# **Module Overview**

Day	Lecture	Lab
1	Introduction	DNA library synthesis (PCR)
2	SELEX I: Building a Library	DNA library purification (agarose gel electrophoresis)
3	SELEX II: Selecting RNA with target functionality	RNA library synthesis
		( <i>In vitro</i> transcription = IVT)
4	SELEX III: Technical advances	RNA purification and heme
	& problem-solving	affinity selection
5	Characterizing aptamers	RNA to DNA by RT-PCR
6	Introduction to porphyrins: chemistry & biology	Post-selection IVT
		Journal Club 1
7	Aptamer applications in biology & technology	Aptamer binding assay
8	Aptamers as therapeutics	Journal Club 2

# **Aptamer Applications**

20.109 Lecture 76 March, 2012

# **Today's Objectives**

 Consider, through discussion, an increasingly important application area for aptamer technology

 Overview of antibodies as affinity reagents to provide a context for appreciating when to consider aptamers versus antibodies

# **Objective**

- As Team Leader at AptUs<sup>™</sup> Biotechnologies (77 Mass. Ave. in Cambridge), you would like to <u>develop a rapid and sensitive method for uniquely and simultaneously detecting 50 proteins</u>. These proteins have homologies ranging from 30-60% (i.e. they are highly similar to each other).
- The relative abundance of these proteins in human serum samples correlates with diabetes risk.
- You are highly motivated to use your team's primary expertise to generate aptamers that can unambiguously distinguish these proteins.
- The newest team member, *Hot Shot*©, asks about using antibodies to tackle this problem. After the massive group laughter subsides, you promise to discuss later.

- What are some key SELEX-related issues you'll need to address?
- What are your strategies for addressing these issues?
- How will you integrate your aptamers into a detection device?
  - What will be your readout?
  - How will you calibrate your system?
  - Keep in mind that you will likely want to analyze hundreds of samples simultaneously!

# **Discuss**

- What are some key SELEX-related issues to address?
  - Large number of proteins and selection optimization space to consider
  - High degree of similarity between the various targets => potential for cross-reactivity is high
  - Obtaining target proteins (all 50?!)
    - Pure?
    - Mixture?

- What is your strategy for addressing these issues?
  - Large number of proteins and selection optimization space to consider
    - Automation (options?)
  - High degree of similarity between the various targets => potential for cross-reactivity is high
    - Counter-selection (how might you efficiently do this?)
    - Perform SELEX on the isolated, non-homologous protein regions only
      - Advantages and disadvantages?
  - Obtaining pure target proteins (all 50?!)
    - Recombinantly express each individually
    - Complex mixture SELEX (problems?)

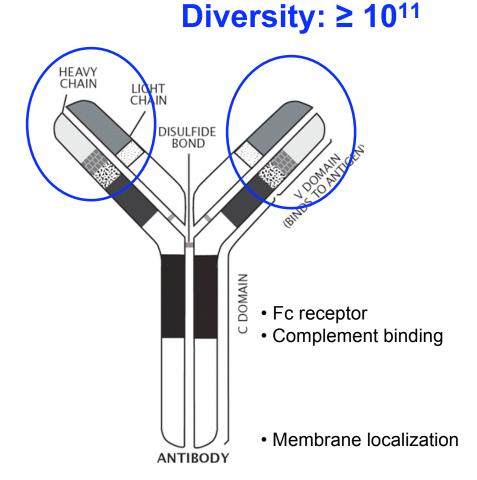
- You've successfully obtained aptamers for 45/50 targets!
  - How do you verify their respective specificities?
  - What cross-reactivities are you concerned about?
    - Between highly related classes of proteins
    - "Non-specific" interactions with other unrelated proteins
      - Especially high abundance proteins (e.g. albumin)

- You now start thinking about how you'll use these aptamers to test the levels of these various proteins in blood samples.
  - What test format do you choose?
    - Solution phase
    - Immobilized (which component do you immobilize?)
    - How do you detect/visualize a binding reaction?
  - You anticipate doing high throughput testing. Which formats are most compatible with having to run tests for 100 samples/day?
    - Microarray
    - Microfluidics

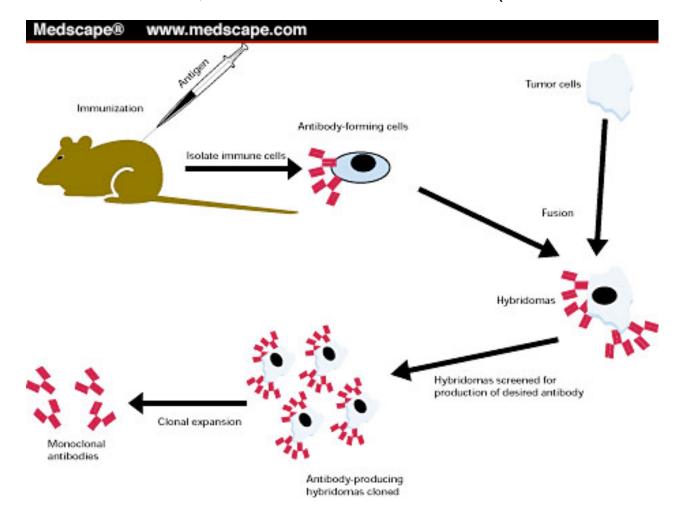
# Aptamers versus antibodies

### **Antibodies: General**

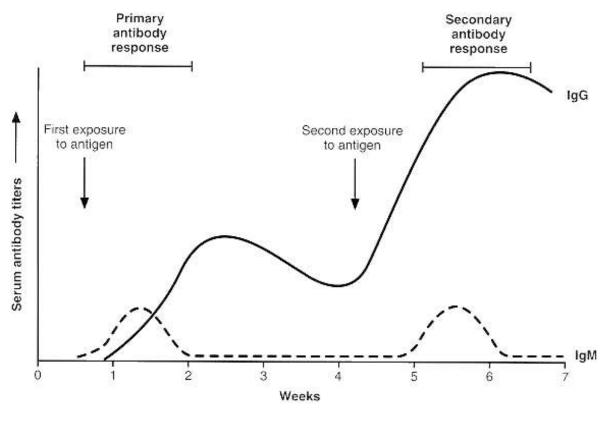
- Antibodies are proteins
  - Produced by B cells
  - Each B cell produces one type of antibody
- Overall structure
  - 2 heavy chains
  - 2 light chains
  - Both intra- and inter- chain disulfide bonds important for maintaining structure and function
- Functional Regions
  - Variable
    - Antigen binding
      - Both heavy & light chains contribute to binding site
  - Constant regions



- How are antibodies produced?
  - Immunize an animal (e.g. mouse, goat, rabbit)
  - Several months later, antibodies can be isolated (if immunization successful)



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Typical time course for antibody production after immunization

- Are there any similarities between antibody and aptamer selections?
  - An animal makes large, randomized antibody pools (B cell clones)
  - Antibodies to the target antigen are selected from this pool
- Do you expect an antibody to every possible target will be present in an animal's antibody repertoire?
  - Consider representation within space considerations
  - Any other factors?
    - Yes--Antibodies that interact strongly with self-proteins are stringently selected against
    - B-cell clones capable of making these antibodies are eliminated from an animal's antibody repertoire

- What are some advantages to using or working with antibodies for binding reactions?
  - Lots of existing expertise!
  - Stability
  - Nature has optimized the selection process
    - Robustness of antibody production process
    - Potentially higher success rate of identifying an antibody
  - Selection stringency high ==> skewed towards identifying high affinity antibodies
  - Selection occurs (unattended) in immunized animal

- What are some disadvantages to using or working with antibodies for binding reactions?
  - Limited to targets that are not toxic to the animal!
  - Cannot easily tune the selection stringency
  - No guarantee that antibody will function in non-physiological conditions
  - Limited antibody reuse
  - Requires using animals (costly to house)
  - Batch-to-batch variability can be high

# **Aptamers** versus antibodies

- Limited to targets that are not toxic to the animal!
- Aptamers to toxic substances can be developed
- Cannot easily tune selection stringency
- Stringency easily tuned
- No guarantee that antibody will function in non-physiological conditions
- Aptamers can be selected under conditions in which they will be used
- Limited antibody reuse
- Aptamers can be refolded and reused
- Requires using animals (costly to house)
- No live animals required
- Batch-to-batch variability can be high
- Chemically well-defined aptamers can be reproducibly synthesized

# Summary

- Aptamers are a viable strategy for binding and distinguishing closely related protein family members
- For multi-target, high throughput protein identification studies, there is a need to:
  - Increase the throughput for producing aptamers to distinct targets
  - Use aptamers in a format compatible with processing many samples in parallel
- Antibodies are the gold standard affinity reagents in biology/biotechnology
  - Many desirable characteristics
  - Aptamers can rival antibodies
  - Must carefully consider your applications to decide which affinity agent is more suitable