

- Announcements, Review HW
- Lab Quiz
- Pre-lab Lecture
 - ❖ DNA Extraction (Miniprep)
 - ❖ Diagnostic Gel Review
 - ❖ Safety + Technical Tips

Announcements

- Your progress report is due in one week
 - OH (room TBA) on: Sun 4-5:30pm
Natalie Sat. 3-5pm (Tue 3-4)
- Please post your colony counts in *talk* page table before leaving – we'll discuss them next time
- read NCB paper for next time
- * use 20109.talk@gmail.com

Previous HW comments

- Analyzing error in linearized gel analysis
 - Error measurement? % error, not absolute difference
 - Sources of error? human measurement → band thickness
gel loading error
- Comments on Figures
 - Titles: Great overall! Hone in on *key* details.
 - Caption:
 - Introduce the players, use consistent naming scheme.
 - Avoid explaining minor methods details.
 - Stick to facts (e.g., expected band sizes), not interpretation.
 - Labeling: often insufficient. Label a few bands on ladder.

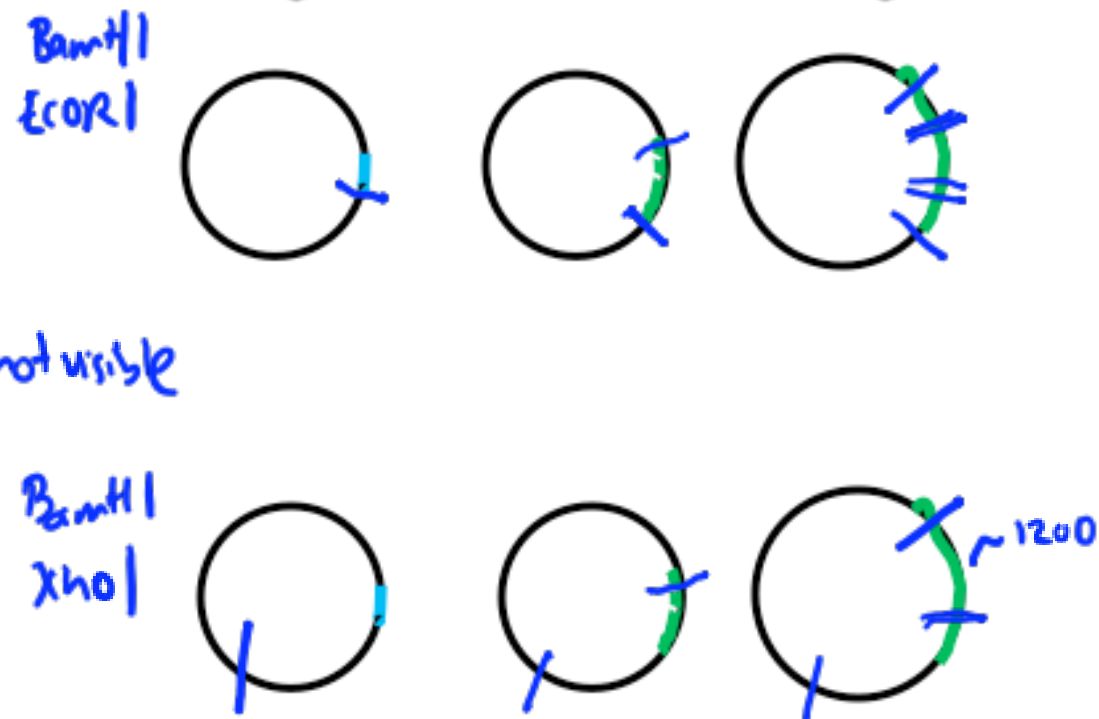
DNA Extraction from Bacteria

Step	Contains	Purpose
Soln. I	EDTA buffer, glucose	→ weakens cell envelope → otherwise stable
Soln. II	SDS Na^+ NaOH	→ disrupt, solubilize lipid membrane (+proteins) → denatures dsDNA $\text{O} \rightarrow$ ssDNA O
Soln. III	acetic acid, KAc.	neutralize pH, precipitate SDS
Transfer	N/A	$\text{O} \rightarrow \text{O}$ renature genomic DNA "crashes"
Final steps	EtOH, H ₂ O dry by	→ precipitates DNA, wash away salts, etc. → EtOH would interfere w/ digest

Diagnostic DNA Gels

kbp	bkb	bkb ins	multiple inserts
~4.6			
~0.6			
~4.6			

Choosing restriction sites for digest



* consider choose one site unique to insert, one on bkb
 * consider results of partial digestion

Today in Lab

- Miniprep three $\Delta 5$ -EGFP candidates, and bacteria transformed with pCX-NNX

control for your technique

- Count and post colony #s



- Visit from Christine Tafoya, EHS: biosafety talk
- Set up digests
– We will add loading dye if lab runs late

4pm