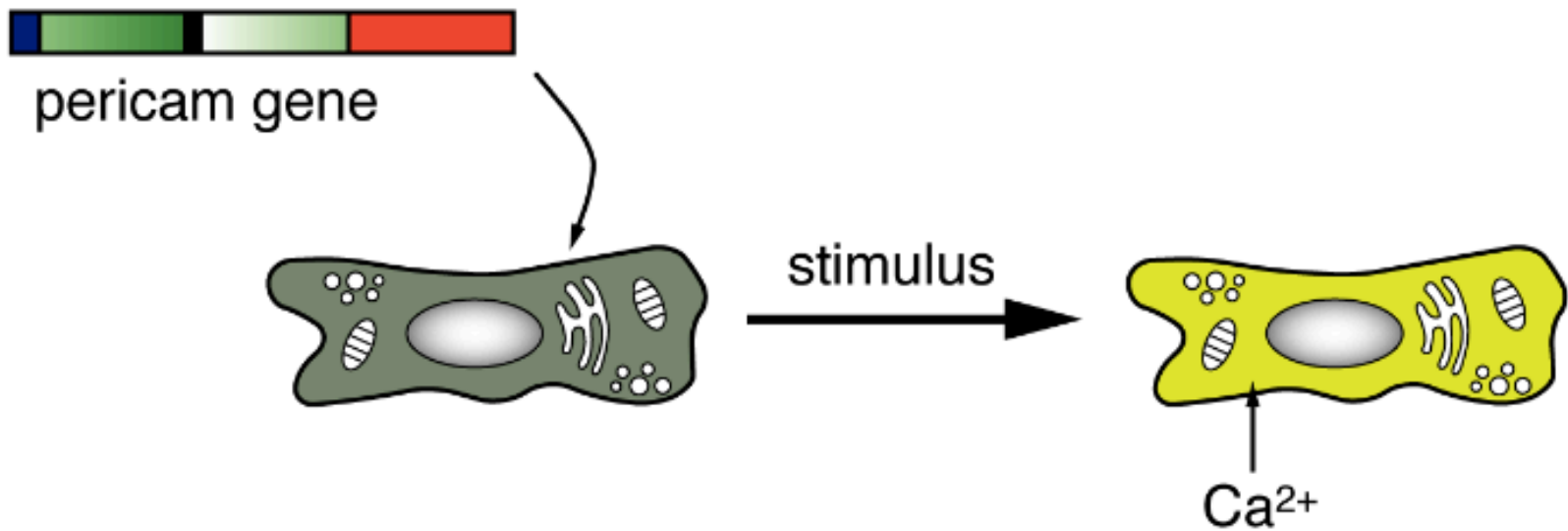
What if you engineered a  $\text{Ca}^{2+}$  sensor?

# Pericam design strategy



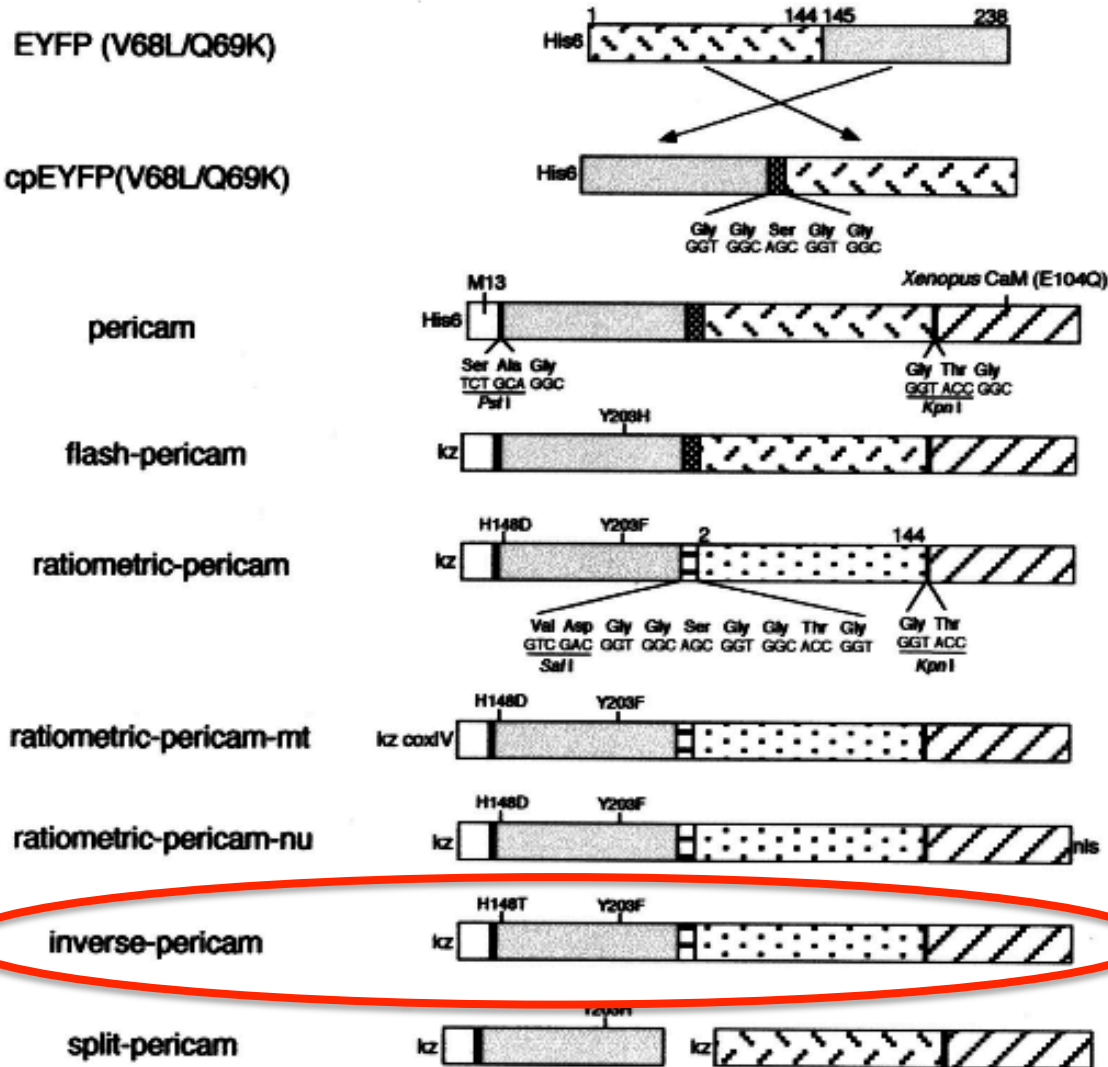
- GFP = green fluorescence protein
- CaM = calmodulin
- M13 = CaM binding peptide of cellular kinase

# Pericam protein monitors free $\text{Ca}^{2+}$

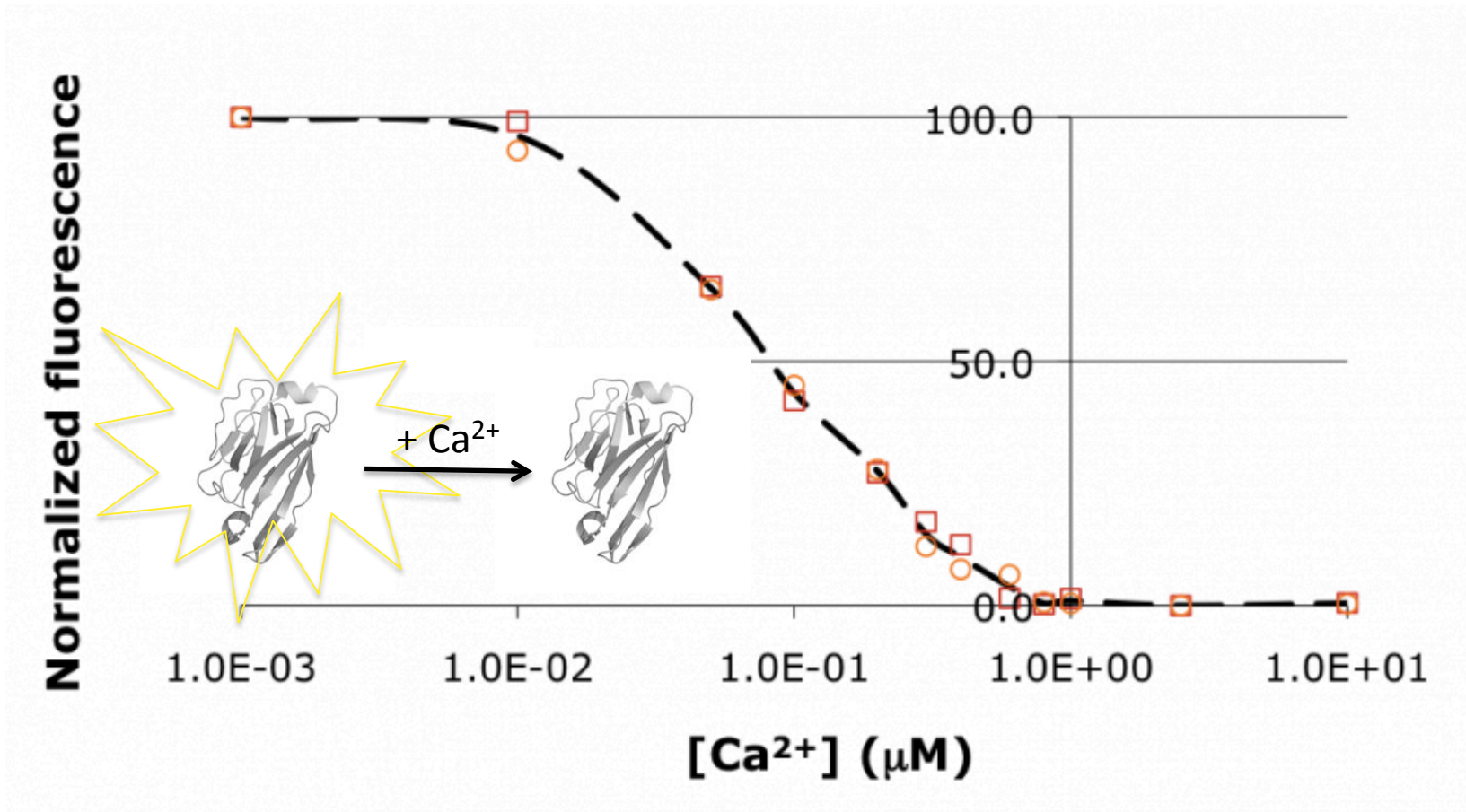


Protein-based machine for measuring  $[\text{Ca}^{2+}]$

# Suite of pericam constructs

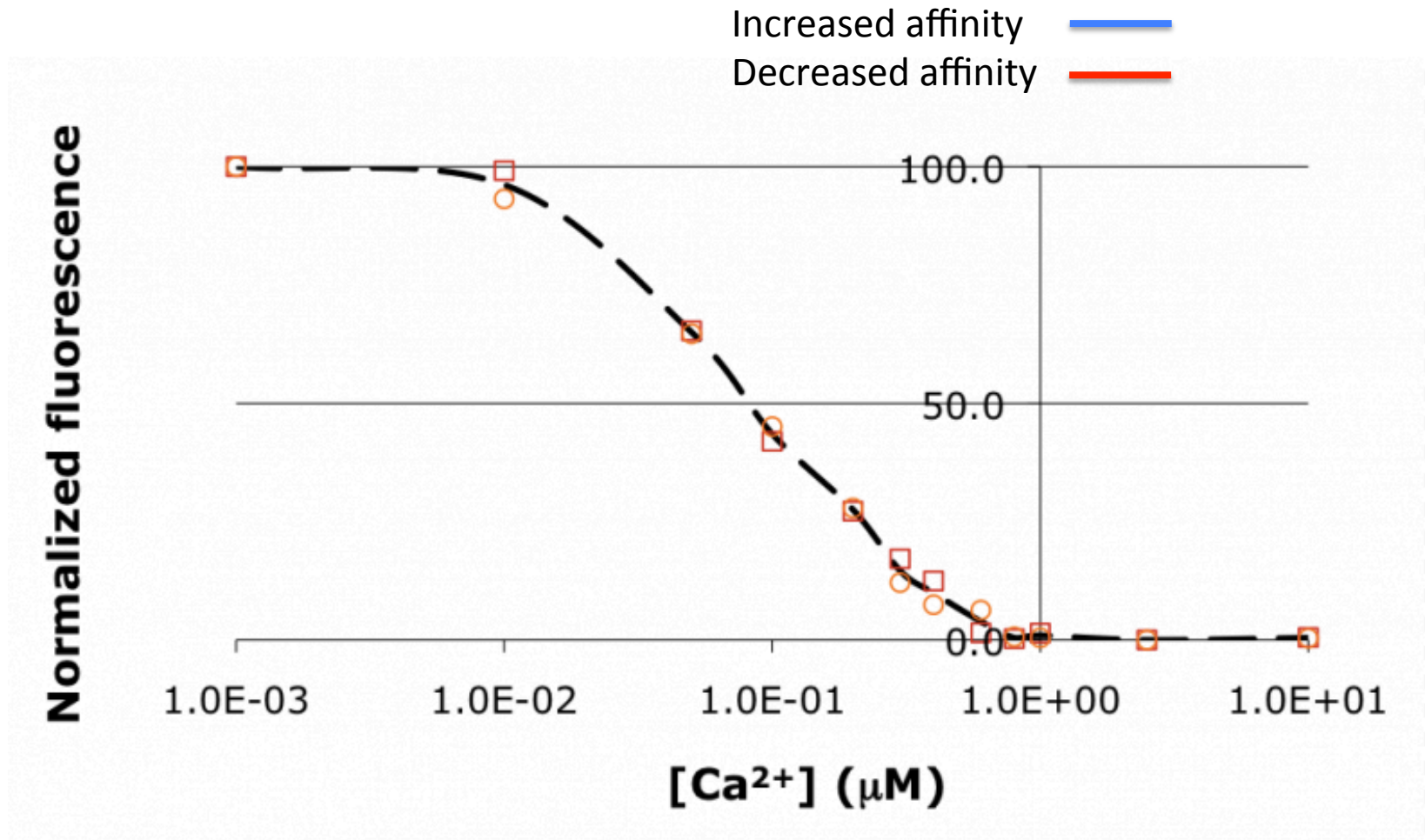


# Inverse pericam (IPC) dims with $\text{Ca}^{2+}$



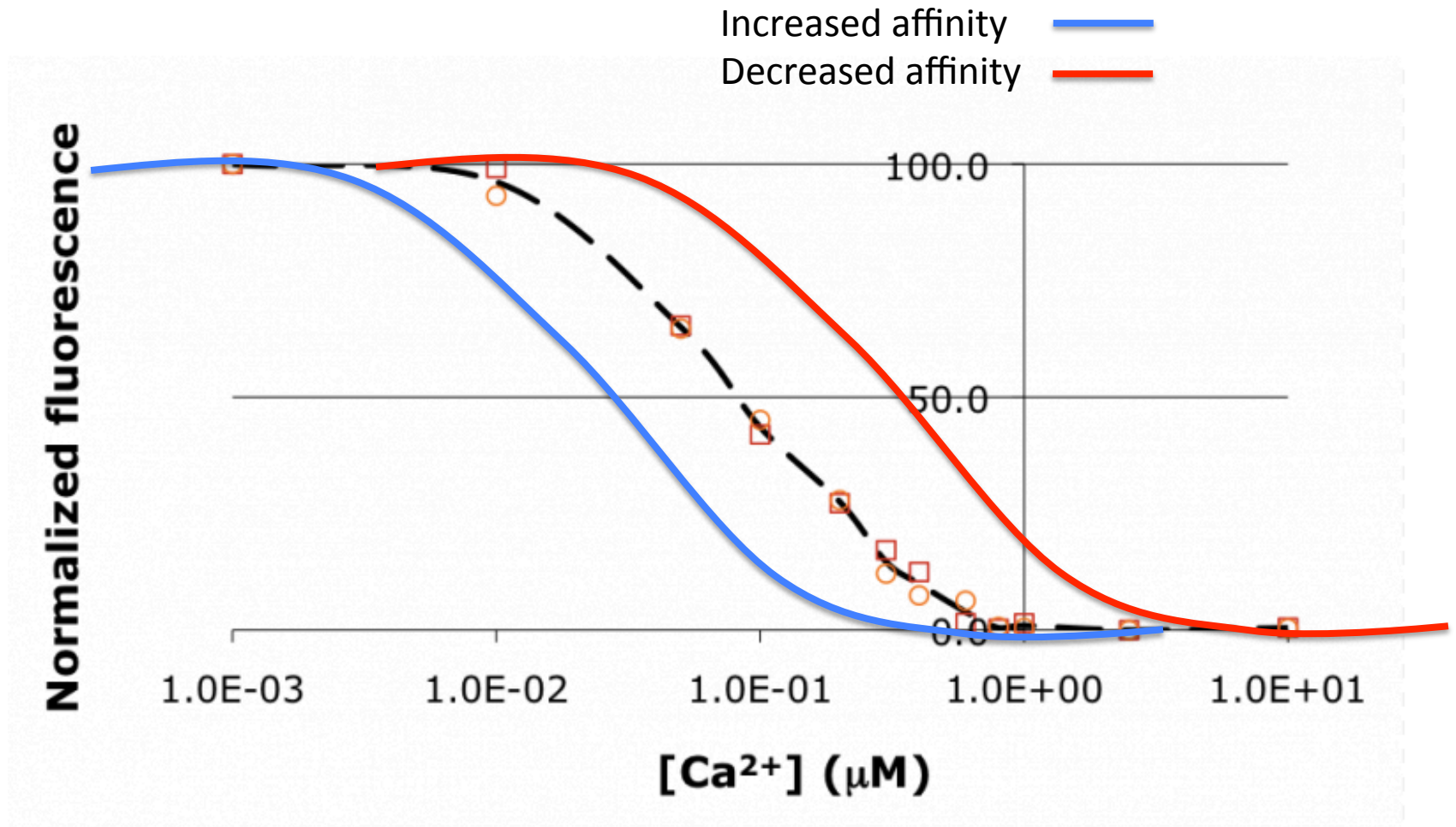
What parameters are we changing?

# How do our mutations alter binding?

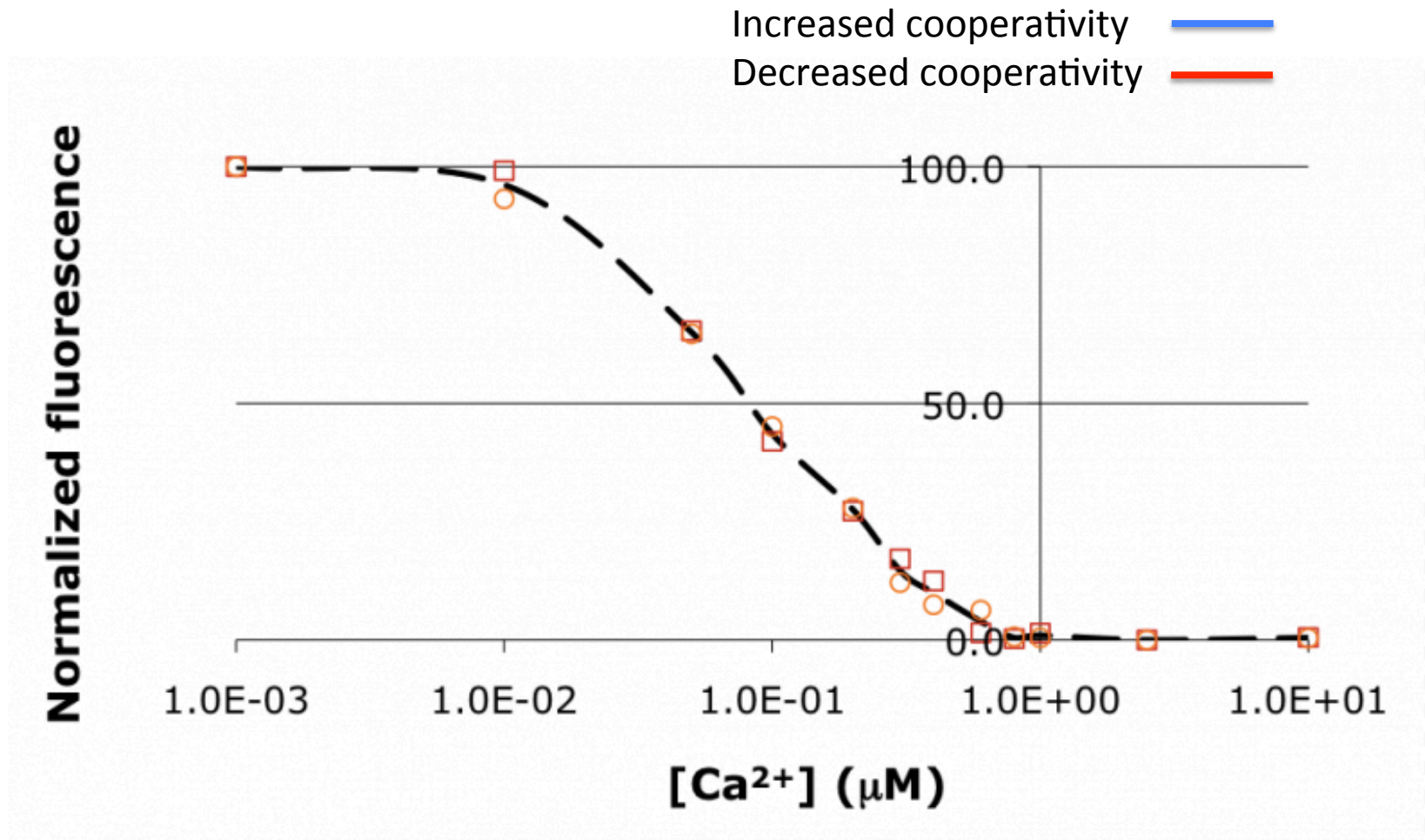




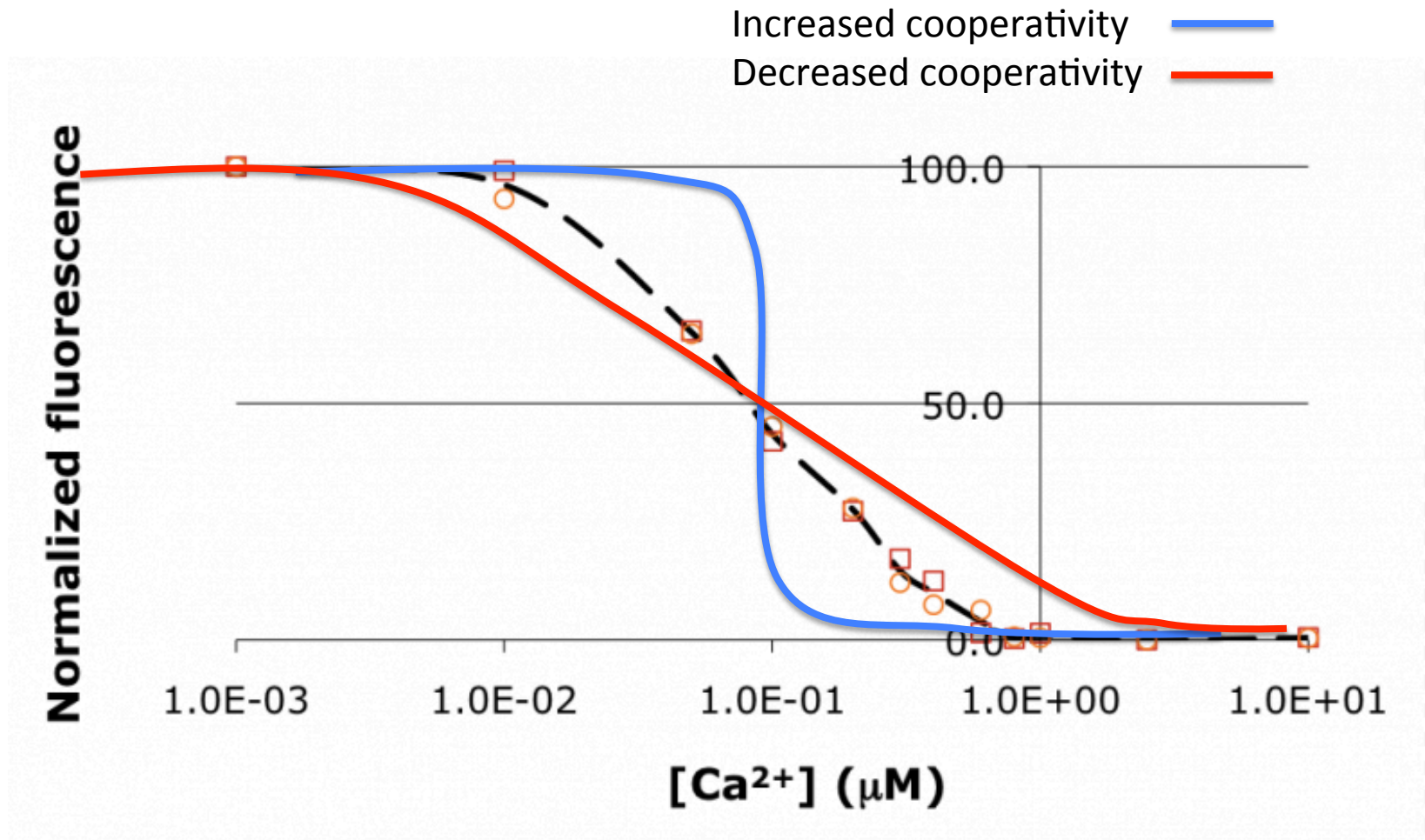
# How do our mutations alter binding?



# How do our mutations alter binding?

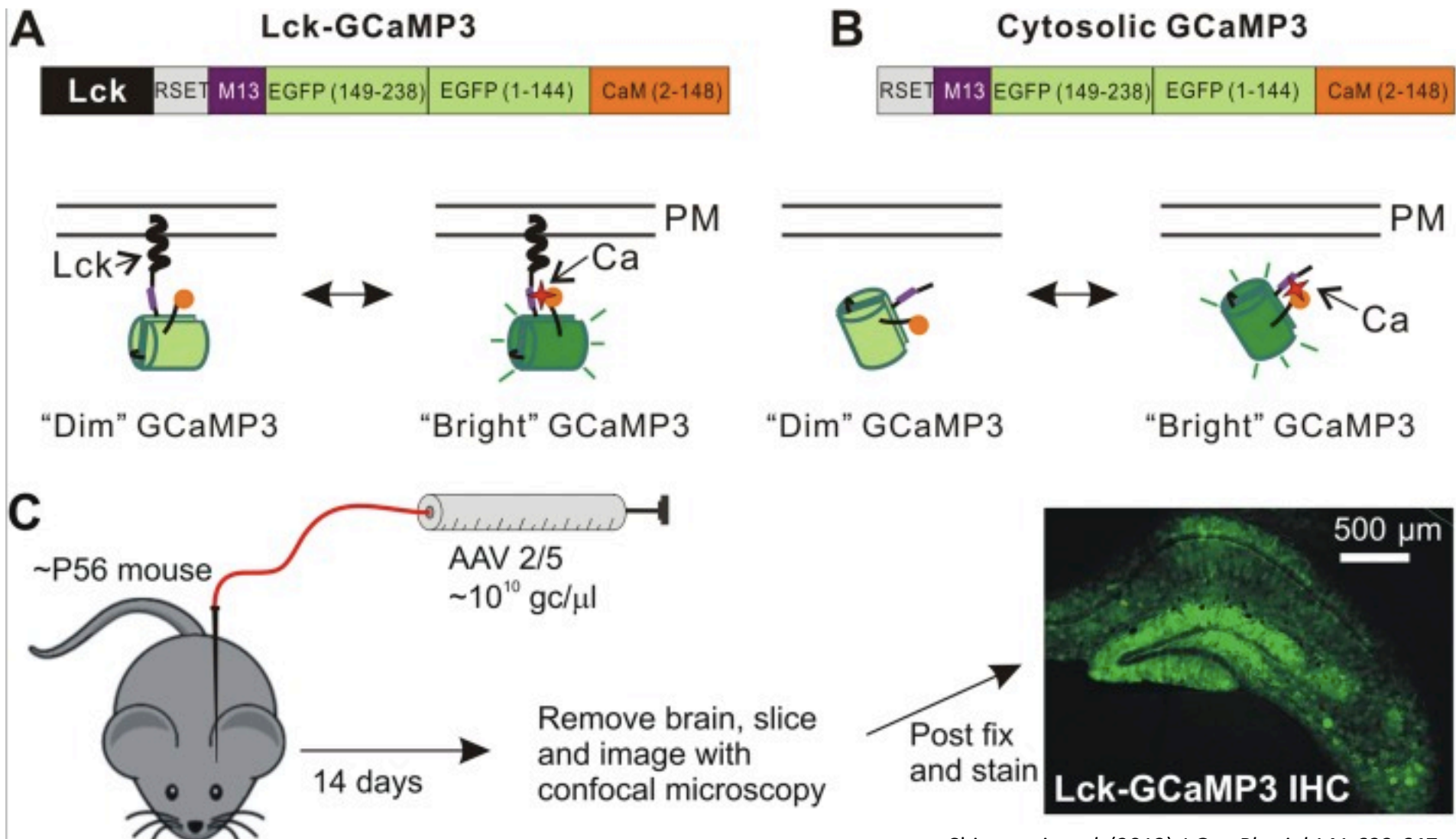


# How do our mutations alter binding?



Why engineer a  $\text{Ca}^{2+}$  sensor?

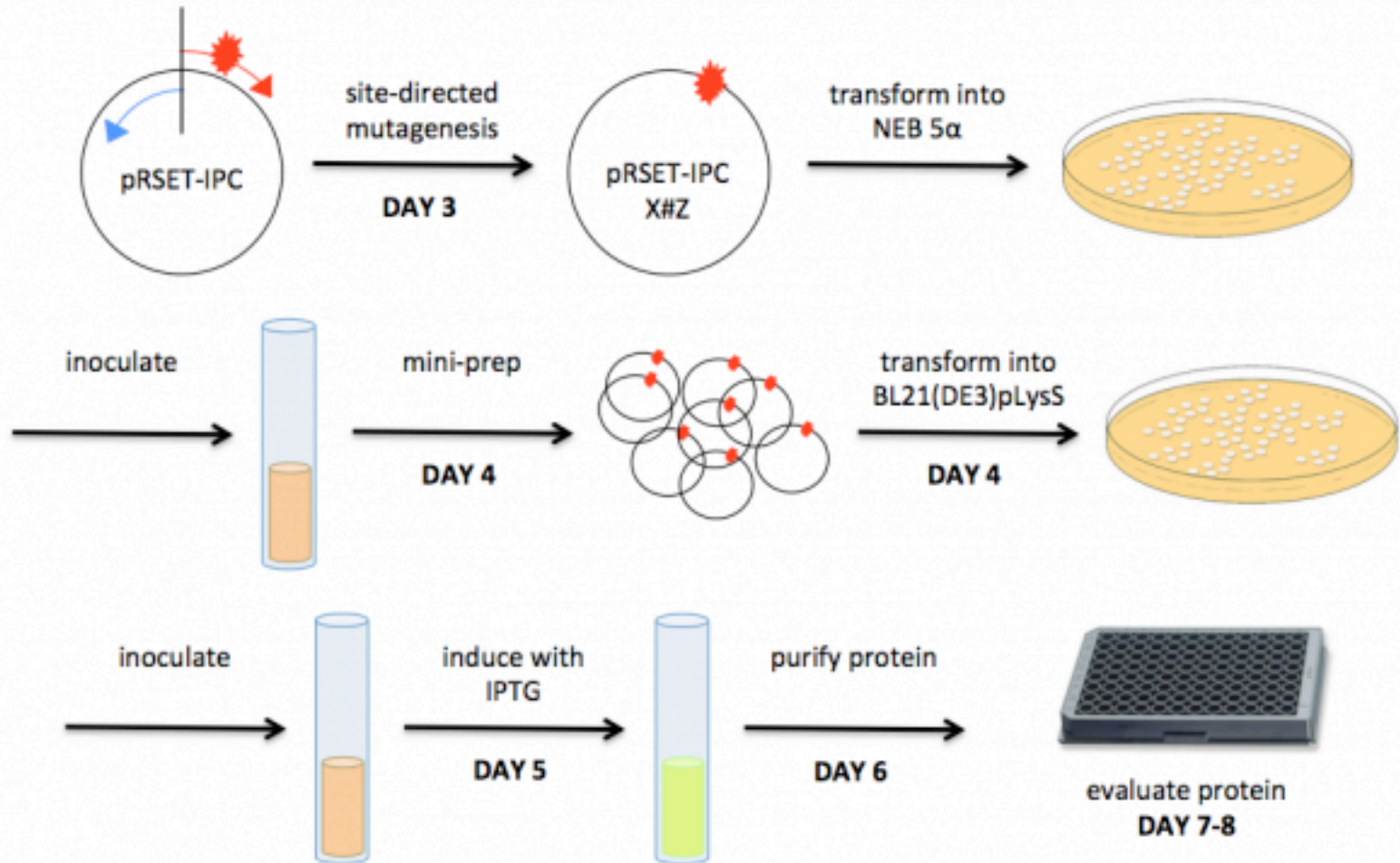
# Astrocytes propagate $\text{Ca}^{2+}$ signals to neurons and blood vessels



# Which brings us back to Mod 2...

- Aim: alter binding affinity and/or cooperativity
- Skills:
  - Use computational tools to rationalize protein engineering strategy
  - Perform mutagenesis reaction to alter protein sequence
  - Purify protein
  - Assess calcium binding behavior with titration assay

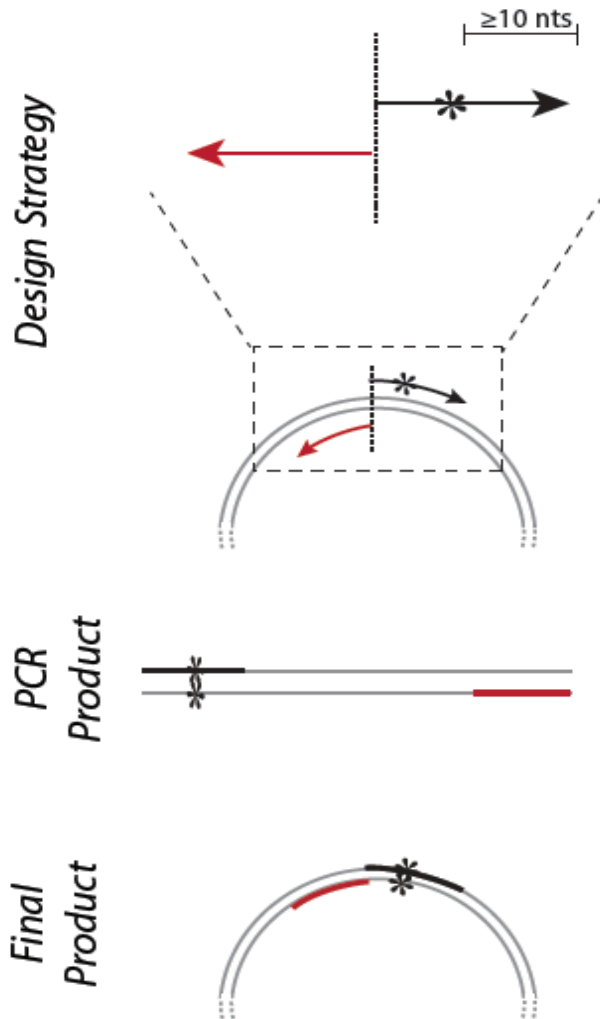
# Mod 2 overview



How do we engineer a  $\text{Ca}^{2+}$  sensor?



# Site-directed mutagenesis



- Can mutate 1-3 bp
  - 1-2 bp is ideal
- Mutation located in center of forward primer
- Forward and reverse primer anneal 'back-to-back'

# In the laboratory...

- Familiarize yourself with IPC sequence
- Identify mutagenesis site
- Design mutagenesis primers
  
- For next time...

## **Circularly permuted green fluorescent proteins engineered to sense $\text{Ca}^{2+}$**

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Communicated by Roger Y. Tsien, University of California, San Diego, CA, December 30, 2000 (received for review December 11, 2000)