

Module 2 overview

lecture

1. Introduction to the module
2. Rational protein design
3. Fluorescence and sensors

lab

1. Start-up protein eng.
2. Site-directed mutagenesis
3. DNA amplification

SPRING BREAK

4. Protein expression

5. Purification and protein analysis
6. Binding & affinity measurements
7. High throughput engineering

4. Prepare expression system
5. Induce protein
6. Characterize expression
7. Assess protein function

Lecture 4: Protein expression & purification

- I. Why express & purify proteins?
 - A. Scientific applications
 - B. Applications in industry, *etc.*

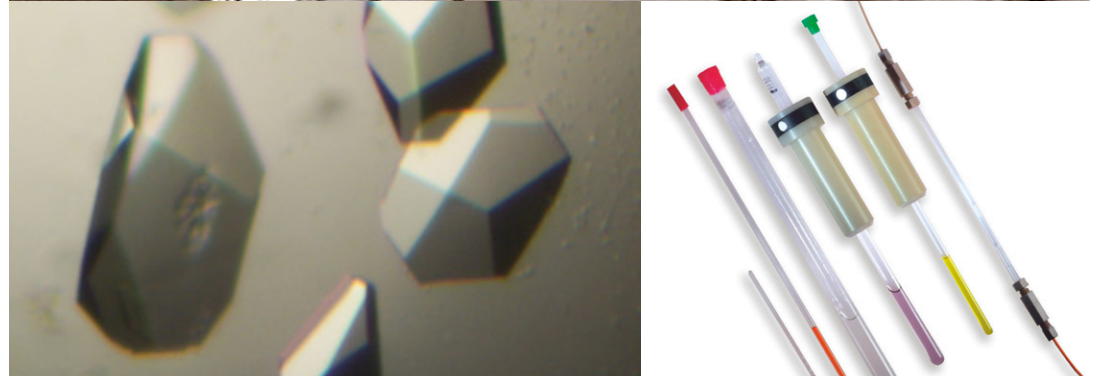
- II. Protein expression systems
 - A. Alternatives to protein expression
 - B. Prokaryotic and eukaryotic systems

Laboratory uses of purified proteins

Biochemistry analysis



Structural biology



Research biochemicals



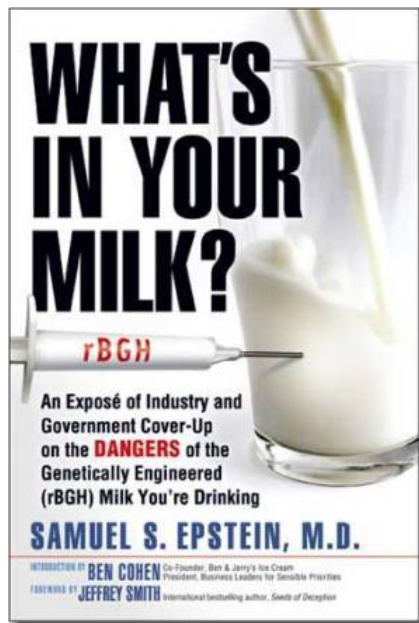
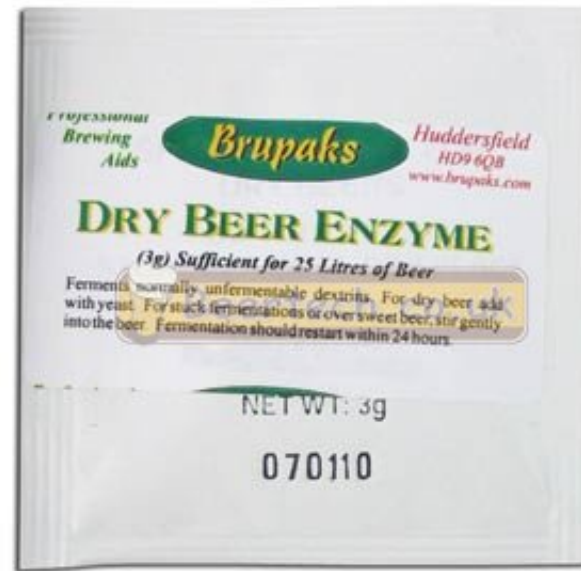
Protein therapeutics

Table 1 Top ten recombinant therapeutic proteins and their global sales between 2001 and 2003

Product (generic)/ marketing company	2001 (\$million)	2002 (\$million)	2003 (\$million)	Growth (decline) 2002– 2003 (%)
Procrit (epoetin alfa)/ Johnson & Johnson	3,430	4,269	3,986	(6.6)
Epogen (epoetin alfa)/ Amgen	2,108	2,261	2,435	7.7
Neupogen (filgrastim)/ Amgen	1,346	1,380	1,268	(8.1)
PEGylated Neulasta (pegfilgrastim)/ Amgen	0	464	1,255	170.5
Novolin (insulin systemic)/ Novo Nordisk	2,244	2,255	2,235	(0.9)
Avonex (interferon beta-1a)/ Biogen IDEC	971	1,034	1,170	13.2
PEGylated PEG-Intron A franchise (pegylated interferon alpha)/ Schering Plough	1,447	2,736	1,851	(32.3)
TNF ligand binding domain + Fc antibody domain Enbrel (etanercept)/ Amgen	856	521	1,300	149.5
epo engineered to have additional glycosylation sites Aranesp (darbepoetin alfa)/ Amgen	42	416	1,544	271.2
NeoRecormon (epoetin-beta)/ Roche	479	766	1,318	72.1
<i>Top ten product sales</i>	<i>12,923</i>	<i>16,102</i>	<i>18,362</i>	<i>14.0</i>
<i>Others</i>	<i>8,547</i>	<i>10,833</i>	<i>13,703</i>	<i>26.5</i>
<i>Total market value</i>	<i>21,470</i>	<i>26,935</i>	<i>32,065</i>	<i>19.0</i>

Source: Datamonitor and company-reported information.

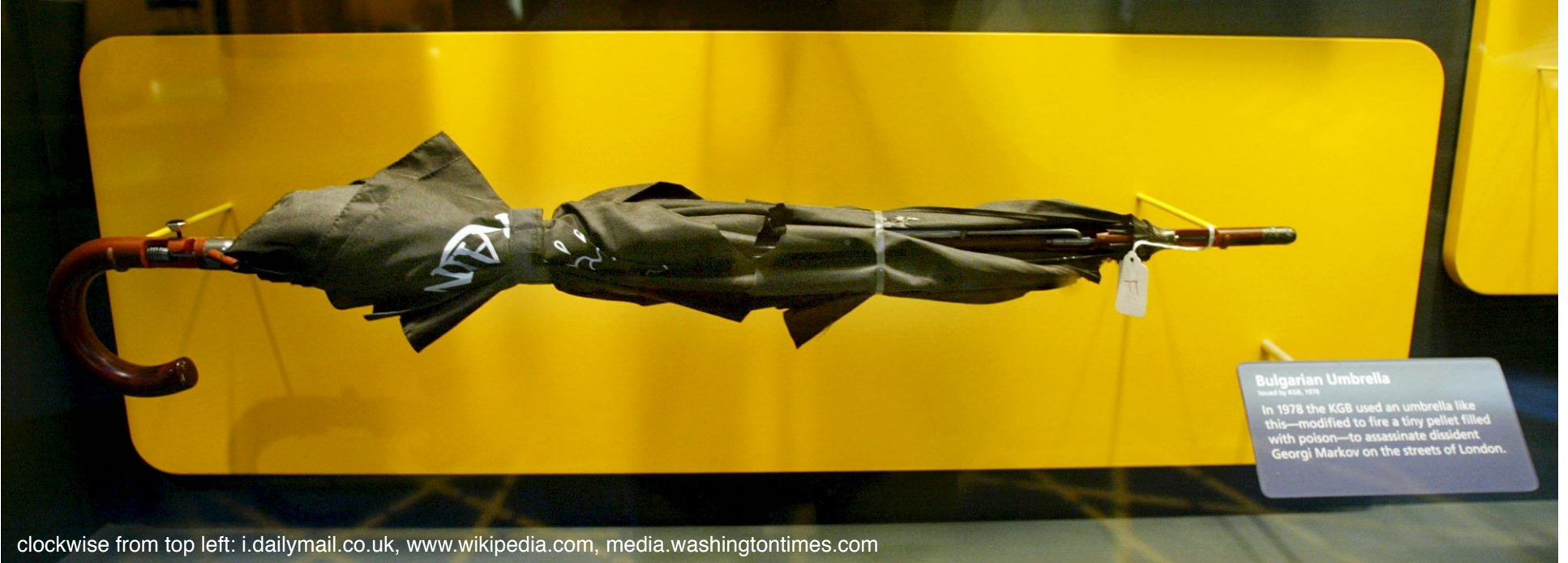
Pavlou & Reichert (2004)
Nat. Biotechnol.



clockwise from top left: s.sears.com, www.beertech.co.uk, www.treatment-skin.com, www.valleynaturals.com, servekrishna.net



<http://www.youtube.com/watch?v=bHWuj5WPvtU&feature=related>



How can proteins be produced?

1. Purify from natural source

advantages: no chemistry or DNA manipulation required, proteins likely to fold properly, assemble with native cofactors, *etc.*

disadvantages: usually only practical for high abundance proteins, source-specific purification method required

2. Synthesize *de novo*

advantages: no DNA manipulation required, synthesis methods well established, proteins produced are relatively pure

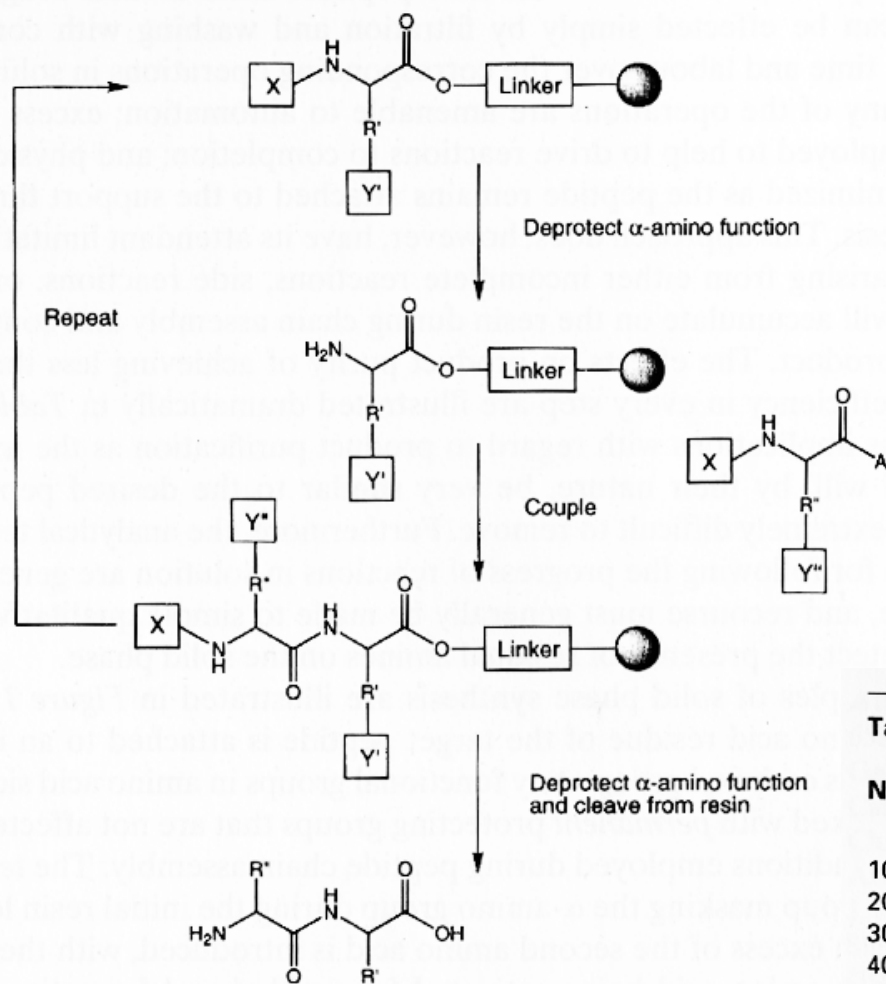
disadvantages: relatively expensive, becomes extremely difficult for polypeptides > 50 amino acids

3. Express and purify from a dedicated expression system

advantages: cheap and frequently high-yield, versatile expression systems already established

disadvantages: cloning required, troubleshooting often needed to obtain high expression and proper folding

Solid phase peptide synthesis is a reliable technique for generating short polypeptides



X = Temporary amino protecting group
 Y = Permanent side-chain protecting group
 A = Carboxy activating group

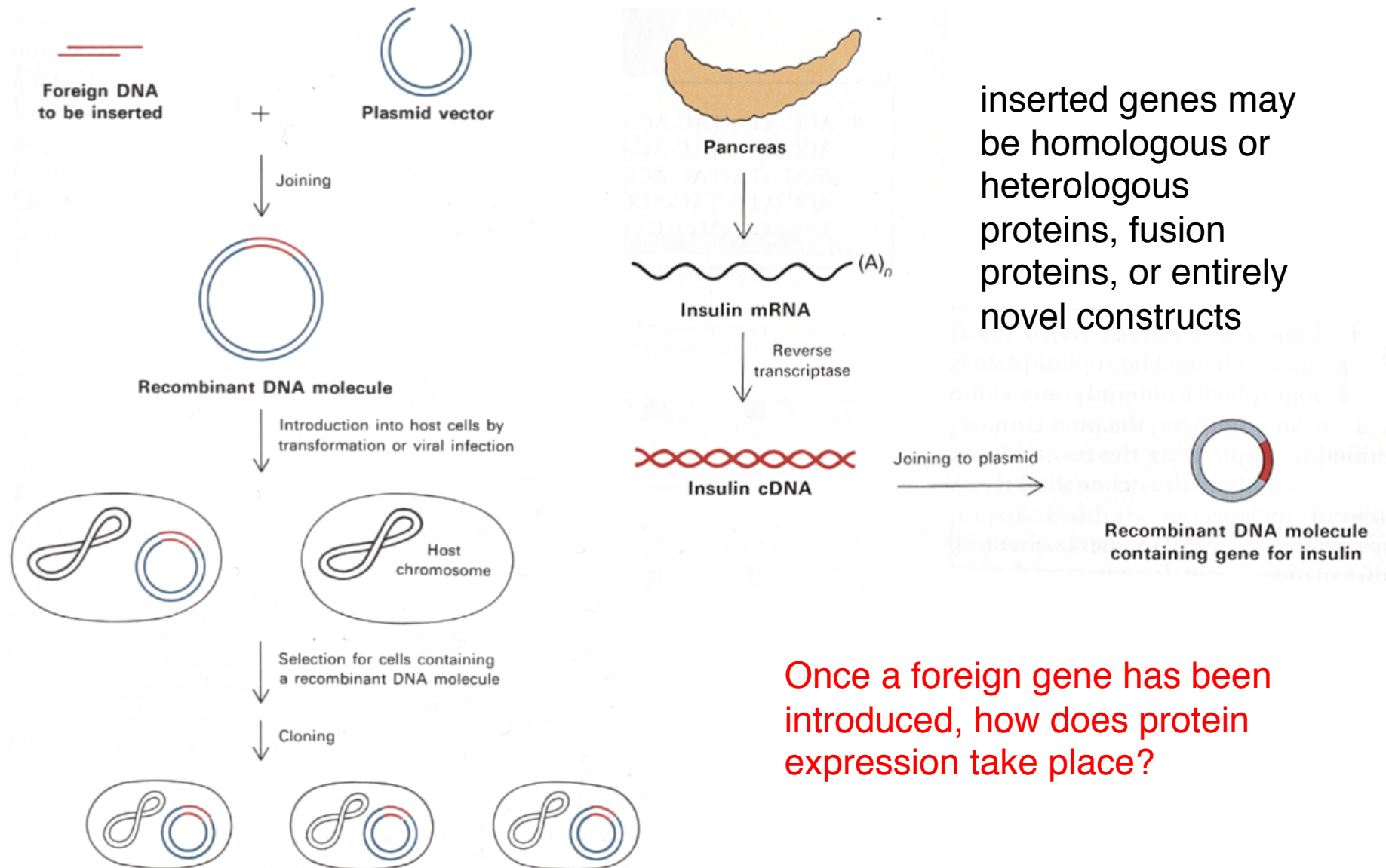


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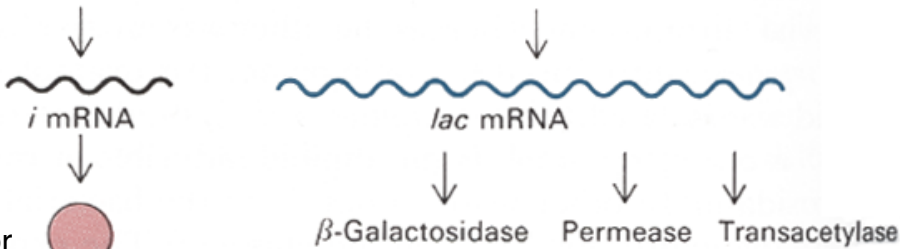
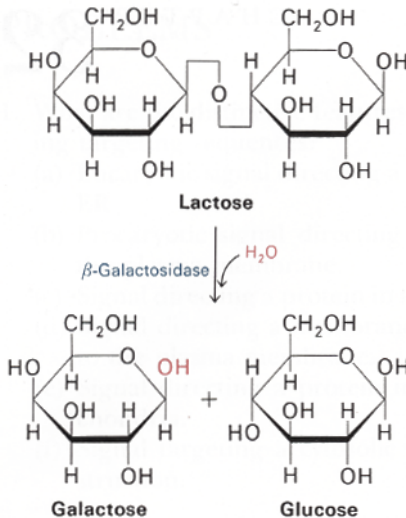
Table 1. Effects of accumulated errors on final product yields

No. of reactions	Overall yields	Yield of each reaction (%)			
		100	99	95	90
10		100	90	60	35
20		100	81	36	12
30		100	74	21	4
40		100	67	13	1
50		100	61	8	< 1

E. coli are the most common host for recombinant gene expression

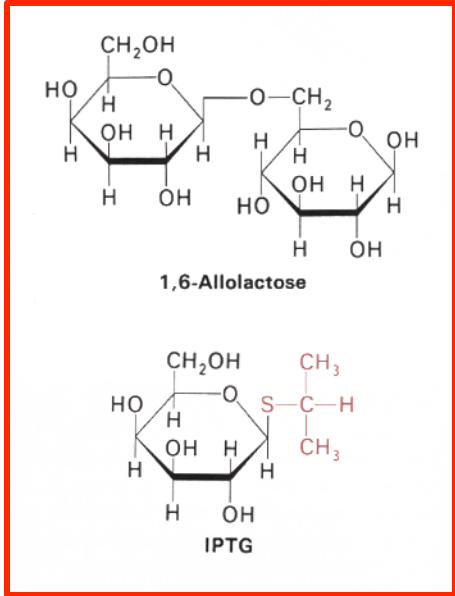


The *lac operon* is the basis for the most common bacterial protein expression systems; allows bacterial growth to be dissociated from protein expression

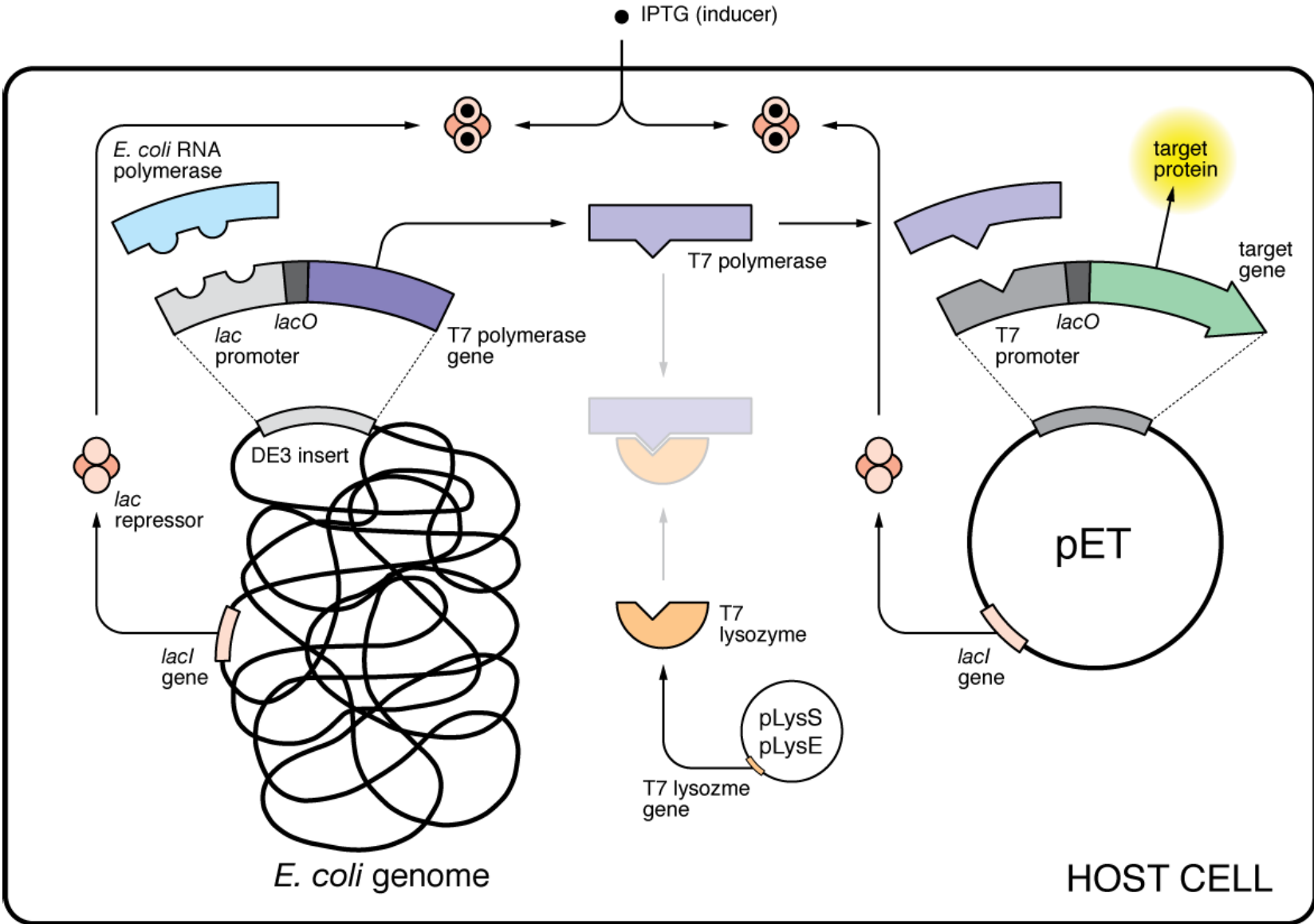


lac repressor

inducer turns on expression



T7 expression system

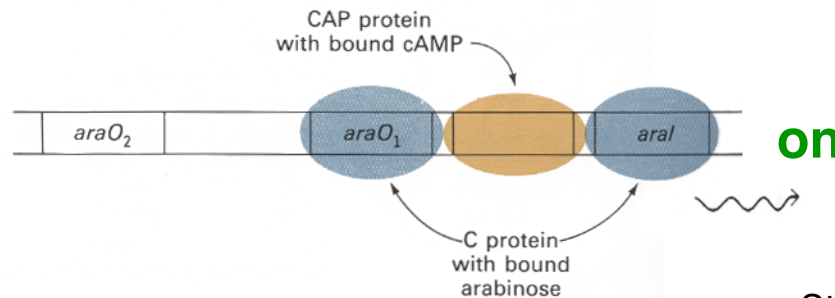
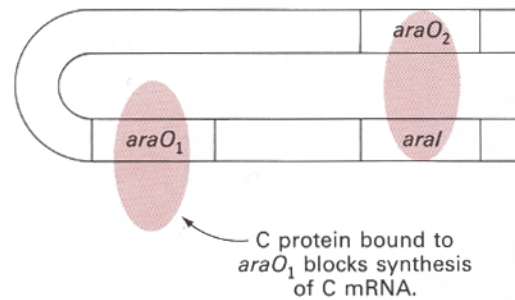
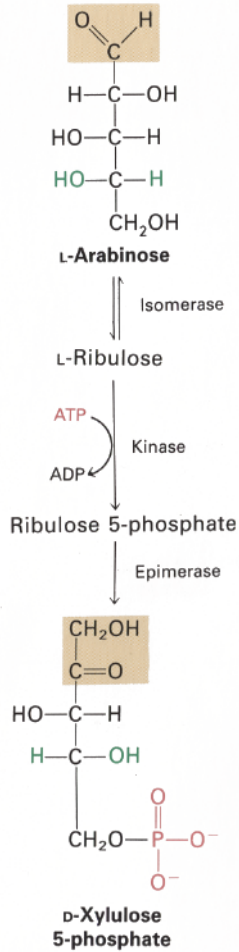
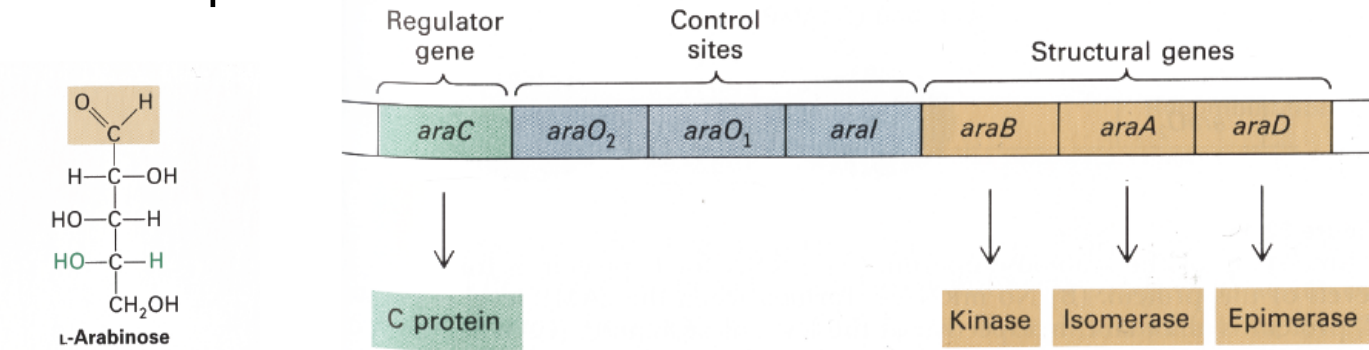




http://farm8.staticflickr.com/7024/6464583867_16c8ccdc9a_z.jpg

<http://www.biomm.com/index.php?p=3,2>

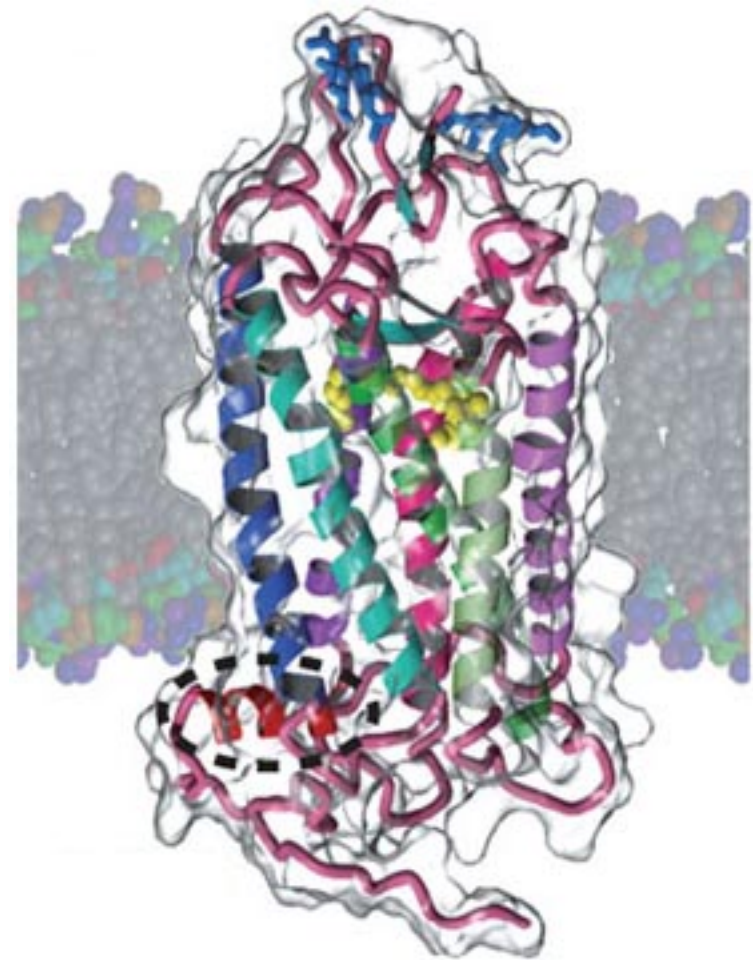
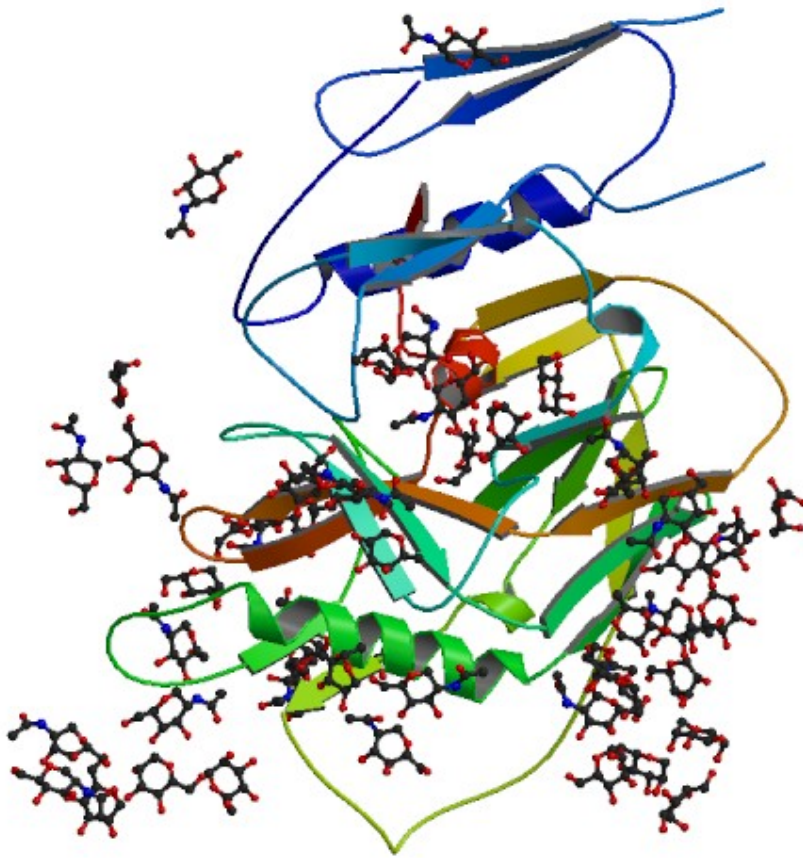
Other induction systems can also be used for protein expression in *E. coli*: arabinose system is considered to be more tightly controlled than the *lac* operon



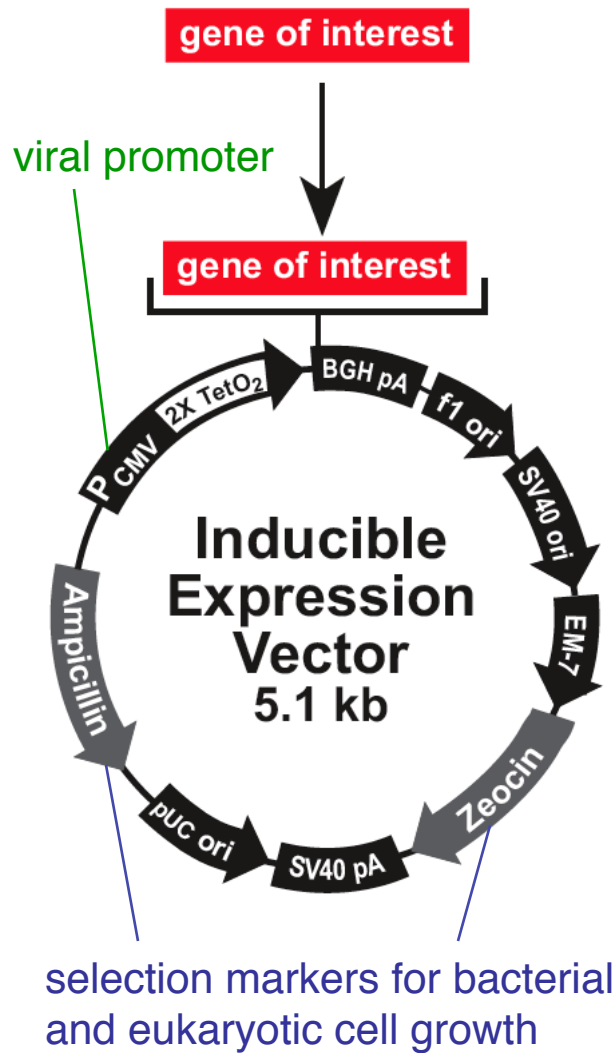
ara system is also compatible with T7-based vectors

Differences between **prokaryotic vs. eukaryotic** proteins sometimes require eukaryotic expression systems.

These two proteins exemplify characteristics that frequently call for eukaryotic expression:



Eukaryotic expression vectors share features with bacterial systems

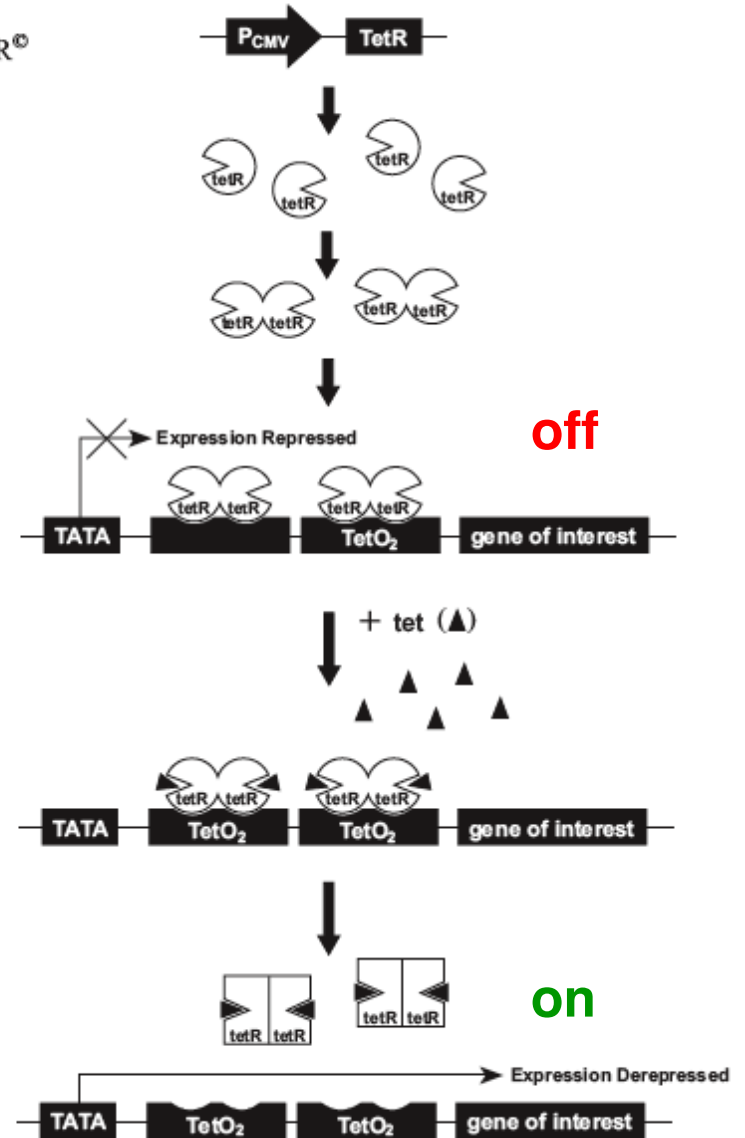


1. Tet repressor (tetR) protein is expressed from pcDNA6/TR[®] in cultured cells.

2. TetR homodimers bind to Tet operator 2 (TetO₂) sequences in the inducible expression vector, repressing transcription of the gene of interest.

3. Upon addition, tetracycline (tet) binds to tetR homodimers.

4. Binding of tet to tetR homodimers causes a conformational change in tetR, release from the Tet operator sequences, and induction of transcription from the gene of interest.



Invitrogen (2006) *T-REx System*

Prokaryotic vs. eukaryotic protein expression

<i>property</i>	<i>prokaryotic</i>	<i>higher eukaryotic</i>
yield/(L culture)	1-100 mg	widely variable
cost/(L medium)	~\$5	~\$50
introduction of DNA	transformation of competent cells	viral or nonviral transfection
handling	sterile needles, <i>etc.</i>	tissue culture hood
cell incubation	shaking incubator	usu. w/CO ₂ -control
induction	usually IPTG	none, tetracycline
glycosylation, <i>etc.</i>	no	yes
<i>notes</i>	best for small, globular proteins	best for complex, eukaryotic proteins